

# Heterozygous neuregulin 1 mice are more sensitive to the behavioural effects of $\Delta^9$ -tetrahydrocannabinol

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## Abstract

**Rationale** Cannabis use may precipitate schizophrenia especially if the individual has a genetic vulnerability to this mental disorder. Human and animal research indicates that neuregulin 1 (*Nrg1*) is a susceptibility gene for schizophrenia.

**Objectives** The aim of this study was to investigate whether dysfunction in the *Nrg1* gene modulates the behavioural effects of  $\Delta^9$ -tetrahydrocannabinol (THC), the major psychotropic component of cannabis.

**Materials and methods** Heterozygous *Nrg1* transmembrane-domain knockout mice (*Nrg1* HET) were treated

with acute THC (0, 5 or 10 mg/kg i.p.) 30 min before being tested using open field (OF), hole board (HB), light-dark (LD), elevated plus maze (EPM), social interaction (SI) and prepulse inhibition (PPI) tests.

**Results** *Nrg1* HET mice showed differences in baseline behaviour with regard to locomotor activity, exploration and anxiety. More importantly, they were more sensitive to the locomotor suppressant actions of THC compared to wild type-like (WT) mice. In addition, *Nrg1* HET mice expressed a greater THC-induced enhancement in % PPI than WT mice. The effects of THC on anxiety-related behaviour were task-dependent, with *Nrg1* HET mice being more susceptible than WT mice to the anxiogenic effects of THC in LD, but not in the EPM, SI and OF tests.

**Conclusions** *Nrg1* HET mice were more sensitive to the acute effects of THC in an array of different behaviours including those that model symptoms of schizophrenia. It appears that variation in the schizophrenia-related neuregulin 1 gene alters the sensitivity to the behavioural effects of cannabinoids.

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## Introduction

Part of a clinical diagnosis of schizophrenia is the characterisation of this mental disorder by a range of psychiatric symptoms frequently classified into positive (e.g., hallucinations, delusions), negative (e.g., impaired emotion, social withdrawal) and cognitive symptoms (e.g., impairment of attention, language, memory) (Wong and Van Tol 2003). These symptoms are at least partially based on dopamine

(DA) hyperstimulation and a dysfunction of either glutamate or its *N*-methyl-D-aspartate (NMDA) receptor subtype (Coyle 1996; Wong and Van Tol 2003). According to the neurodevelopmental theory of schizophrenia, such altered neurochemistry is thought to arise due to an interaction between genetic and environmental factors promoting defects of brain circuitry maturation (Duncan et al. 1999; Farber et al. 1995). This results in abnormalities of synaptic connectivity, myelination or structural changes in the cortex of schizophrenia patients.

Genetic factors play an important role in the aetiology of schizophrenia (Cardno and Gottesman 2000; Sullivan et al. 2003). A meta-analysis of genome scans has reported a schizophrenia susceptibility locus on chromosomes 8p12–p23.1. Furthermore, significant associations with a candidate gene on 8p12, neuregulin 1 (*NRG1*), have been reported in Caucasian (Stefansson et al. 2002) and Asian populations (Tang et al. 2004; Yang et al. 2003), although some studies failed to replicate this association (Duan et al. 2005; Iwata et al. 2004). Nevertheless, *Nrg1* with its more than 15 isoforms (Falls 2003), has an impact on schizophrenia-related brain processes such as the expression and function of central nervous system (CNS) neurotransmitter receptors for NMDA and gamma-aminobutyric acid (GABA), the activation of glial cells and the regulation of myelin and oligodendrocyte-related gene expression. Furthermore, altered *NRG1* mRNA expression can be found in postmortem dorsolateral prefrontal cortex and hippocampus of schizophrenia patients (Hashimoto et al. 2004; Law et al. 2006).

Several studies have proposed a schizophrenia-related phenotype for transgenic mice heterozygous for transmembrane *Nrg1* (Falls 2003): the phenotype includes hyperactivity (reversible with clozapine), deficits in prepulse inhibition and habituation processes and fewer functional NMDA receptors (O'Tuathaigh et al. 2006; Stefansson et al. 2002). Importantly, transmembrane *NRG1* was recently found to associate with schizophrenia (Walss-Bass et al. 2006). Thus, the heterozygous *Nrg1* transmembrane-domain knockout mouse may provide a putative animal model of genetic vulnerability to schizophrenia.

The use of cannabis increases the risk of developing schizophrenia (Linszen et al. 1994), and people with psychotic disorders are more likely to use or have used psychoactive drugs than other psychiatric patients or the general population (Arseneault et al. 2004; Schneier and Siris 1987). It has been hypothesised that chronic (Degenhardt and Hall 2002; Leweke et al. 2004; Linszen et al. 1994) and less robustly acute (Favrat et al. 2005) cannabis consumption has an impact on the aetiology of schizophrenia in subjects with a predisposition for this mental disorder. Importantly, not only cannabis consumption during puberty/adolescence but also a variety of other external/environmental factors such as obstetric complications (Boksa 2004) or nutritional deficiency

(Susser and Lin 1992) may unmask schizophrenia in individuals who have a prior vulnerability to the disorder (Degenhardt and Hall 2002; Leweke et al. 2004). This is consistent with the two-hit hypothesis of schizophrenia, which poses that psychotic patients harbour one or various susceptibility genes that are necessary but not sufficient to cause schizophrenia (hit 1). Environmental stressors, such as cannabis abuse, may then help to trigger the onset of schizophrenia by interacting with this genetic vulnerability to the mental disorder (hit 2) (Bayer et al. 1999).

The availability of transgenic mice offers a unique opportunity to systematically investigate such gene–environment interactions, while avoiding the many confounding factors that plague human studies on cannabis-induced psychosis (Henquet et al. 2005). In this study, we examine, whether dysfunction in the *Nrg1* gene modulates the behavioural effects of  $\Delta^9$ -tetrahydrocannabinol (THC), the major psychotropic component of cannabis, using heterozygous *Nrg1* transmembrane domain knockout (*Nrg1* HET) mice. The effects of different dosages of acutely administered THC on wild type-like and *Nrg1* HET mice were assessed using a multitiered phenotyping strategy focusing on behavioural domains of locomotion, exploration, anxiety and sensorimotor gating.

