Review Article



Pharmacogenomics of osteoporosis: Opportunities and challenges

T.V. Nguyen and J.A. Eisman

Bone and Mineral Research Program, Garvan Institute of Medical Research, Sydney, Australia

Abstract

The genetics of osteoporosis can be considered in two broad areas: disease susceptibility and drug activity. While the former has been studied, the latter is still largely untouched. Pharmacogenomics is the utilization of genetic information to predict outcome of drug treatment, with respect to both beneficial and adverse effects. The pharmacotherapy of osteoporosis is characterized by variability in therapeutic response with limited prediction of response on a patient-by-patient basis. This is particularly problematic in a clinical situation where therapy is typically required for several years before outcomes can be evaluated for an individual. Thus, the emerging field of pharmacogenomics holds great potential for refining and optimising pharmacological treatment of osteoporosis. Key components for future development of the pharmacogenomics of osteoporosis should include improved understanding of mechanisms of drug action, identification of candidate genes and their variants and expansion of clinical trials to include genetic profiling. This approach could provide clinicians and scientists with powerful tools to dissect novel molecular pathways involved in osteoporosis and to identify new drug targets. The iterative combination of innovative genomics with classical endocrinological approaches in osteoporosis research can be examined as a model of biological research and innovate therapeutical approaches in a continuing interaction between clinical science and basic research.

Keywords: Osteoporosis, Fracture, Bone Mineral Density, Vitamin D Receptor Gene, Collagen I alpha-1 gene, Pharmacogenetics

Background

Osteoporosis, the most common and serious skeletal "disease", is highly prevalent among the elderly of both sexes, with a combined lifetime risk for hip fracture, forearm fracture and vertebral fractures coming to clinical attention being around 40%, equivalent to the risk for cardiovascular disease¹. In the United States alone, osteoporosis affects 25 million people, and incurs estimated costs of \$60 billion. The lifetime risk of hip fracture, the most serious fracture, in Caucasian women is one in six; somewhat higher than the risk of diagnosis of breast cancer (one in nine)2. Mortality is a frequent occurrence in the

increased mortality in both men and women³. Osteoporosis is defined as a systemic skeletal disease characterized by low bone mass and microarchitectural deterioration of bone tissue resulting in increased bone fragility and susceptibility to fracture. Bone mass is traditionally quantified by bone mineral density (BMD). At any specific

months immediately following a hip fracture³. Fifty percent of surviving individuals will need help with daily living activities,

and 15 to 25 percent will need to enter a long-term care insti-

tution shortly after the fracture³. Moreover, all major osteo-

porotic fractures are associated with a two- to three-fold

skeletal region, BMD can be accurately and reliably measured by dual X-ray absorptiometry (DXA); as the amount of mineral per area of bone imaged.

Although fracture is the clinically relevant endpoint of osteoporosis, BMD is a primary predictor of fracture risk⁴⁻⁶. Each standard deviation decrease in BMD is associated with a two-fold increase in the fracture risk⁷. The BMD – fracture relationship generally applies across the skeleton, with some site-specificity; i.e., hip fracture risk is more related to BMD measurements at the hip than lumbar spine or forearm. The relationship between fracture risk and BMD measurements

Dr. Nguyen has no conflict of interest. Dr. Eisman has consultancies with several companies.

Corresponding author: Dr. Tuan V. Nguyen, Bone and Mineral Research Program, Garvan Institute of Medical Research, 384 Victoria Street, Darlinghurst, NSW 2010, Australia

E-mail: t.nguyen@garvan.org.au

Accepted 1 December 2005

is comparable to the relationship between stroke and blood pressure readings. In the same way that "hypertension" relates to cut-off value for blood pressure measurements, osteoporosis is based on a value for BMD below a cut-off threshold. Moreover, as for blood pressure, there is no threshold of BMD that discriminates absolutely between those who will or will not have a clinical event. Hence, a "normal" BMD measurement is no guarantee that fracture will not occur; only the risk is relatively low. Conversely, if BMD is in the osteoporotic range, then fractures are more likely, but still may not occur. At age 50 years, the proportion of women with osteoporosis who will fracture their hip, spine, forearm or proximal humerus in the next 10 years - i.e., positive predictive value - is about 45%. The detection rate for these fractures (sensitivity) is lower, i.e., a significant proportion of fractures occur in women above this BMD threshold.

BMD is not the only predictor of fracture; rather, it is the interaction of increased force, i.e., in falls, and decreased bone "strength". Although falls prevention may influence osteoporotic fracture prevention, determinants of bone "strength" have been the major focus to date with many treatments being developed and applied effectively clinically. BMD is a valuable predictor of future fracture risk, and partly accounts for bone size, but not bone structure, including mass distribution and quality, which are considered to contribute to bone strength. Quantitative ultrasound measurements, broadband ultrasound attenuation (BUA) and speed of sound (SOS), have been proposed as additional measures of bone quality, related to trabecular connectivity and bone matrix. Moreover, many clinical studies have supported a role for normalization of elevated bone turnover as part of the anti-fracture efficacy of the widely used anti-resorptive therapies.

Osteoporosis is, therefore, a complex disease. Its complexity is not just characterized by the multiplicity of clinical aspects, but also multiple determinants. Like many other multifactorial diseases, osteoporosis is determined by environmental factors, by genetic susceptibility and likely by the interaction between these factors. Genetic variations do not necessarily cause osteoporosis or fracture, but they can influence a subject's susceptibility to specific environmental factors and so modify the disease risk. This implies that each subject in the population has a unique risk profile that can change with time. Hence, population data can be only cautiously extrapolated to the individual subject. Yet, at present, decisions about diagnosis and treatment of osteoporosis are still based on statistical data of the subjects' general population. Clearly, this generalized average approach is suboptimal compared with an individualized approach, according to individual genetic and environmental risk profile. Osteoporosis presents an ideal case for such an approach, because of its strong genetic precipitation and high variability in the susceptibility of fracture risk among individuals. In this framework, the principles of pharmacogenomics, which seek to correlate phenotypes and biomarkers by taking advantage of genomic technology, could be applied to identify the actual genetic basis of inter-individual variation in drug efficacy.

Genetics of osteoporosis

Heritability of fracture. Data on heritability of fracture per se are scarce. In a Finnish twin study, approximately 35% of the variance in the liability to fracture (in both males and females) was attributable to genetic factors8. In a recent family study, approximately 25% of the liability to one fracture type, i.e., Colles' fracture of the wrist, was attributable to genetic factors9. Familial analysis within the Study of Osteoporotic Fracture¹⁰ suggests that women, whose mother had had a hip fracture, had a two-fold increase in risk of hip fracture compared with controls. The risk of hip or other fractures was three-fold higher with a paternal history of wrist fracture. In two small studies of osteoporotic women with vertebral or hip fractures, their daughters had bone density deficits intermediate between their mothers and "expected" at the site of their mothers' fracture i.e., lumbar spine or proximal femur^{11,12}. Similar observations have been made in both elderly men and women¹³.

