

Predictive and prognostic biomarkers for neoadjuvant chemoradiotherapy in locally advanced rectal cancer

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Accepted 5 May 2015

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Abstract

Locally advanced rectal cancer is regularly treated with trimodality therapy consisting of neoadjuvant chemoradiation, surgery and adjuvant chemotherapy. There is a need for biomarkers to assess treatment response, and aid in stratification of patient risk to adapt and personalise components of the therapy. Currently, pathological stage and tumour regression grade are used to assess response. Experimental markers include proteins involved in cell proliferation, apoptosis, angiogenesis, the epithelial to mesenchymal transition and microsatellite instability. As yet, no single marker is sufficiently robust to have clinical utility. Microarrays that screen a tumour for multiple promising candidate markers, gene expression and microRNA profiling will likely have higher yield and it is expected that a combination or panel of markers would prove most useful. Moving forward, utilising serial samples of circulating tumour cells or circulating nucleic acids can potentially allow us to demonstrate tumour heterogeneity, document mutational changes and subsequently measure treatment response.

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Keywords: Rectal cancer; Neoadjuvant; Biomarker; Chemoradiation

1. Introduction

Colorectal cancer is one of the leading causes of mortality in the developed world, with rectal cancers comprising a third of cases. Rectal cancers carry a worse prognosis and in the locally advanced setting, are treated differently to colon cancers, with trimodality therapy consisting of neoadjuvant chemoradiation, surgery and adjuvant chemotherapy. Neoadjuvant therapy consists of either long-course chemoradiation or short-course radiation. Long-course chemoradiation is given as 45–50.4 Gy radiation in 25–28 fractions, typically over 5–6 weeks, and short-course as 25 Gy in 5 fractions over a week. Long-course therapy is recommended for patients with mid to lower tumours as it provides a higher rate of sphincter preservation due to tumour downstaging [1]. Higher tumours may be treated with short-course radiotherapy. Short-course radiation has been shown to have effective local control and no difference in overall survival compared to long-course chemoradiotherapy [2,3]. Surgery follows neoadjuvant therapy, consisting of proctectomy with total mesorectal excision. Despite trimodality therapy, around 40% of patients still recur [1].

1.1. Current benchmark – tumour regression and downstaging

The clinicopathological variables that are currently used to guide therapy and prognosticate include post-treatment pathological stage (ypT and ypN) and tumour regression grade (TRG) that are assessed microscopically [4,5]. TRG and tumour downstaging have become universally accepted for assessing tumour response to neoadjuvant therapy in locally advanced rectal cancer [4–6]. Hence it is used in the majority of studies as the benchmark for comparison with newer biomarker candidates. TRG and downstaging after

neoadjuvant therapy have resulted in an increased rate of sphincter preservation with curative resection, 9–30% with complete tumour regression and 46–60% with tumour downstaging [4,7]. A survival benefit has also been demonstrated, with pathological complete response associated with superior survival. A recent study reported both complete and intermediate tumour regression to be associated with improved long-term outcomes with a follow-up period of 132 months, based on the Dworak classification [8]. It is important to note that there is no standardised method of reporting regression, with various different grading classifications, based on degree of fibrosis and percentage of viable tumour cells [9–11]. This lack of standardisation perhaps explains the lack of association between tumour regression and survival in a study of 297 patients that looked at both cellular response/regression (based on Mandard and Rodel classifications) and downstaging [12]. Nevertheless, tumour regression is included as a prognostic factor in the 7th edition cancer staging manual of the American Joint Committee on Cancer (AJCC) [13].

Studies have also demonstrated the importance of pathological stage [8,14–16]. A retrospective study of 77 patients correlated the residual pathological stage and survival outcomes, both local and distant [5]. Survival by pathologic stage was reported to be 100% for ypT0–2, N0 cancers, 80% for ypT3–4, N0 and 73% for ypTx, N1–2. Local recurrence of disease was observed in no patients with ypT0–2, N0 as compared with 13% (3/24) in ypT3–4, N0 and 16% (5/31) in ypT0–4, N1–2 patients. Downstaging was shown in multivariate analyses to be significant for survival in the aforementioned study [12].

Other histological factors have also been investigated along with tumour response in a multivariate analysis in 297 patients [7]. This demonstrated that a pathological response of more than 95%, positive lymph nodes, lymphovascular invasion and perineural invasion were significantly

associated with recurrence-free and overall survival. The treatment consisted of long-course radiotherapy with concurrent chemotherapy (93% treated with 5-fluorouracil (5-FU) and the other 7% treated with irinotecan). Interestingly, they showed that there may not be a difference between a pathological response of greater than 95% and pCR. Correlation of clinicopathological factors and tumour characteristics with TRG was explored in another study of 98 patients with rectal cancer treated with chemoradiotherapy [17]. Non-fixed tumours and tumours with circumferential extent less than or equal to 50% were associated with good response. Tumour circumferential extent of greater than 60% was associated with lower pCR in a study of 562 patients treated with concurrent radiotherapy +/- chemotherapy (5-FU in 77% and uracil with tegafur in 2%) [18]. This remained significant in multivariate analysis ($p = 0.033$). Greater tumour circumferential extent ($p = 0.020$) as well as greater distance from anal verge ($p = 0.010$) predicted lower tumour downstaging.