## Materials and methods

### Animals

The heterozygous *Nrg1* transmembrane domain knockout mice were provided by Prof. Richard Harvey (Victor Chang Cardiac Research Institute, Sydney Australia) and have been described previously (Stefansson et al. 2002). Test animals were heterozygous *Nrg1*<sup>+/-</sup> (*Nrg1* HET) and wild type-like control *Nrg1*<sup>+/+</sup> (WT) littermates (backcrossed for 15 generations onto a C57BL6/J background). Genotypes were determined after weaning (postnatal day 21) by tail biopsy and polymerase chain reaction (primers for mutant *Nrg1* mice: Neo173F 5'-ATGAAGTGCAGGACGAGGCA-3' and Neo6301R 5'-GCCACAGTCGATGAATCCAG-3'; primers for wild-type-like control mice: 5'-AACAGCTGACTGTAAACACC-3' and 5'-TGCTGTCCATCTGACGAGACTA-3'). Male, adult, age-matched ( $\pm 14$  days) test animals of the same genotype were pair-housed in Macrolon cages under a 12:12 h light:dark schedule [light phase: white light (illumination: 80 lx) - dark phase: red light (illumination: <2 lx)] with food and water available ad libitum. Male, adult, age-matched ( $\pm 7$  days), group-housed A/J mice (Animal Resources Centre, Canning Vale, Australia) were used as standard opponents in the social interaction test. All research and animal care procedures were approved by the "Garvan Institute/St. Vincent's

Hospital Animal Experimentation Ethics Committee” and were in agreement with the “Australian Code of Practice for the Care and Use of Animals for Scientific Purposes”.

### Drug treatment

$\Delta^9$ -tetrahydrocannabinol (THC; Sigma-Aldrich, Sydney, Australia) was dissolved in 4% ethanol, 1% Tween 80 and 95% saline (Balerio et al. 2006; Varvel et al. 2006). The drug was administered intraperitoneally (i.p.) in a volume of 10 ml/kg of body weight (BW), 30 min before behavioural testing. Two doses of THC were used in this study, 5 and 10 mg/kg BW (latter one was shown to be CB1 receptor-specific: see e.g., Compton et al. 1996). Relatively high doses of THC were chosen, as cannabis psychosis has been associated with acute administration of high doses of cannabis (Hall and Degenhardt 2000; Johns 2001) and as the C57BL/6J background strain is known for its reduced sensitivity to some of the neurobehavioural effects of THC compared to other mouse strains (Onaivi et al. 1995). Mice were treated with similar volumes of either vehicle (4% ethanol, 1% Tween 80 and 95% saline) or one of the two dosages of THC in a counterbalanced, quasi-randomised design, allowing for within-animal comparison. A washout period of at least 7 days was guaranteed before the animals were retested.

### Behavioural testing

All animals were tested at the age of 6–7 months to investigate whether acute THC treatment has a differential effect on the behavioural performance of *Nrg1* HET compared to WT mice. *Nrg1* hypomorphic mice exhibit an age-dependent hyperactive phenotype, as mutant animals of 3–4 months of age show similar behaviours as wild type-like control littermates, whereas *Nrg1* HET mice older than 4 months exhibit an increase in locomotion and exploration compared to WT mice (Karl et al. 2006). A first set of animals ( $n=10$  per genotype) was tested 30 min postinjection in domains such as locomotion, exploration and anxiety using the open field (OF), the hole board (HB) and the elevated plus maze (EPM) tasks, respectively. A second set of mice ( $n=10$ –13) was tested 30 min postinjection for anxiety, social behaviours and sensorimotor gating using the light–dark (LD), the social interaction (SI) and the prepulse inhibition (PPI) tests (see Table 1). All behavioural testing was conducted during the light cycle under highly standardized test conditions (Crawley 1999; Karl et al. 2003). For habituation purposes, all test animals were transported to the testing room 1 h before behavioural testing (holding and test room were part of the same facility having therefore identical light, air pressure and temperature conditions). Environmental odours were removed from the different test apparatus by

**Table 1** Test biography of both sets of wild type-like (WT) and heterozygous *Nrg1* transmembrane domain knockout (*Nrg1* HET) mice

Sets	Genotype (and animal numbers)	Battery of behavioural paradigms (30 min after i.p. injection of either vehicle or THC)
Set 1	WT: $n=10$ ; <i>Nrg1</i> HET: $n=10$	Behavioural test battery A OF HB EPM
Set 2	WT: $n=10$ ; <i>Nrg1</i> HET: $n=13$	Behavioural test battery B LD SI PPI

A washout period of at least 7 days was guaranteed before the animals were retested in the same battery of behavioural paradigms. Thus, all test animals were observed repeatedly 30 min postinjection in the behavioural test battery A [open field (OF), the hole board (HB) and the elevated plus maze (EPM); set 1] or test battery B [light–dark test (LD), the social interaction test (SI) and the prepulse inhibition paradigm (PPI); set 2]. Two sets of animals were tested over a period of 3 weeks. Mice were treated with either vehicle or one of the two dosages of THC (5 or 10 mg/kg bodyweight) in a counterbalanced, quasi-randomised design, allowing for within-animal comparison.

cleaning the equipment after each trial with a 30% ethanol solution.