Heritability of bone mineral density. Most genetic studies on osteoporosis have focused on the predictive bone phenotypes, such as BMD. Although BMD is determined by both environmental and genetic factors, it has been estimated from twin studies that 70% to 80% of variance of BMD measured at the lumbar spine and femoral neck is attributable to genetic factors in twin samples¹⁴⁻¹⁸. Heritability of forearm BMD appears to be lower than that in either the femoral neck or lumbar spine^{17,18}. In these studies, there is evidence for pleiotropic effects, i.e., BMD in various skeletal sites being determined by both common and site-specific sets of genes^{16,21}.

Genetic influence on bone turnover. Change in BMD during adult life is the result of the net imbalance between bone formation and bone resorption. These are typically assessed by measurements (in blood or urinary excretion) of various products of osteoblast (bone formation) and osteoclasts (bone resorption) cell activity. Indices of bone formation include osteocalcin, bone-specific alkaline phosphates, procollagen I carboxy-terminal and amino-terminal propeptides. Indices of bone resorption include urinary excretion of hydroxyproline or more specifically pyridinoline cross-links, and more recently, urinary type I collagen cross-linked Ntelopeptides and urinary or serum type I collagen C-telopeptide breakdown products. Genetic factors have been shown to contribute significantly to the inter-individual variance of bone formation markers (both osteocalcin and collagen Cterminal propeptide of type 1 collagen) in premenopausal twins¹⁹⁻²¹.

Heritability of quantitative ultrasound (QUS). The genetic influence on different types of QUS measurements, namely BUA and SOS, has been shown to be 0.53 to 0.82²². BUA measurements have been reported to be more strongly correlated between mothers and their postmenopausal rather than their premenopausal daughters; i.e., the reverse of what has been reported for DXA measurements^{23,24}. These observations suggest that different genetic influences

act on components of the bone phenotype as measured using QUS and DXA. There are no data on heritability of speed-of-sound measures along cortical bone. Genetic correlations observed between transmission QUS and BMD measurements have been moderate, 0.32 to 0.59²². Thus, genes that influence variation in BMD might, but not necessarily, influence variation in QUS, and vice versa. This is consistent with QUS measuring additional non-density characteristics of bone. In any case, a significant part of the variability of QUS and DXA BMD measurements appears unrelated, consistent with their assessment of some distinct bone phenotypic characteristics.

The recognition that various bone-related traits are largely determined by genetic factors has led to an intensive search for specific genes either linked or associated with these traits. Gene-search studies have focused on bone mineral density using the two major approaches of genome-wide screening and candidate genes. The candidate gene approach is based on a priori knowledge of the potential function of the gene involved, and takes advantage of the relevant and known biochemical pathway of bone physiology. Based on this commonly used approach, currently 16 genes have been proposed as potential candidates for bone mineral density, including vitamin D receptor, collagen type $I\alpha 1$, osteocalcin, IL-1 receptor antagonist, calcium sensing receptor, α2HS glycoprotein, vitamin D binding protein, osteopontin, osteonectin, estrogen receptor α, interleukin-6, calcitonin receptor, collagen type $I\alpha 2$, parathyroid hormone, and transforming growth factor α1. As well as the above gene polymorphisms, other polymorphisms in genes including other steroid receptor, cytokine, and bone matrix proteins genes, and more "distant" osteoporosis candidate genes such as apolipoprotein E have also been suggested^{25,44-50}. A feature of these candidate gene studies has been the wide range in positive and negative outcomes and, even in consistent studies, the wide range of effect sizes.

The candidate genes identified so far have been neither strong nor consistent enough to have major clinical predictive value. More importantly, since their relationship with bone biology was the basis of their initial study as "candidates", none could provide novel targets for development of new therapies. It seems likely that minor variations in the regulation or function in several genes, each making relatively small contributions, interact to make up the genetic component of osteoporosis. Under this scenario, individual studies seeking to establish an association between a candidate gene and markers of the disease may yield spurious results. The best strategies for resolving the genetic and environmental contributions to such a polygenic disease such as osteoporosis are not clear.

The less common alternative approach of genome-wide scan has yielded interesting findings. By using linkage analysis of data from a family with osteoporosis-pseudoglioma syndrome (OPS), a disorder characterised by severely low bone mass and eye abnormality, investigators were able to localise the OPS locus to chromsomal region 11q12-13⁵¹. At

the same time, a genome-wide linkage analysis of an extended family with 22 members among whom 12 had very high bone mass (HBM) suggested that the HBM locus also located within 30cM region of the same locus⁵². In follow-up studies using the positional candidate approach both research groups found that a gene encoding the low-density lipoprotein receptor-related protein 5 (LRP5) was linked to both OPS and high bone mass^{53,54}. The finding that LRP5 gene is linked to high bone mass was subsequently confirmed in a family study which included individuals with exceptionally high BMD but were otherwise phenotypically normal⁵⁵. This study showed that a missense mutation (G171V) was found in high-BMD individuals⁵⁵. A recent family study further identified six novel mutations in LRP5 among 13 confirmed polymorphisms that were associated with different conditions with increased BMD⁵⁶. The conditions included endosteal hyperostosis, Van Buchem disease, autosomal dominant osteosclerosis, and osteopetrosis type I. The association between LRP5 and BMD has also recently been shown in a general unselected population⁵⁷.

An example of the success of the combined linkage and association strategy was demonstrated in a recent study in 207 nuclear families with 1323 individuals of Icelandic ancestry. In this study, by first performing a genome-wide scan in the families, the BMP2 gene was found to be linked to the variation in BMD; and by association analysis, with increased fracture risk and BMD⁵⁸.

Pharmacology of osteoporosis

In the past decade there have been remarkable advances in the understanding of basic bone biology leading to targeted approaches both in the prevention and effective treatment of osteoporosis. Among the approved pharmacological therapies for osteoporosis, estrogen replacement therapy, selective estrogen receptor modulators (SERMs), calcitonin, vitamin D derivatives, potent bisphosphonates, and parathyroid hormone have been introduced to clinical use⁵⁹. Most of these drugs, with the exception of parathyroid hormone, act as anti-resorptive agents, and thus decrease bone loss.

Paradoxically, although these agents have modest effects of increasing bone density, they exert a much greater effect on fracture risk reduction. For example, alendronate (a potent bisphosphonate) can improve BMD by 4 to 8%, which based on the BMD-fracture relationship, could be expected to reduce fracture risk by 12 to 28%; but in reality, the fracture risk among alendronate-treated patients was reduced by about 50%. Potential mechanisms for this higher-than-BMD-expected reduction are still being evaluated.

A common feature of all clinical trials involved pharmacological intervention in osteoporosis is that the efficacy and safety are highly variable among patients, such that the range of response is considerable, ranging from "success" to little or no response. For example, the standard deviation of change in BMD induced by potent bisphosphonates is more than twice the rate of change⁵⁹. As a result, while the majority of patients experience an increase in BMD, a small proportion (perhaps 5 to 10%) of patients apparently still lose bone. Thus, although very few patients experience absolutely no therapeutic effects following typical anti-resorptive treatment, no treatment currently prevents all fractures. Moreover, some subjects experience significant adverse effects. These effects, both positive and negative, are experienced over years. Thus, while these treatments are overall beneficial, no reliable means exist to predict who will experience unfavourable or an adverse effect of some type.