**Open field (OF)** Locomotor activity, explorative-like tendencies and anxiety-related behaviours were evaluated by placing the mouse into an infrared photobeam-controlled ( $x$ -,  $y$ -, and  $z$ -axis) open field activity test chamber (MED, Vermont, USA). This paradigm mimics the natural conflict in mice between the tendency to explore a novel environment and to avoid an exposed open area (Crawley 1985; DeFries et al. 1966). The arena (43.2 cm $\times$ 43.2 cm) was divided into a central and a peripheral zone (MED software coordinates for central zone: 3/3, 3/13, 13/3, 13/13). Animals were tested for 30 min (illumination at floor level: 20 lx), and the animal’s total distance travelled (horizontal activity:  $x$ - and  $y$ -axis) and vertical activity ( $z$ -axis) was recorded automatically (software settings: box size: 3; ambulatory trigger: 2; resting delay: 1,000 ms; resolution: 100 ms) in the central and peripheral areas. The ratio of central to total distance travelled and the time spent in the central zone were taken as measures of anxiety (Denenberg 1969).

**Hole board (HB)** The HB test provides independent measures of locomotor activity and directed exploration (Boissier et al. 1964; van Gaalen and Steckler 2000). Mice were placed into the open field activity test chamber, which was equipped with a hole board floor insert for mice (MED: 16 holes; diameter: 1.6 cm). The infrared photobeams

provided automated measures of the distance travelled, ambulatory frequency and *head-dipping* frequency.

**Elevated plus maze (EPM)** The EPM represents the natural conflict between the tendency of mice to explore a novel environment and the tendency to avoid a brightly lit, elevated, open area (Montgomery 1958). The grey plus maze had a central platform (6 cm×6 cm), two alternate enclosed arms (35 cm×6 cm; height of enclosing walls 28 cm; dimly illuminated: 10 lx) and two alternate open arms (35 cm×6 cm; without side walls; highly illuminated: 70 lx) with ledges (4 mm×6 mm). The arms' surface was raised 70 cm above the floor. The mouse was placed onto the centre field of the “+” (faced to an enclosed arm) and was allowed to explore the maze for 5 min. Closed arm entries were taken as measures of motor activity, whereas entries in and time spent on open arms and the percentage of open arm entries (open arm entry ratio) were recorded online as a measure of anxiety (Hogg 1996; Pellow et al. 1985). In addition to recording behaviours online by repeatedly trained research staff, the sessions were reanalysed for parameters such as time spent in the different compartments of the EPM. An individual entry was recorded when the animal entered the arm with at least half of its body length.

**Light-dark (LD)** In the LD test, the time spent in a brightly illuminated zone compared to a dark zone and the occurrence of associated exploratory behaviour (vertical activity) were used to assess anxiety in rodents (Costall et al. 1989; Crawley 1999). The test animals were placed into the open field chamber, which was equipped with a dark box insert for mice (covering half the area of the chamber: MED). An opening located in the centre of the partition connected light (illumination: 20 lx) and dark (illumination: <2 lx) compartments. At the start of the experiment, mice were placed into the lit compartment. The time spent in, entries into and horizontal activity in the differentially illuminated compartments, and vertical activity were recorded during a 10 min test session. The ratio of distance travelled in the light compartment to total distance travelled (distance ratio) and the time mice spent in the light compartment were taken as measures of anxiety.

**Social interaction (SI)** The SI model is widely used to measure anxiety-like behaviours and to detect anxiogenic- and anxiolytic-like effects of drugs (File 1988; Kask et al. 2001). Test animals were placed together with an A/J standard opponent into the open field activity test chamber (in opposite corners), where they were allowed to explore the arena and each other freely for 10 min. The behaviour of the test mouse was recorded online. Frequency and total duration (so-called active social interaction time) of the active socio-positive behaviours *general sniffing*, *anogenital*

*sniffing*, *allogrooming*, *following* and *crawling over/under* were recorded.

**Prepulse inhibition (PPI)** Patients with schizophrenia show impaired sensorimotor gating. PPI is the operational measure of sensorimotor gating, in which a weak pre-stimulus (prepulse) attenuates the startle response (Wang et al. 2003). PPI was tested in two startle chambers (SR-Lab: San Diego Instruments, San Diego, USA). Animals were habituated to the test device for three consecutive days (day 1: 2×2 min; days 2–3: 1×10 min) before being tested 24 h later. The protocol used was adapted from methods developed by Geyer and Swerdlow (1998). Briefly, after a 5 min acclimation period with a 70 dB background noise, the test session began. Each session consisted of 76 trials in a pseudorandomised order: 10×90 dB startle response trials, 18×120 dB startle response trials, 2× prepulse alone trials (prepulse intensities of 74, 78, 82 or 86 dB), 8× PPI response trials (prepulse followed 80 ms later by a 120 dB startle stimulus), and 8× no pulse trials (background noise only). We chose a variable intertrial interval of averaged 15 s (range 10–20 s), prepulse duration of 20 ms and startle duration of 40 ms. Startle response was measured as the average mean amplitude. Percentage of PPI was calculated as [(startle response 120 dB - PPI response)×100/startle response 120 dB]. We also analysed the mean % PPI of all summed prepulse responses for each genotype.

#### Statistical analysis

One-way analysis of variance (ANOVA) was applied to investigate “baseline” differences in the behavioural performance of vehicle-treated WT and *Nrg1* HET mice. The behavioural response of WT and *Nrg1* HET mice to THC was analysed using two-way ANOVA to investigate the effect of the factors “genotype” and “THC dose” on several behavioural domains. This was followed by one-way ANOVA (factor: “genotype” or “THC dose” split by the corresponding factor) and the Student–Newman–Keuls post hoc test, in which vehicle data were compared to each individual dose of THC (5 and 10 mg/kg) within each genotype. Differences were regarded as statistically significant when  $p < 0.05$ . Results present the *degrees of freedom*, *F*-values and *p*-values of one-way (for baseline comparison) or two-way ANOVA (for “THC dose” effects), while in the figures and tables, the *p*-values of the corresponding post hoc tests are provided. Significant post hoc effects of *Nrg1* HET animals vs WT are indicated by “#” ( $p < 0.05$  and “##”  $p < 0.01$ ), whereas significant effects of acute THC treatment vs vehicle treatment are shown by asterisk (\* $p < 0.05$  and \*\* $p < 0.01$ ). All data are presented as means ± standard error of the mean (SEM).