Pharmacogenomics

On the clinical level, drug response is affected by many factors, including age, sex, ethnicity, and concomitant disease or drug therapy. However, it is also possible that genetic factors affect the variability in drug response⁶⁰. Indeed, evidence of genetic influence on inter-subject drug response had been reported as early as the 1940s, in the case of peripheral neuropathy in a substantial number of patients treated with the anti-tuberculosis drug, isoniazid⁶¹. It was also observed that African-American soldiers given antimalarial drugs were more likely than their Caucasian colleagues to develop haemolytic anaemia, due to inherited metabolic enzyme differences⁶². Further evidence of genetics of drug response was found in twin studies, in which identical twins were more similar than non-identical twins in regard to the plasma half-life of numerous drugs, providing the best experimental indication of strong genetic components in drug elimination⁶³. Thus, genetic factors may determine an individual's response to pharmacological therapy of osteoporosis and their susceptibility to adverse drug reactions, for each specific drug. Although drug elimination and metabolism can be relatively easily studied, other components that influence drug effects are either unknown or are more difficult to study.

On the molecular level, pharmacological agents act by interacting with proteins such as receptors, enzymes and intracellular signalling proteins. Therefore, when a drug is taken, its absorption, distribution, excretion and pharmacological responses are likely determined by the interactions among those factors, including carrier proteins, transporters, metabolising enzymes, receptors, and co-factors. Members of the cytochrome P450 (CYP), including CYP2D6, 3A4/3A5, 1A2, 2E1, 2C9, and 2C19 are known to influence drug efficacy and toxicity^{64,65}. For example, patients who are homozygous for the CYP2D6 null alleles exhibit a poor metabolizer phenotype, with impaired degradation and excretion of many drugs, including debrisoquine, metoprolol, nortriptyline, and propafone⁶⁴. These poor metabolizers are more likely to exhibit adverse drug reactions. The frequency of this recessive trait ranges from 1% to 2% in Asians, 5% in African Americans and up to 6% to 10% in Caucasian populations⁶⁶⁻⁶⁹. Similarly, patients who are homozygous for the "null" allele of the P450 isoform CYP2C19 are highly sensitive to omeprazole, diazepam, propranolol, mephenytoin, amitriptyline, hexobarbital and

other drugs⁶⁴. The CYP2C19 poor metabolizer phenotype comprises 2% to 5% of Caucasians and 3% to 23% of Asians, resulting largely from a single base pair mutation (A→G) in exon 5 of the coding region 7⁷⁰. Another polymorphically expressed member of the cytochrome P450 family, CYP2C9, metabolizes ibuprofen, naproxen, piroxicam, tetrahydrocannabinol, phenytoin, tolbutamide, and S-warfarin⁷¹; some of which have narrow therapeutic indices. Amino acid substitutions at codons 144 and 359 in the coding region of CYP2C9 result in a 5-fold decline in metabolic activity. Although the frequency of these 2 allelic variants is uncertain, approximately 25% of Caucasians appear to be heterozygous for one or the other variant, leading to a predicted frequency of 5% for the compound homozygous genotype⁷².

Pharmacogenomics of osteoporosis

Some drugs used in osteoporosis therapy, bisphosphonates for example, are not subject to metabolism, but many others are metabolized to active components or as part of their elimination pathway. Despite the evidence of genetic effects on the variation in efficacy and safety of pharmacological agents in other diseases, these are still largely untested in the treatment of osteoporosis, but their potential is underlined by their rapid adoption in disciplines such as obesity and hypertension.

Nevertheless, recent evidence suggests that genetic factors may mediate the response to drug treatment⁷³, and modify the dynamic association between bone turnover markers and bone density. A recent series of studies by Palomba and colleagues⁷⁴⁻⁷⁶ suggested that among postmenopausal women who were on alendronate and hormone replacement therapy (HRT) treatments, the b allele of the VDR's Bsm-I polymorphisms was associated with a greater increase in BMD than those carriers of the B allele. However, interestingly, among patients on RLX the B allele carriers were associated with a greater increase in BMD than the b allele carriers. As a result of the opposite effects, among those on combined ALN and RLX there was no significant association between VDR polymorphisms and BMD change. These results clearly illustrate the interaction between VDR polymorphisms and various anti-resorptive drug therapies in BMD change.

In a study of 21 premenopausal Caucasian women who were homozygous for the VDR genotypes (BB or bb), it was found that baseline osteocalcin, 1,25-(OH)₂D, type I collagen carboxyterminal telopeptide, and inorganic phosphate levels were significantly higher and spinal bone mineral density was significantly lower in the BB allelic group. However, after calcitriol administration, similar serum levels of 1,25-(OH)₂D were attained in both genotypic groups. The increase in serum osteocalcin levels in the BB group was significantly less than that in the bb group. The genotype-related baseline difference in osteocalcin levels was not apparent at similar serum 1,25-(OH)₂D levels. By contrast, baseline differences in phosphate and type I collagen carboxytermi-

nal telopeptide persisted throughout the study. Moreover, parathyroid hormone was less suppressed in the low bone density group despite similar ionized calcium levels⁷⁷.

The VDR gene polymorphisms may also affect the dynamic association between dietary calcium intake and bone density. For example, on lower dietary calcium intakes, gut calcium absorption in women with VDR's *BB* genotype did not increase, but those with *bb* genotype did^{78,79}. The difference in gut calcium absorption between the two alternate homozygotes for the vitamin D receptor start codon polymorphism was 42% 80. As with other studies of bone and genetics, a number of studies have found positive relationships between the vitamin D receptor gene alleles and calcium homeostasis 80-82, while other studies have been negative 83-86.

Similarly, longitudinal studies have also shown differences of the bone density response to calcium intake according to vitamin D receptor genotype. In one study, the vitamin D receptor heterozygotes responded to calcium intake while the alternate homozygotes either gained or lost bone irrespective of calcium intake⁸⁷. By contrast, in a second study, the BB homozygotes gained some bone when supplemented from a very low basic calcium intake⁸⁸.

However, despite apparent differences in gut calcium absorption, a number of studies have not found any difference in intestinal vitamin D receptor level^{86,89,90}. By contrast, differences in parathyroid gland regulation have been related to vitamin D receptor polymorphisms^{91,92}.

Another potential gene-environment interaction for the vitamin D receptor gene would be in relation to simple vitamin D itself or the active hormonal forms of vitamin D. Differences in response of bone density to the vitamin D metabolites and analogs have been reported according to the vitamin D receptor genotypes, particularly in Japanese studies 93,95 . The more common bb genotype in Japanese cohorts (about 75% of the subjects) was more responsive compared with the heterozygotes, who did not respond well or actually worsened. Given that the heterozygote is the most common genotype in most Caucasian groups, these differences have an intriguing parallel to the differences that have been observed in response to the active vitamin D compounds in clinical studies of osteoporosis between Japanese and Caucasian groups. In another study, the response to simple vitamin D varied according to vitamin D receptor genotype⁹⁶.

The mechanism by which any changes in the vitamin D receptor alleles may account for changes in calcium and bone homeostasis is not clear. At a simple level it is possible that there may be subtle differences in the regulation of the gene or in stability of the mRNA product. Some initial *in vitro* studies suggested that change in stability of mRNA product^{37,97}; however other studies do not confirm this effect⁹⁸⁻¹⁰⁰. Another mechanism may relate to changes in alternative transcripts from the recently reported multiple promoters of single human vitamin D receptor gene¹⁰¹.

The differences in the vitamin D allelic effects may relate to genetic backgrounds and/or environmental factors such as calcium and vitamin D intakes. Genetic backgrounds may relate to other allelic gene effects, e.g., the estrogen receptor genotype ^{102,103}. By this mechanism, allelic effects could differ between environments. Some effects could be quite unexpected as, for example, the apparent protection against some chronic infections reported in a recent African study ¹⁰⁴ and relationship to risk of osteoarthritis of spine and hip ¹⁰⁵⁻¹⁰⁷.

Elucidating pharmacogenomic mechanisms

Currently, most pharmacogenomic studies depend on comparing expression profiles at the mRNA (genomics) or protein (proteomics) level for a given tissue or cell type after a relevant stimulus. Comparison of expression profiles at the mRNA level is attractive, particularly with the advent of recent availability of microarrays allowing concurrent analyses of tens of thousands of genes. This new technology can rapidly genotype individuals to provide information on polymorphic drug metabolism genes, and also identify genes differentially expressed in response to a drug. In fact, one gene chip, CYP2C6/CYP2C19, is already available for identifying potential poor drug metabolizers. On the other hand, this genomics-based technology might also help to understand the biological drug responses and to interpret therapeutic trials¹⁰⁸.

Comparing mRNA expression profiles can be used to explore which genes are up-regulated or down-regulated in osteoporosis treatment by comparing the expression profiles in tissue taken from affected and unaffected individuals. The potential difficulty with this approach is that small variations in the cellular constituents of the tissue might produce large fluctuations in mRNA and/or protein, giving rise to false positive (or negative) results. Another potential problem is that the logistical difficulties of dealing with data on thousands of gene products (which by definition may have no known function) are considerable. These problems can be avoided to some extent by simplifying the experimental design. For example, one approach is to use cultured human bone cells from a single individual and then to compare expression profiles after treatment with, say, bisphosphonates.

Apart from the logistical difficulties in sampling from bone and obtaining comparable bone tissue, study design will also be a major issue. As with genetic linkage studies, a major challenge for pharmacogenomics of osteoporosis lies in the design of meaningful studies for use of these technologies. In any mRNA-level study, a reasonable number of paired replicates must be performed and relevant time points examined. In practice, it may be possible to reduce this to a baseline and two different time points for this kind of experiment. However, even then, with an appropriate number of replicates, the number of samples to be processed and the logistics of multiple samples, at least in humans, remain daunting if not ethically impossible.

On the other hand, large epidemiological studies are required to identify associations between specific gene polymorphisms and predisposition to osteoporosis before these could be useful in clinical settings. At present, large-scale SNP-based association studies in osteoporosis are feasibly prevented by limitations in genotyping resources and biostatistical models. Large-scale association studies involving SNPs will be more practical when high-throughput and affordable SNP scoring methods are available 109. The progress has, nevertheless, been impressive: to date, the Human Genome Project has provided more than two million SNPs as genetic markers¹¹⁰. Within the next few years, SNPs located every 3-50 kb will likely be characterized, it will be possible to perform genome-wide association studies to obtain information about major genes that contribute to the disease or pharmacological differences, as well as secondary, modifier, genes that also affect the disease. The recent development of a single mouthwash method for obtaining genomic DNA clinical studies¹¹¹ may be suitable for large community-based studies in which samples can be collected by the participants themselves.

The advance of genomic research gives rise to several ethical issues that need to be resolved. While information such as race and ethnicity have long been used in predicting therapeutic response, a growing number of critics view the use of this information as potentially prejudicial¹¹². Collecting and storing genetic information from individuals raise questions of privacy as well as security and ethical dilemmas, since the information also provides information about potentially non-censored relatives. Thus, guidelines need to be developed to protect the privacy and confidentiality of participants and their family members. A critical component of any such study will be to ensure that the ethical principle of beneficence is fulfilled. The analysis of DNA samples, including those from large population-based studies, in research is very important for understanding genetic influences of disease susceptibility, but the benefit must be weighed against risk to persons, including the potential for discrimination and invasion of privacy. Although these issues are difficult, it has been suggested that treating participants as limited partners in genetic research can provide a framework for addressing many of these concerns¹¹³.

In summary, data accumulated during the last three decades clearly indicate that genetic factors are a major determinant of bone mineral density, quantitative ultrasound of bone, and bone turnover. Many genetic factors appear to be involved in the determination of BMD as well as bone architecture in various skeletal sites. From the clinical as well as economic points of view, aggressive strategies to search for osteoporosis genes are warranted. With the progress of the Human Genome Project, a new era of post-genomics genotype – phenotype correlations in osteoporosis is heralded. The identification of relevant genes should enhance our understanding not just of disease mechanisms, but also explain why the clinical course of osteoporosis is so variable among individuals. Much more operational research is required to design studies capable of deciphering the complex interactions between individuals' genetic differences, predisposition to the disease, and drug-gene interactions; and the integration and interrogation of the vast data sets that

such studies will produce. While such research effort is undoubtedly complex, the ongoing development of molecular techniques in pharmacogenomics may allow not only individual prediction of drug efficacy and toxicity but also the development of innovative, more active and safer drugs. Genotyping individuals can help determine the influence of the polymorphisms on the pharmacokinetics of the drug on a number of enzymes and transporters that influence the processes of drug absorption and metabolism. Genotyping could also be used to stratify patients for phase III trials, to reduce the necessary sample size. Likewise, genotyping may become part of routine investigations to help clinicians tailor drug therapy effectively. Recent studies demonstrate the feasibility and the importance of these concepts^{108,114}.

The Human Genome Project, coupled with new molecular technologies and new statistical methods, will collectively enhance the search for osteoporosis genes and help translate the prediction of genetically complex osteoporosis into the realm of the possible.