## Results

### Locomotor activity and exploration

The results from the OF, HB and LD are shown in Table 2. One-way ANOVA revealed that vehicle-treated *Nrg1* HET mice exhibited a hyperactive phenotype at baseline as the total distance travelled was significantly increased in the OF [ $F(1,18)=21.0$ ,  $p<0.001$ ] and in the LD [ $F(1,21)=8.8$ ,  $p<0.01$ ] compared to vehicle-treated WT mice. Furthermore, *Nrg1* HET mice crossed significantly more often between the light and the dark compartment of the LD [ $F(1,21)=9.6$ ,  $p<0.01$ ] and entered more often the enclosed arms of the EPM [ $F(1,18)=5.1$ ,  $p<0.05$ ] than WT mice. This hyperactivity was not only evident with regard to locomotion but could be confirmed in explorative-like behaviours as measured by an increased frequency of head dipping in the HB [ $F(1,17)=5.1$ ,  $p<0.05$ ] and of exploration-related vertical activity in the LD test [ $F(1,21)=7.1$ ,  $p<0.05$ ] compared to vehicle-treated WT mice.

THC treatment significantly decreased the total distance travelled in the OF [ $F(2,52)=34.6$ ,  $p<0.001$ ] and the LD test [ $F(2,59)=9.9$ ,  $p<0.001$ ], and the frequency of total crossings [ $F(2,59)=11.4$ ,  $p<0.01$ ] and vertical activity [ $F(2,59)=15.4$ ,

$p<0.001$ ] in the LD test as measured by two-way ANOVA. Interestingly, a differential motor activity- and exploration-suppressing effect of THC was observed between WT and *Nrg1* HET mice as we detected significant genotype by drug interactions for all these parameters: total distance travelled in OF [ $F(2,52)=10.3$ ,  $p<0.001$ ] and LD [ $F(2,59)=5.2$ ,  $p<0.01$ ], frequency of total crossings in LD [ $F(2,59)=5.2$ ,  $p<0.01$ ], and vertical activity [ $F(2,59)=4.2$ ,  $p<0.05$ ] in LD. Thus, there exists a genotype-specific difference in the motor activity-suppressant actions of THC. This conclusion was further supported by post hoc analyses for motor activity/exploration measures conducted within each genotype. In the OF, 10 mg/kg THC significantly reduced the total distance travelled in both genotypes compared to vehicle. However, 5 mg/kg THC significantly inhibited the total distance travelled selectively in the *Nrg1* HET mice (see Table 2). In the LD test, both the 5 and 10 mg/kg doses of THC were effective in reducing the total number of crossings and the frequency of vertical activity of *Nrg1* HET mice. Nevertheless, both cannabinoid doses (5 and 10 mg/kg) were ineffective in reducing these measures in WT mice (see Table 2). Acute THC treatment also decreased the frequency of head dipping [ $F(2,50)=44.1$ ,  $p<0.001$ ] in the HB test as measured by two-way ANOVA. No significant genotype by

**Table 2** Locomotor activity, exploration and anxiety-related behaviours

Parameters	WT mice			<i>Nrg1</i> HET mice		
	Vehicle	THC 5 mg/kg	THC 10 mg/kg	Vehicle	THC 5 mg/kg	THC 10 mg/kg
Open field						
Total distance [cm] “+”	2,017±121	1,607±131	1,190±194**	3,382±272#	1,358±241**	1,073±200**
Time spent in the centre [s]	294±59	68±21**	31±25**	370±50	134±50**	9±2**
Hole board						
Head dipping fq	15±2	3±1**	1±0.4**	23±3#	5±3**	1±0.5**
Elevated plus maze						
Enclosed arm entries	6±1	9±2	4±1	9±1#	12±3	12±3
Time spent in open arms [s]	16±4	10±5	7±3	30±6#	11±5*	14±6
Open arm entries ratio [%]	36±2	19±5	25±7	38±3	16±6*	24±5
Light–dark						
Total distance [cm] “+”	1,248±166	1,172±186	859±145	1,881±137##	981±156**	911±156**
Total crossings “+”	49±12	46±16	22±11	105±13##	29±12**	18±6**
Vertical activity “+”	35±5	25±10	11±7	63±8#	13±8**	6±4**
Time spent in light zone [s] “+”	72±15	86±32	40±25	147±18##	51±27**	25±11**
Distance travelled in light zone [cm] “+”	274±62	264±87	147±77	542±63##	182±78**	71±35**
Entries into light zone fq “+”	24±6	23±8	11±6	52±6##	14.5±6**	9±3**
Distance ratio [%] “+”	18±4	17±5	12±5	29±2#	13±4**	6±3**
Social interaction						
SI duration [s]	42±5	22±5**	12±2**	44±3	16±2**	9±2**
Sniffing fq	29±2	15±1**	9±1**	32±2	14±2**	7±1**
Anogenital sniffing fq	6±1	2±0.3**	1±0.3**	8±1	2±0.5**	0.4±0.2**

“fq” represent frequencies of any given behaviour; parameters with a significant genotype by THC dose interaction are presented with “+”; significant post hoc effects of *Nrg1* HET animals vs WT control animals at baseline are indicated by “#” or “##” ( $\#p<0.05$  and  $\#p<0.01$ ), whereas significant effects of acute THC treatment vs vehicle treatment are shown by “\*” or “\*\*” ( $*p<0.05$  and  $**p<0.01$ ); all data are presented as means ± standard error of the mean (SEM)

dose interaction was observed for this parameter. Both doses of THC (5 and 10 mg/kg) significantly reduced head dipping compared to vehicle in either genotype (Table 2).

### Anxiety

The results from the animal models of anxiety are shown in Table 2. Mice were repeatedly tested in the various animal models of anxiety (OF, EPM, LD and SI) according to a within-subjects, counterbalanced, quasi-randomised design with a 1 week interval between testing. To confirm that the effects of THC were consistent with repeated testing over weeks of administration, we analysed the data using two-way ANOVA (factor 1: week of testing and factor 2: THC dose). Our analyses found no significant week of testing by THC dose interaction effects for any parameter recorded in the different anxiety paradigms (data not shown). This highlights that the effects of THC were reproducible irrespective of whether the mice were naïve or pre-exposed to the test apparatus.