References

- Kanis JA. Diagnosis of osteoporosis and assessment of fracture risk. Lancet 2002; 359:1929-1936.
- Cummings SR, Melton LJ III. Epidemiology and outcomes of osteoporotic fractures. Lancet 2002; 359:1761-1767.
- 3. Center JR, Nguyen TV, Schneider D, Sambrook PN, Eisman JA. Mortality after all major types of osteoporotic fracture in men and women: an observational study. Lancet 1999; 353:878-882.
- Nguyen TV, Sambrook PN, Kelly PJ, Jones G, Lord SR, Freund J, Eisman JA. Prediction of osteoporotic fractures by postural instability and bone density. Br Med J 1993; 307:1111-1115.
- Hui SL, Slemenda CW, Johnton CC. Age and bone mass as predictors of fracture in prospective studies, J Clin Invest 1987; 81:1804-1809.
- Melton LJ III, Atkinson EJ, O'Fallon WM, Wahner HW, Riggs BL. Long-term fracture risk prediction by bone mineral density assessed at different skeletal sites. J Bone Miner Res 1993; 8:1227-1233.
- Marshall D, Johnell O, Wedel H. Meta-analysis of how well measures of bone mineral density predict occurrence of osteoporotic fractures. Br Med J 1996; 312:1254-1259.
- Kannus P, Palvanen M, Kaprio J, Parkkari J, Koskenvuo M. Genetic factors and osteoporotic fractures in elderly people: prospective 25-year follow-up of a nationwide cohort of elderly Finnish twins. Br Med J 1999; 319:1334-1337.
- Deng HW, Chen WM, Recker S, Stegman MR, Li JL, Davies KM, Zhou Y, Deng H, Heaney R, Recker RR. Genetic determination of Colles' fracture and differential bone mass in women with and without Colles' fracture. J Bone Miner Res 2000; 15:1243-1252.

- Cummings SR, Nevitt MC, Browner WS, Stone K, Fox KM, Ensrud KE, Cauley J, Black D, Vogt TM. Risk factors for hip fracture in white women. N Engl J Med 1995; 332:767-773.
- Seeman E, Hopper JL, Bach LA, Cooper ME, Parkinson E, MacKay J, Jerums G. Reduced bone mass in daughters of women with osteoporosis. N Engl J Med 1989; 320:554-558.
- 12. Seeman E, Tsalamandris C, Formica C, Hopper JL, McKay J. Reduced femoral neck bone density in the daughters of women with hip fractures: the roles of low peak bone density in the pathogenesis of osteoporosis. J Bone Miner Res 1994; 9:739-743.
- 13. Evans RA, Marel GM, Lancaster EK, Kos S, Evans M, Wong SYP. Bone mass is low in relatives of osteoporotic patients. Ann Intern Med 1988; 109:870-873.
- 14. Pocock NA, Eisman JA, Hopper JL, Yeates GM, Sambrook PN, Ebert S. Genetic determinants of bone mass in adults: a twin study. J Clin Invest 1987; 80:706-710.
- Young D, Hopper JL, Nowsen CA. Green RM, Sherwin J, Kaymacki B, Smid M, Guest GS, Larkins RG, Wark JD. Determinants of bone mass in 10 to 26-year-old females: a twin study. J Bone Miner Res 1995; 10:558-567.
- Nguyen TV, Howard GM, Kelly PJ, Eisman JA. Bone mass, lean mass and fat mass: same genes or same environments. Am J Epidemiol 1998; 147:3-16.
- 17. Smith DM, Nance WE, Kang KW, Christian JC, and Johnston CC. Genetic factors in determining bone mass. J Clin Invest 1973; 52:2800-2808.
- 18. Flicker L, Hopper JL, Rodgers L, Kaymakci B, Green RM, Wark JD. Bone density determinants in elderly women: a twin study. J Bone Miner Res 1995; 10:1607-1613.
- Tokita A, Kelly, PJ, Nguyen TV, Sambrook PN, Eisman JA. Genetic influences on type I collagen synthesis and degradation: further evidence for genetic regulation of bone turnover. J Clin Endocrinol Metab 1994; 78:1461-1466.
- Harris M, Nguyen TV, Howard GM, Kelly PJ, Eisman JA. Genetic and environmental correlations between bone formation and bone mineral density: a twin study. Bone 1998; 22:141-145.
- Garnero P, Arden NK, Griffiths G, Delmas PD, Spector TD. Genetic influence on bone turnover in postmenopausal twins. J Clin Endocrinol Metab 1996; 81:140-146.
- Howard GM, Nguyen TV, Harris M, Kelly PJ, Eisman JA. Genetic and environmental contributions to the association between quantitative ultrasound and bone mineral density measurements: a twin study. J Bone Miner Res 1998; 13:1318-1327.
- 23. Arden NK, Spector TD. Genetic influences on muscle strength, lean body mass and bone mineral density: a twin study. J Bone Miner Res 1997; 12:2076-2081.
- Danielson ME, Cauley JA, Baker CE, Newman AB, Dorman JS, Towers JD, Kuller LH. Familial resem-

- blance of bone mineral density (BMD) and calcaneal ultrasound attenuation: the BMD in mothers and daughters study. J Bone Miner Res 1999; 14:102-110.
- Dohi Y, Iki M, Ohgushi H, Gojo S, Tabata S, Kajita E, Nishino H, Yonemasu K. A novel polymorphism in the promoter region for the human osteocalcin gene: the possibility of a correlation with bone mineral density in postmenopausal women. J Bone Miner Res 1998; 13:1633-1639.
- 26. Keen RW, Woodford-Richens KL, Lanchburry JS, Spector TD. Allelic variation at the interleukin-1 receptor antagonist gene is associated with early postmenopausal bone loss at the spine. Bone 1998; 23:367-371.
- 27. Cole DEC, Trang HM, Vieth R, Peltekova VD, Pierratos A, Wong BYL, Rubin LA, Hendy GN. Calcium excretion is independently associated with the vitamin D receptor (VDR) and calcium sensing receptor (CASR) polymorphisms in a nephrolithiasis population. Bone 1998; 23:S248.
- Zmuda JM, Eichner JE, Ferrell RE, Bauer DC, Kuller LH, Cauley JA. Genetic variation in α2HS-glycoprotein is related to calcaneal broadband ultrasound attenuation in older women. Calcif Tissue Int 1998; 63:5-8.
- Papiha SS, Francis RM, Allcroft LC, Datta HK. Association of vitamin D receptor and intronic Alu repeats in the vitamin D binding protein alleles with bone mineral density in men. J Bone Miner Res 1996; 11:S207.
- 30. Willing M, Sowers M, Aron D, Clark MK, Burns T, Bunten C, Crutchfield M, D'Agostino D, Jannausch M. Bone mineral density and its change in white women: estrogen and vitamin D receptor genotypes and their interaction. J Bone Miner Res 1998; 13:695-705.
- 31. Kobayashi S, Inoue S, Hosoi T, Ouchi Y, Shiraki M, Orimo H. Association of bone mineral density with polymorphism of the estrogen receptor gene. J Bone Miner Res 1996; 11:306-311.
- 32. Tsuji S, Munkhbat B, Hagihara M, Tsuritani I, Abe H, Tsuji K. HLA-A*24-B*07-DRB1*01 haplotype implicated with genetic disposition of peak bone mass in healthy young Japanese women. Hum Immunol 1998; 59:243-249.
- 33. Murray RE, McGuigan FEA, Grant SFA, Reid DM, Ralston SH. Polymorphisms of the interleukin-6 gene are associated with bone mineral density. Bone 1997; 21:89-92.
- 34. Tsukamoto K, Yoshida H, Watanabe S, Suzuki T, Miyao M, Hosoi T, Orimo H, Emi M. Association of radial bone mineral density with CA repeat polymorphism at the interleukin-6 locus in postmenopausal Japanese women. J Hum Genet 1999; 44:148-151.
- 35. Gong G, Jihnson ML, Barger-Lux MJ, Heany RP, Kimmel DB, Recker RR. Association of PTH gene polymorphism with metacarpal diameter and rate of change in upper radius in women. J Bone Miner Res 1995; 10:S462.

- 36. Kohlmeier M, Saupe J, Schaefer K, Asmus G. Bone fracture history and prospective bone fracture risk of hemodialysis patients are related to apolipoprotein E genotype. Calcif Tissue Int 1998; 62:278-281.
- Morrison NA, Yeoman R, Kelly PJ, Eisman JA. Contribution of trans-acting factor alleles to normal physiological variability: vitamin D receptor gene polymorphisms and circulating osteocalcin. Proc Natl Acad Sci USA 1992; 89:6665-6669.
- 38. Morrison NA, Qi JC, Tokita A, Kelly PJ, Crofts L, Nguyen TV, Sambrook PN, Eisman JA. Prediction of bone density from vitamin D receptor alleles. Nature 1994; 367:284-287.
- 39. Grant SFA, Reid DM, Blake G, Herd R, Fogelman I, Ralston SH. Reduced bone density and osteoporotic vertebral fracture associated with a polymorphic Sp1 binding site in the collagen type Iα1 gene. Nature Genet 1996; 14:203-205.
- 40. Yamada Y, Miyauchi A, Goto J, Takagi Y, Okuizumi H, Kanematsu M, Hase M, Takai H, Harada A, Ikeda K. Association of a polymorphism of the transforming growth factor-b1 gene with genetic susceptibility to osteoporosis in postmenopausal Japanese women. J Bone Miner Res 1998; 13:1569-1576.
- 41. Zmuda JM, Cauley JA, Danielson ME, Wolf RL, Ferrell RE. Vitamin D receptor gene polymorphisms, bone turnover, and rates of bone loss in older African-American women. J Bone Miner Res 1997; 12:1446-1452.
- Jorgensen HL, Scholler J, Sand JC, Bjuring M, Hassager C, Christiansen C. Relation of common allelic variation at vitamin D receptor locus to bone mineral density and postmenopausal bone loss: cross-sectional and longitudinal population study. Br Med J 1996; 313:586-590.
- Taboulet J, Frenkian M, Frendo JL, Feingold N, Julienne A, de Vernejoul MC. Calcitonin receptor polymorphism is associated with a decreased fracture risk in postmenopausal women. Hum Mol Genet 1998; 7:2129-2133.
- 44. Keen RW, Woodford-Richens KL, Lanchburry JS, Spector TD. Allelic variation at the interleukin-1 receptor antagonist gene is associated with early postmenopausal bone loss at the spine. Bone 1998; 23:367-371.
- 45. Cole DEC, Trang HM, Vieth R, Peltekova VD, Pierratos A, Wong BYL, Rubin LA, Hendy GN. Calcium excretion is independently associated with the vitamin D receptor (VDR) and calcium sensing receptor (CASR) polymorphisms in a nephrolithiasis population. Bone 1998; 23:S248.
- Zmuda JM, Eichner JE, Ferrell RE, Bauer DC, Kuller LH, Cauley JA. Genetic variation in α2HS-Glycoprotein is related to calcaneal broadband ultrasound attenuation in older women. Calcif Tissue Int 1998; 63:5-8.
- 47. Papiha SS, Francis RM, Allcroft LC, Datta HK. Association of vitamin D receptor and intronic Alu repeats in the vitamin D binding protein alleles with bone mineral density in men. J Bone Miner Res 1996; 11:S207.

- 48. Gong G, Jihnson ML, Barger-Lux MJ, Heany RP, Kimmel DB, Recker RR. Association of PTH gene polymorphism with metacarpal diameter and rate of change in upper radius in women. J Bone Miner Res 1995; 10:S462.
- 49. Kohlmeier M, Saupe J, Schaefer K, Asmus G. Bone fracture history and prospective bone fracture risk of hemodialysis patients are related to Apolipoprotein E genotype. Calcif Tissue Int 1998; 62:278-281.
- Taboulet J, Frenkian M, Frendo JL, Feingold N, Julienne A, de Vernejoul MC. Calcitonin receptor polymorphism is associated with a decreased fracture risk in postmenopausal women. Hum Mol Genet 1998; 7:2129-2133.
- 51. Gong Y, Vikkula M, Boon L, Liu J, Beighton P, Ramesar R, Peltonen L, Somer H, Hirose T, Dallapiccola B, De Paepe A, Swoboda W, Zabel B, Superti-Furga A, Steinmann B, Brunner HG, Jans A, Boles RG, Adkins W, van den Boogaard MJ, Olsen BR, Warman ML. Osteoporosis-pseudoglioma syndrome, a disorder affecting skeletal strength and vision, is assigned to chromosome region 11q12-13. Am J Hum Genet 1996; 59:146-151.
- 52. Johnson ML, Gong G, Kimberling W, Recker SM, Kimmel DB, Recker RR. Linkage of a gene causing high bone mass to human chromosome 11 (11q12-13). Am J Hum Genet 1997; 60:1326-1332.
- 53. Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, et al.: Osteoporosis-Pseudoglioma Syndrome Collaborative Group. LDL receptor related protein 5 (LRP5) affects bone accrual and eye development. Cell 2001; 107:513-523.
- 54. Boyden LM, Mao J, Belsky J, Mitzner L, Farhi A, Mitnick MA, Wu D, Insogna K, Lifton RP. High bone density due to a mutation in LDL-receptor-related protein 5. N Engl J Med 2002; 346:1513-1521.
- 55. Little RD, Carulli JP, Del Mastro RG, Dupuis J, Osborne M, Folz C, et al. A mutation in the LDL receptor-related protein 5 gene results in the autosomal dominant high-bone-mass trait. Am J Hum Genet 2002; 70:11-19.
- 56. Van Wesenbeeck L, Cleiren E, Gram J, Beals RK, Benichou O, Scopelliti D, et al. Six novel missense mutations in the LDL receptor-related protein 5 (LRP5) gene in different conditions with an increased bone density. Am J Hum Genet 2003; 72:763-767.
- 57. Ferrari SL, Deutsch S, Choudhury U, Chevalley T, Bonjour JP, Dermitzakis ET, Rizzoli R, Antonarakis SE. Polymorphisms in the low-density lipoprotein receptor-related protein 5 (LRP5) gene are associated with variation in vertebral bone mass, vertebral bone size, and stature in whites. Am J Hum Genet 2004; 74:866-875.
- 58. Styrkarsdottir U, Cazier J-B, Kong A, Rolfsson O, Larsen H, Bjarnadottir E, Johannsdottir VD, Sigurdardottir MS, Bagger Y, Christiansen C, Reynisdottir I,