In the LD, one-way ANOVA for baseline behaviour revealed an anxiolytic-like phenotype for the vehicle-treated *Nrg1* HET mice. Accordingly, *Nrg1* HET mice spent significantly more time in the light compartment [ $F(1,21)=9.0$ ,  $p<0.01$ ], showed increased locomotion rates [ $F(1,21)=8.8$ ,  $p<0.01$ ] in the area and exhibited an elevated light compartment entry score [ $F(1,21)=9.6$ ,  $p<0.01$ ] compared to vehicle-treated WT mice. Importantly, this anxiolytic-like profile of *Nrg1* HET mice holds when the locomotion-related parameters were corrected for the total distance travelled in this paradigm as highlighted by a significant increase in distance ratio [ $F(1,21)=6.8$ ,  $p<0.05$ ] for *Nrg1* hypomorphs within the aversive light compartment. Furthermore, vehicle-treated *Nrg1* HET mice spent significantly more time on the more aversive open arms of the EPM [ $F(1,18)=4.4$ ,  $p=0.05$ ]. The reduced anxiety observed in these mutant mice appears to be task-dependent as SI performance was not different between vehicle-treated WT and *Nrg1* HET mice.

Two-way ANOVA revealed that THC had a significant anxiogenic-like effect as it reduced the time mice spent on the open arms of the EPM [ $F(2,52)=4.7$ ,  $p<0.05$ ] and the ratio of open arm entries [ $F(2,52)=7.7$ ,  $p<0.01$ ]. Furthermore, THC treatment resulted in a decrease of time spent [ $F(2,59)=5.8$ ,  $p<0.01$ ] and distance travelled [ $F(2,59)=9.3$ ,  $p<0.001$ ] in the light compartment of the LD, of the frequency of entries into the same compartment [ $F(2,59)=11.4$ ,  $p<0.001$ ] and of distance ratio in this area [ $F(2,59)=7.7$ ,  $p<0.001$ ]. We also found a genotype by THC dose interaction for time spent in the light compartment of the LD [ $F(2,59)=3.4$ ,  $p<0.05$ ], for the frequency of entries into [ $F(2,59)=5.2$ ,  $p<0.01$ ] and the distance travelled in [ $F(2,59)=4.2$ ,  $p<0.05$ ] the same area. In addition, there was a trend for a genotype by THC dose interaction for the distance ratio in the light compartment [ $F$

(2,59)=2.9,  $p=0.06$ ]. Post hoc analysis revealed that THC, at 5 mg/kg, decreased the open arm entry ratio in the EPM for *Nrg1* HET but not WT mice. Furthermore, THC administration, at both doses of 5 and 10 mg/kg, reduced all behaviours measured in the LD in the *Nrg1* HET but not the WT mice (see Table 2).

In the social interaction test, two-way ANOVA revealed no differential effect of THC treatment on *Nrg1* HET and WT mice in any measures derived from this test. THC treatment dose-dependently decreased the total time spent in social interaction [ $F(2,59)=51.8$ ,  $p<0.001$ ], the frequency of sniffing [ $F(2,59)=82.2$ ,  $p<0.001$ ] and the frequency of anogenital sniffing [ $F(2,59)=55.8$ ,  $p<0.001$ ] in a similar fashion in WT and *Nrg1* HET mice. No aggressive interactions were observed between test animals and the opponent A/J mice.

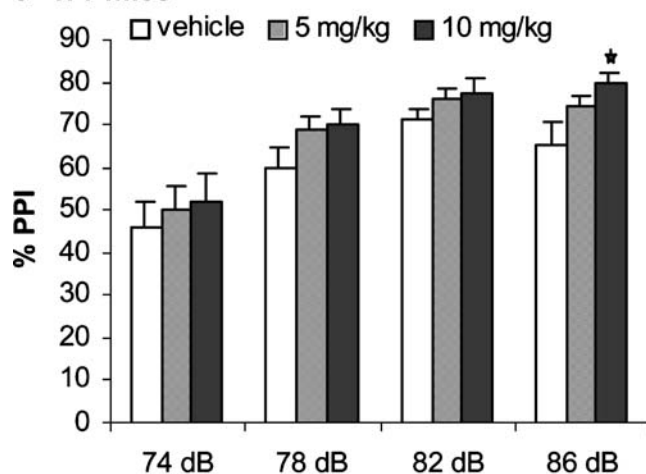
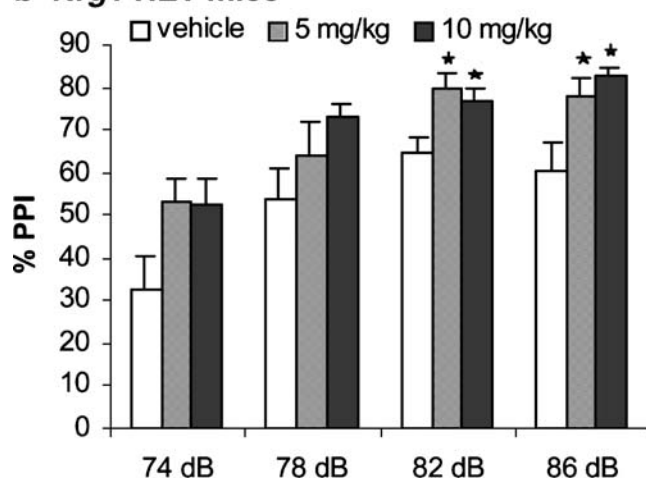
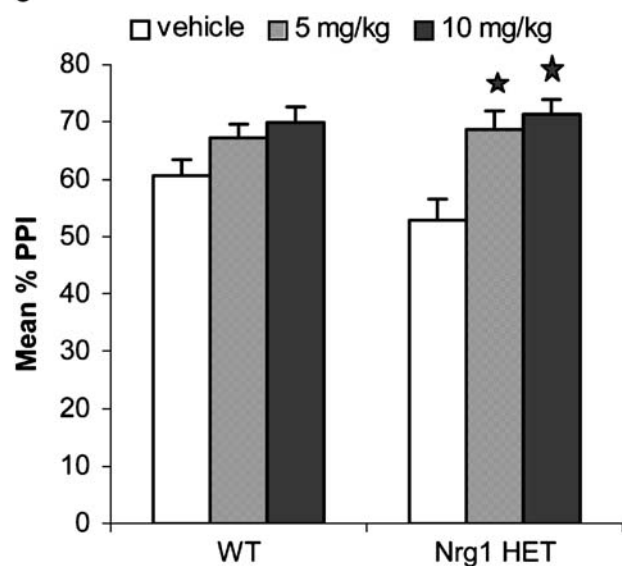
Vehicle-treated *Nrg1* HET mice showed no significant difference to vehicle-treated WT mice in the total time spent in the centre of the arena of the OF. However, two-way ANOVA showed that THC treatment significantly decreased the time spent in the centre of the OF [ $F(2,52)=32.7$ ,  $p<0.001$ ], which is a parameter for anxiety-related behaviour. Post hoc analysis revealed that in both genotypes, acute THC treatment (both 5 and 10 mg/kg) decreased the time spent in the centre of the OF compared to vehicle-treated animals.