- Grant SF, Jonasson K, Frigge ML, Gulcher JR, Sigurdsson G, Stefansson K. Linkage of osteoporosis to chromosome 20p12 and association with BMP2. PLoS Biol 2003; 1:1-10.
- 59. Delmas PD. Treatment of postmenopausal osteoporosis. Lancet 2002; 359:2018-2026.
- 60. Kalow W. Pharmacogenetics in biological perspective. Pharmacol Rev 1997; 49:369-379.
- Hughes HB, Biehl JP, Jones AP, Schmidt LH. Metabolism of isoniazid in man as related to the occurrence of peripheral neuritis. Am Rev Tuberculosis 1954; 70:266-273.
- 62. Carson PEF, Flanagan CL, Ickes CE, Alving AS. Enzymatic deficiency in primaquine-sensitive erythrocytes. Science 1956; 124:484-485.
- 63. Vesell ES. Advances in pharmacogenetics and pharmacogenomics. Clin Pharmacol 2000; 40:930-938.
- Touw DJ. Clinical implications of genetic polymorphisms and drug interactions mediated by cytochrome P450 enzymes. Drug Metab Drug Interact 1997; 14:55-82.
- 65. Evans WE, Relling MV. Pharmacogenomics: translating functional genomics into rational therapeutics. Science 1999; 286:487-491.
- 66. Evans DAP. Genetic factors in drug therapy: Clinical and Molecular Pharmacogenetics. Cambridge University Press, Cambridge, MA; 1993:303-305.
- 67. Buchert E, Woosley RL. Clinical implications of variable antiarrhythmic drug metabolism. Pharmacogenetics 1992; 2:2-11.
- 68. Dahl AK, Bertilsson L. Genetically variable metabolism of antidepressants and neuroleptic drugs in man. Pharmacogenet 1993; 3:61-70.
- Tanaka E, Hisawa S. Clinically significant pharmacokinetic drug interactions with psychoactive drugs: antidepressants and antipsychotics and the cytochrome P450 system. J Clin Pharm Ther 1999; 24:7-16.
- Morais S, Wilkinson GR, Blaisdell J, Nakamura K, Meyer UA, Goldstein JA. The major genetic defect responsible for the polymorphism of S-mephenytoin metabolism in humans. J Biol Chem 1994; 269:15419-15422.
- 71. Daly AK. Molecular basis of polymorphic drug metabolism. J Mol Med 1995; 73:39-53.
- Gonzales FJ. Pharmacogenetic phenotyping and genotyping. Present status and future potential. Clin Pharmacokinet 1994; 26:59-70.
- Nakamura M, Zhang ZQ, Shan L, Hisa T, Sasaki M, Tsukino R, Yokoi T, Kaname A, Kakudo K. Allelic variants of human calcitonin receptor in the Japanese population. Hum Genet 1997; 99:38-41.
- 74. Palomba S, Orio F, Russo T, Cascella T, Nappi C, Colao A, Nunziata V, Mastratonio P, Lombardi G, Zullo F. BsmI vitamin D receptor genotypes influence the efficacy of anti-resorptive treatments in post-menopausal women. A one-year multicentre randomized controlled trial. Osteoporos Int 2005 Mar 1; [Epub ahead of print].

- Palomba S, Numis FG, Mossetti G, Rendina D, Vuotto P, Russo T, Zullo F, Nappi C, Nunziata V. Effectiveness of alendronate treatment in postmenopausal women with osteoporosis: relationship with BsmI vitamin D receptor genotypes. Clin Endocrinol (Oxf) 2003; 58:365-371.
- Palomba S, Numis FG, Mossetti G, Rendina D, Vuotto P, Russo T, Zullo F, Nappi C, Nunziata V. Raloxifene administration in postmenopausal women with osteoporosis: effect of different BsmI vitamin D receptor genotypes. Hum Reprod 2003; 18:192-198.
- 77. Howard G, Nguyen T, Morrison N, Watanabe T, Sambrook P, Eisman J, Kelly PJ. Genetic influences on bone density: physiological correlates of vitamin D receptor gene alleles in premenopausal women. J Clin Endocrinol Metab 1995; 80:2800-2805. Also see correction in J Clin Endocrinol Metab 1998; 83:1043.
- Ongphiphadhanakul B, Rajatanavin R, Chanprasertyothin S, Chailurkit L, Piaseu N, Teerarungsikul K, Sirisriro R, Komindr S, Puavilai G. Vitamin D receptor gene polymorphism is associated with urinary calcium excretion but not with bone mineral density in postmenopausal women. J Endocrinol Invest 1997; 20:592-596.
- Dawson-Hughes B, Harris SS, Finneran S. Calcium absorption on high and low calcium intakes in relation to vitamin D receptor genotype. J Clin Endocrinol Metab 1995; 80:3657-3661.
- 80. Ames S, Ellis K, Gunn S, Copeland K, Abrams S. Vitamin D receptor gene Fok1 polymorphism predicts calcium absorption and bone mineral density in children. J Bone Miner Res 1999; 14:740-746.
- 81. Kiel DP, Myers RH, Cupples LA, Kong XF, Zhu XH, Ordovas J, Schaefer EJ, Felson DT, Rush D, Wilson PW, Eisman JA, Holick MF. The BsmI vitamin D receptor restriction fragment length polymorphism (bb) influences the effect of calcium intake on bone mineral density. J Bone Miner Res 1997; 12:1049-1057.
- 82. Wishart JM, Horowitz M, Need AG, Scopacasa F, Morris HA, Clifton PM, Nordin BEC. Relations between calcium intake, calcitriol, polymorphisms of the vitamin D receptor gene, and calcium absorption in premenopausal women. Am J Clin Nutr 1997; 65:798-802.
- 83. Francis RM, Harrington F, Turner E, Papiha SS, Datta HK. Vitamin D receptor gene polymorphism in men and its effect on bone density and calcium absorption. Clin Endocrinol (Oxf) 1997; 46:83-86.
- 84. Rauch F, Radermacher A, Danz A, Schiedermaier U, Golucke A, Michalk D, Schönau E. Vitamin D receptor genotypes and changes of bone density in physically active German women with high calcium intake. Exp Clin Endocrinol Diabetes 1997; 105:103-108.
- 85. Gunnes M, Berg JP, Halse J, Lehmann EH. Lack of relationship between vitamin D receptor genotype and forearm bone gain in healthy children, adolescents, and young adults. J Clin Endocrinol Metab 1997; 82:851-855.
- 86. Kinyamu HK, Gallagher JC, Knezetic JA, DeLuca HF,