### Prepulse inhibition

Figure 1a–c presents the data from PPI testing. The prepulse inhibition (measured as % PPI) was elevated for both genotypes with increased prepulse intensities. At baseline, no differences were observed between vehicle-treated WT and *Nrg1* HET mice with regard to % PPI (Fig. 1a,b).

Two-way ANOVA revealed that acute THC administration significantly enhanced % PPI [ $F(2,59)=6.4$ ,  $p<0.01$ ; Fig. 1a,b]. Post hoc analyses indicated that in the *Nrg1* HET mice, the increased % PPI was observed for both THC doses at prepulses of 82 and 86 dB (Fig. 1b), whereas in WT mice, only treatment with 10 mg/kg THC significantly increased % PPI at the 86 dB prepulse (Fig. 1a). When % PPI was calculated as the % PPI of the mean PPI responses across all prepulse intensities, post hoc analysis revealed that both the 5 and 10 mg/kg THC doses increased % PPI in the *Nrg1* HET animals with no corresponding effects being observed in the WT mice (Fig. 1c).

No differences were observed between vehicle-treated *Nrg1* HET and WT mice when measuring their startle response to a 120 dB startle stimulus (Fig. 2). Two-way ANOVA revealed that THC treatment [ $F(2,59)=6.1$ ,  $p<0.01$ ] significantly reduced the startle response. Post hoc analysis revealed that the startle response-inhibiting effect of THC was only evident in *Nrg1* HET after treatment with 10 mg/kg THC and was not observed in the WT animals.

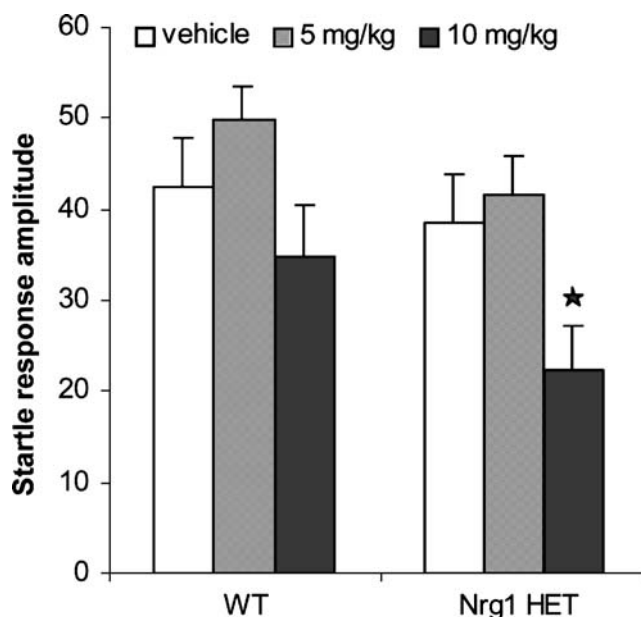
**a WT mice****b *Nrg1* HET mice****c**

**Fig. 1a–c** Prepulse Inhibition (PPI). The effect of acute THC treatment (30 min after i.p. injection of either vehicle or THC: 5 or 10 mg/kg BW) on % PPI [(startle response 120 dB–PPI response)×100/startle response 120 dB] was measured in WT and *Nrg1* HET mice ( $n=10–13$ ). Different prepulse stimuli were used (4/8/12/16 dB above 70 dB background noise): **a** % PPI of WT mice, **b** % PPI of *Nrg1* HET mice, and **c** mean of % PPI of all summed prepulse responses for WT and *Nrg1* HET test animals. All data are presented as means  $\pm$  standard error of the mean (SEM). Significant effects of acute THC treatment vs vehicle treatment are shown by “\*” ( $p<0.05$ )

**Discussion**

Vehicle-treated *Nrg1* HET mice expressed a hyperactive, hyper-exploratory and anxiolytic-like phenotype compared to WT animals. Importantly, mice hypomorphic for the *Nrg1* gene were more susceptible to THC's locomotor suppressant actions and its stimulating effect in an animal model for sensorimotor gating. In animal models of anxiety, less anxious *Nrg1* HET animals were no more affected by acute THC administration than their WT controls in an array of different models (e.g., social interaction and EPM). However, the *Nrg1* HET mice showed a greater anxiogenic response to THC than WT in the LD test, highlighting that the effects of THC on anxiety in these animals might be task-dependent.