- Prahl JM, Lanspa SJ. Effect of vitamin D receptor genotypes on calcium absorption, duodenal vitamin D receptor concentration, and serum 1,25 dihydroxyvitamin D levels in normal women. Calcif Tissue Int 1997; 60:491-495.
- 87. Ferrari S, Rizzoli R, Chevalley T, Slosman D, Eisman JA, Bonjour JP. Vitamin-D-receptor-gene polymorphisms and change in lumbar spine bone mineral density [see comments]. Lancet 1995; 345:423-424.
- 88. Krall EA, Parry P, Lichter JB, Dawson-Hughes B. Vitamin D receptor alleles and rates of bone loss: influences of years since menopause and calcium intake. J Bone Miner Res 1995; 10:978-984.
- 89. Barger-Lux MJ, Heaney RP, Hayes J, DeLuca HF, Johnson ML, Gong G. Vitamin D receptor gene polymorphism, bone mass, body size, and vitamin D receptor density. Calcif Tissue Int 1995; 57:161-162.
- 90. Kinyamu HK, Gallagher JC, Prahl JM, DeLuca HF, Petranick KM, Lanspa SJ. Association between intestinal vitamin D receptor, calcium absorption, and serum 1,25 dihydroxyvitamin D in normal young and elderly women. J Bone Miner Res 1997; 12:922-928.
- 91. Carling T, Kindmark A, Hellman P, Lundgren E, Ljunghall S, Rastad J, Akerstrom G, Melhus H. Vitamin D receptor genotypes in primary hyperparathyroidism [see comments]. Nature Med 1995; 1:1309-1311.
- 92. Carling T, Kindmark A, Hellman P, Holmberg L, Akerstrom G, Rastad J. Vitamin D receptor alleles b, a, and T: risk factors for sporadic primary hyperparathyroidism (HPT) but not HPT of uremia or MEN 1. Biochem Biophys Res Comm 1997; 231:329-332.
- 93. Shiraki M, Shiraki Y, Aoki C, Hosoi T, Inoue S, Kaneki M, Ouchi Y. Association of bone mineral density with apolipoprotein E phenotype. J Bone Miner Res 1997; 12:1438-1445.
- 94. Yamagata Z, Miyamura T, Iijima S, Asaka A, Sasaki M, Kato J, Koizumi K. Vitamin D receptor gene polymorphism and bone mineral density in healthy Japanese women [letter]. Lancet 1994; 344:1027.
- 95. Tokita A, Kelly PJ, Nguyen TV, Qi JC, Morrison NA, Risteli L, Risteli J, Sambrook PN, Eisman JA. Genetic influences on type I collagen synthesis and degradation: further evidence for genetic regulation of bone turnover. J Clin Endocrinol Metab 1994; 78:1461-1466.
- 96. Graafmans WC, Lips P, Ooms ME, van Leeuwen JP, Pols HA, Uitterlinden AG. The effect of vitamin D supplementation on the bone mineral density of the femoral neck is associated with vitamin D receptor genotype. J Bone Miner Res 1997; 12:1241-1245.
- 97. Arai H, Miyamoto K, Taketani Y, Yamamoto H, Iemori Y, Morita K, Tonai T, Nishisho T, Mori S, Takeda E. A vitamin D receptor gene polymorphism in the translation initiation codon: effect on protein activity and relation to bone mineral density in Japanese women. J Bone Miner Res 1997; 12:915-921.
- 98. Mocharla H, Butch AW, Pappas AA, Flick JT, Wein-

- stein RS, De Togni P, Jilka RL, Roberson PK, Parfitt AM, Manolagas SC. Quantification of vitamin D receptor mRNA by competitive polymerase chain reaction in PBMC: lack of correspondence with common allelic variants. J Bone Miner Res 1997; 12:726-733.
- Gross C, Musiol IM, Eccleshall TR, Malloy PJ, Feldman D. Vitamin D receptor gene polymorphisms: analysis of ligand binding and hormone responsiveness in cultured skin fibroblasts. Biochem Biophys Res Comm 1998; 242:467-473.
- 100. Durrin LK, Haile RW, Ingles SA, Coetzee GA. Vitamin D receptor 3'-untranslated region polymorphisms: lack of effect on mRNA stability. Biochim Biophys Acta 1999; 1453:311-320.
- 101. Crofts L, Hancock M, Morrison N, Eisman JA. Multiple promoters direct the tissue-specific expression of novel N-terminal variant human vitamin D receptor gene transcripts. Proc Natl Acad Sci (USA) 1998; 95:10529-10534.
- 102. Gennari L, Becherini L, Masi L, Mansani R, Gonnelli S, Cepollaro C, Martini S, Montagnani A, Lentini G, Becorpi AM, Brandi ML. Vitamin D and estrogen receptor allelic variants in Italian postmenopausal women: evidence of multiple gene contribution to bone mineral density. J Clin Endocrinol Metab 1998; 83:939-944.
- 103. Willing M, Sowers M, Aron D, Clark MK, Burns T, Bunten C, Crutchfield M, D'Agostino D, Jannausch M. Bone mineral density and its change in white women: estrogen and vitamin D receptor genotypes and their interaction. J Bone Miner Res 1998; 13:695-705.
- 104. Bellamy R, Ruwende C, Corrah T, McAdam KP, Thursz M, Whittle HC, Hill AV. Tuberculosis and chronic hepatitis B virus infection in Africans and variation in the vitamin D receptor gene. J Infect Dis 1999; 179:721-724.
- 105. Jones G, White C, Sambrook P, Eisman J. Allelic variation in the vitamin D receptor, lifestyle factors and lumbar spinal degenerative disease. Ann Rheum Dis 1998; 57:94-99.
- 106. Uitterlinden AG, Burger H, Huang Q, Odding E, Duijn EM, Hofman A, Birkenhager JC, van Leeuwen JP, Pols HA. Vitamin D receptor genotype is associated with radiographic osteoarthritis at the knee. J Clin Invest 1997; 100:259-263.
- 107. Keen RW, Hart DJ, Lanchbury JS, Spector TD. Association of early osteoarthritis of the knee with a Taq I polymorphism of the vitamin D receptor gene. Arthritis Rheum 1997; 40:1444-1449.
- 108. Joyce DE, Gelbert L, Ciaccia A, DeHoff B, Grinnell BW. Gene expression profile of antithrombotic protein c defines new mechanisms modulating inflammation and apoptosis. J Biol Chem 2001; 276:11199-11203.
- 109. Lai E, Riley J, Purvis I, Roses A. A 4-Mb high density single nucleotide polymorphism-based map around APOE. Genomics 1998; 54:31-38.
- 110. Venter JC, Adams MD, Myers EW, Li PW, Mural RJ,

- Sutton GG, et al. The sequence of the human genome. Science 2001; 291:1304-1351.
- 111. Lum A, Le Marchand L. A simple mouthwash method for obtaining genomic DNA in molecular epidemiological studies. Cancer Epidemiol Biomarkers Prev 1998; 7:719-724.
- 112. Dries DL, Exner DV, Gersh BJ, Cooper HA, Carson PE, Domanski MJ. Racial differences in the outcome of left ventricular dysfunction. N Engl J Med 1999;
- 340:609-616. Also see Erratum in N Engl J Med 1999; 341:298; and discussions in N Engl J Med 1999; 341:287-288.
- 113. Greely HT. Genomics research and human subjects. (Editorial). Science 1998; 282:625.
- 114. Zhao B, Bowden RA, Stavchansky SA, Bowman PD. Human endothelial cell response to Gram-negative lipopolysaccharide assessed with cDNA microarrays. Am J Physiol Cell Physiol 2001; 281:C1587-C1595.