The acute administration of THC decreased spontaneous locomotor activity and exploration in the open field test. Importantly, THC promoted a greater reduction in locomotor and exploratory activity in the *Nrg1* HET mice



**Fig. 2** Startle response. The startle response (averaged startle amplitude) in WT and *Nrg1* HET mice ( $n=10–13$ ) was measured for a 120-dB startle stimulus 30 min after i.p. injection of vehicle or THC (5 or 10 mg/kg BW). All data are presented as means  $\pm$  standard error of the mean (SEM). Significant effects of acute THC treatment vs vehicle treatment are shown by “\*” ( $p<0.05$ )

compared to their WT controls, suggesting that the mutant mice are more susceptible to the well-established locomotor suppressant effects of cannabinoids (Arnold et al. 1998, 2001). It could be argued that this hypersensitivity may simply reflect the *Nrg1* HET mice being relatively hyperactive at baseline, thus, providing more room to delineate the locomotor suppressant effects of THC. However, a floor effect can be ruled out, as locomotor activity of both genotypes is not completely suppressed by the acute THC treatment (see, e.g., total distance in OF—Table 2). One likely explanation for the *Nrg1* HET animals' enhanced sensitivity to the locomotor inhibitory effects of cannabinoids may be based on developmental adaptations during ontogeny in the germline *Nrg1* hypomorphic mice (e.g., increased expression of CB1 receptors or greater sensitivity of CB1 receptors to cannabinoid ligands in motor-related regions). The observation that *Nrg1* HET mice are hyperactive, combined with the finding that *Nrg1* administration acts to inhibit locomotor behaviour in hamsters (Snodgrass-Belt et al. 2005), indicates that *Nrg1* may function as an inhibitory counterbalance to excitatory neural processes mediating locomotor behaviour.

Vehicle-treated *Nrg1* hypomorphic mice exhibit an anxiolytic-like phenotype, which might be influenced by their hyperactivity. However, we detected an increase in the time hypomorphs spent in aversive areas. Thus, the decrease in anxiety is evident within a behavioural parameter, which is relatively unaffected by an unspecific increase in motor activity. These baseline differences exist for spatio-temporal (i.e., LD and EPM) rather than socio-temporal paradigms (i.e., SI). This task-dependent anxiolytic phenotype suggests an impact of *Nrg1* on certain explorative/locomotive aspects of anxiety (Rodgers 1997; Rodgers and Johnson 1995). Importantly, anxiety symptoms (and high autonomic arousal) are very variable accompaniments to schizophrenia—often schizophrenia patients are diagnosed with more than one anxiety disorder (Braga et al. 2005; Tibbo et al. 2003; Townsend and Wilson 2005). Obviously, any animal model for candidate genes of schizophrenia models only some, but not all, aspects of the mental disorder. Future research has to clarify if the specific anxiolysis of *Nrg1* hypomorphs is related to the comorbidity between anxiety disorders and psychosis reported in schizophrenia or if it is a direct effect of reductions in *Nrg1* expression levels.

It has been widely reported that administration of cannabinoids stimulates anxiety-related behaviours in rodents (Onaivi et al. 1990, 1995; Rutkowska et al. 2006). In our study, we observed a task-specific increased susceptibility of *Nrg1* HET mice to the anxiogenic effects of THC compared to control animals. A confounding effect of the increased sensitivity of the *Nrg1* HET mice to the locomotor suppressant effects of THC can be ruled out, as spatiotemporal parameters of other anxiety paradigms

(i.e., OF and EPM) were not differentially affected by THC treatment. It is possible that the enhanced sensitivity of the *Nrg1* HET animals to the anxiogenic effects of THC, as measured in the LD test, may reflect a specific anxiogenic-like action of THC on exploration/locomotion-related features of anxiety, which might be expressed to a greater extent in the mutant mice. These features are linked to how an organism adapts and responds to its spatial surrounding rather than to its social environment. Interestingly, both altered habituation to a novel environment in animal models (O'Tuathaigh et al. 2006) and social withdrawal in humans (Dixon et al. 1994) are described within the schizophrenia context. Further investigations should use a battery of different anxiety models screening for conditioned (conflict and other paradigms) and unconditioned responses (exploration, social, antipredator and other) after THC treatment to clarify this task-specificity (Rodgers 1997; Rodgers and Johnson 1995).

Within-subjects designs are commonly utilised in psychopharmacological mutant mice studies due to the difficulty in generating sufficient animal numbers. Importantly, altered anxiety-related behaviour (Espejo 1997; Lee and Rodgers 1990; Nyberg et al. 2003; Rodgers et al. 1996) and modulated sensitivity to the effects of psychotropic drugs upon repeated testing of mice in the same anxiety model (Holmes and Rodgers 1999; Rodgers et al. 1992; Rodgers and Shepherd 1993) are well-known phenomena. For instance, modulators of GABA<sub>A</sub> function (e.g., benzodiazepines) have diminished anxiolytic effects when administered to animals with prior test experience of the EPM. This phenomenon is known as “one trial tolerance” (Bertoglio and Carobrez 2002; File et al. 1990; Holmes and Rodgers 1999; Rodgers et al. 1992; Rodgers and Shepherd 1993). Our statistical analyses revealed no such altered effectiveness of THC with repeated testing, as mice displayed consistent anxiogenic responses to THC, whether they were naïve or had prior experience of the test apparatus. However, many studies demonstrating “one trial tolerance” to the effects of benzodiazepines use a 24 h test–retest interval (Albrechet-Souza et al. 2005; Cruz-Morales et al. 2002; File 1990; Holmes and Rodgers 1999). Therefore, it is possible that THC may have distorted effects with a shorter than 7 day intertrial interval, although the selective CB1 receptor antagonist AM 251 also showed consistent anxiety-modulating effects on EPM-naïve or -experienced mice using a 24 h test–retest interval (Rodgers et al. 2005). Interestingly, Bouwknecht et al. (2004) reported a relatively small impact of repeated testing on baseline anxiety behaviour of C57BL/6 mice using a 7 day intertrial interval. Furthermore, they showed consistent effects of the 5HT<sub>1A</sub> receptor agonist, flesinoxan, with repeated testing in anxiety models. Our results suggest that with a 1 week intertrial period, THC administered according to a within-subjects design provides



consistent behavioural results in various animal models of anxiety.

PPI is a phenomenon, whereby a pre-stimulus reduces the magnitude of the normal startle response to an intense startling stimulus (Graham 1975; Hoffman and Searle 1968). Importantly, motor activity differences (as shown for our animal model) do not have an impact on the startle reflex to an acoustic startle stimulus (Leng et al. 2004). In the current study, 5 and 10 mg/kg THC exposure significantly increased PPI in the *Nrg1* HET mice, with the corresponding effect of THC not being as pronounced in WT animals. This increased effect of acute THC on PPI in the *Nrg1* HET mice appears to be valid, at least for the 5 mg/kg THC dose, as it was ineffective in reducing the startle response. The observation that 10 mg/kg THC significantly reduced startle in the *Nrg1* HET animals further supports that these mice are more sensitive to sedative actions of cannabinoids than WT animals.

Studies investigating the effects of cannabinoids on PPI in rodents are replete with inconsistencies. The anandamide reuptake and degradation inhibitor, AM404, and the synthetic cannabinoid receptor agonists, WIN 55,212-2 and CP 55,940, have been shown to promote deficits in PPI in some studies (Fernandez-Espejo and Galan-Rodriguez 2004; Mansbach et al. 1996; Martin et al. 2003; Schneider and Koch 2002) but not in others (Bortolato et al. 2005, 2006; Stanley-Cary et al. 2002). Two recent studies have directly examined the effects of THC on PPI (Malone and Taylor 2006; Nagai et al. 2006). Similar to our findings using wild type-like control mice, Malone and Taylor (2006) showed that THC (1 and 3 mg/kg) had no effect on PPI when administered to group-housed control rats. However, isolation-reared rats—suggested as another animal model for schizophrenia—exhibited PPI disruptions after THC treatment. Importantly, a genetic and environmental model should not be treated as identical models for schizophrenia as too little is known about biochemical and behavioural differences between these models. For example, socially isolated rats express increased anxiety-like behaviours (Wright et al. 1991) unlike *Nrg1* HET mice, which display reduced anxiety-related behaviour. Such differences and the fact that these studies were conducted on distinct species may account for the opposing effects of THC on PPI. Furthermore, it is widely accepted that any given animal model for schizophrenia can only represent certain aspects of this mental disorder not its entire complexity (Ellenbroek and Cools 2000; van den Buuse et al. 2005). Therefore, a direct comparison of a singular animal model (i.e., isolation rearing) with a two-hit model (i.e., *Nrg1* depletion combined with drug abuse) would be overly simplistic.

Nagai et al. (2006), using almost identical doses of THC (6 and 10 mg/kg) to that employed here, demonstrated CB1 receptor-mediated PPI deficits in mice. The lack of

correlation between the findings of Nagai et al. (2006) and our results in wild type-like mice are likely due to strain differences in the effects of cannabinoids as the C57BL6/J background strain used in our study has been shown to be less sensitive to the neurobehavioural effects of THC compared to other inbred strains (Arnold et al. 2001; Onaivi et al. 1995).

Given that chronic and -less robust -acute (Favrat et al. 2005; Linszen et al. 1994) THC exposure is thought to precipitate psychosis in vulnerable individuals (Degenhardt et al. 2003; Hall 1998) and that other known psychosis-promoting agents such as phencyclidine and amphetamine (Geyer et al. 2001; Mansbach and Geyer 1989; Mansbach et al. 1988; Martin et al. 2003) disrupt PPI, the observation that THC exposure increased PPI selectively in *Nrg1* HET animals was unexpected. The self-medication theory is an alternative theory for why people with schizophrenia are more likely to administer drugs of abuse than the general population (Arseneault et al. 2004; Degenhardt et al. 2001; Schneier and Siris 1987; Schofield et al. 2006). According to this theory, schizophrenia patients try to alleviate the symptoms associated with the disorder and/or to counter the side effects of antipsychotic medication. Interestingly, the most widely used drug in the schizophrenia population, nicotine, increases PPI in rats and reverses PPI deficits in schizophrenia patients (Acri et al. 1994; Kumari et al. 2001; Postma et al. 2006). Thus, schizophrenia patients may self-medicate with nicotine to reverse their cognitive deficits. Similarly, our observation of THC-induced enhancement of PPI supports the hypothesis that acute THC may have a partly beneficial action in schizophrenia patients by improving a negative symptom such as attentional dysfunction. Unfortunately, there is no satisfactory animal model for a variety of positive symptoms (i.e., hallucinations, delusions) of schizophrenia; hence, the effects of THC on these core aspects of the mental disorder cannot be concluded upon by the current investigation. However, it is interesting to note that THC, like clozapine, acts to reverse the hyperactive phenotype of *Nrg1* HET mice (Stefansson et al. 2002). On the contrary in human studies, it appears cannabis may acutely exacerbate positive schizophrenia symptoms (Grech et al. 2005; Linszen et al. 1994; van Os et al. 2002). Another study discusses the potential bidirectional interactions between cannabis use and psychosis (Hides et al. 2006).

Given that *Nrg1* HET mice provide a putative animal model of schizophrenia, the current results are consistent with the notion that *acutely* administered THC may reverse this schizophrenia phenotype, decreasing hyperactivity and enhancing attention as measured by PPI. As mice were tested repeatedly, behavioural alterations in *Nrg1* HET mice could be influenced by disrupted habituation and novelty processing—both have been considered central to the cognitive deficits observed in schizophrenia (O’Tuathaigh

et al. 2006). These results may provide an animal model of genetic vulnerability to self-medication using cannabinoids, where users may initiate cannabis use as a means to dampen some symptoms of schizophrenia. However, future investigations are needed to examine the effects of *chronic use* of cannabinoids on the *Nrg1* HET mice, as it is the extended use of cannabis that is more strongly associated with precipitating psychosis and exacerbating symptoms in schizophrenia patients.

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