Time to surgery is also becoming a focus of interest, based on the observation that a longer duration from radiation to surgery results in greater tumour regression [19]. One hundred and sixty-seven patients treated with neoadjuvant radiation were assessed for predictive and prognostic markers of tumour downstaging and tumour response [16]. Time before surgery was found to be a predictor of tumour downstaging.

These current factors that determine prognosis are measured on the surgical specimen, and as yet there are no robust biomarkers that can aid in stratification of patient risk in order to adapt and personalise therapy. In studies that have adopted a 'watch-and-wait' approach, a combination of radiologic, clinical and endoscopic criteria is used after completion of neoadjuvant treatment, and intense surveillance is adopted in cases of complete clinical response that did not proceed to surgery [20]. There is a great need to find markers that can predict tumour radiosensitivity and response, particularly upfront or early on in the treatment course. Hence this is particularly relevant in the neoadjuvant phase, given that patients who experience a complete response may not need to proceed with surgery or additional therapy.

Many biomarkers have been investigated and compared against the current standard of TRG. These include tumour tissue biomarkers of either biopsy or surgical specimens, and blood-based biomarkers as well as tumour related nucleic acids and circulating tumour cells. Our review aims at providing a summary of standard and potentially novel clinical and pathological biomarkers after treatment with neoadjuvant chemoradiation. Selected studies on short-course radiation are included to illustrate certain points. For the most part, interesting correlations exist. However, some data are conflicting and there is as yet no single marker that is robust

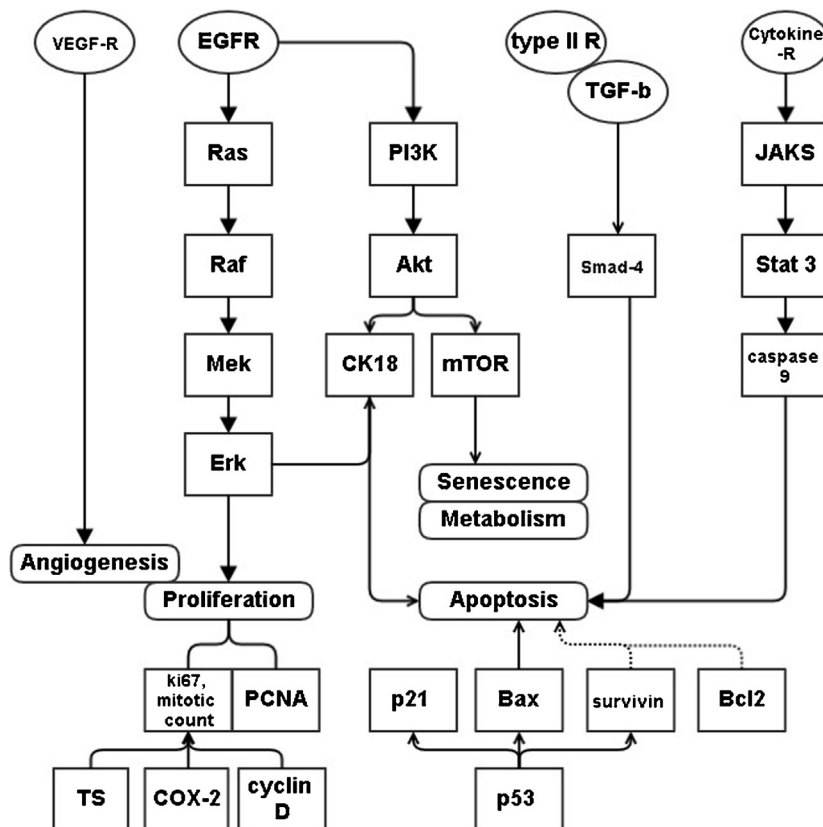


Fig. 1. Pathway modulators that have been investigated as potential biomarkers of response in rectal cancer (dotted lines indicate an opposing effect).

enough to be clinically useful on its own. It is expected that a combination of these markers may be required to have the most clinical utility.

2. Solid tissue-based biomarkers

2.1. Biomarkers under investigation

Potential biomarker candidates have included markers of cell proliferation, apoptosis, and angiogenesis (Fig. 1). Aberrations in these pathways have been implicated in radioresistance. Specific examples include EGFR Sp1-216 G/T polymorphisms, DNA double-strand break repair pathways, and PI3K-Akt hypoxia-related resistance [21–24]. Many studies have investigated a single biomarker, while others have classified biomarkers based on mechanism of action, or included multiple candidates that have shown promise in earlier studies (Table 1). Many of these studies have relatively small patient number, are retrospective in nature and differ in methodologies, making it difficult to draw firm conclusions or carry out cross-study comparisons. Moreover, there is a lag time bias in these studies, as the effect of chemoradiotherapy is continual over time [25]. Some pertinent studies are discussed herein to illustrate certain concepts in an attempt to draw meaningful conclusions.

Biomarker candidates include p53, epidermal growth factor receptor (EGFR), thymidylate synthase, Ki67, p21, bax/bcl-2, COX-2, cyclin D1, survivin, dihydropyrimidine dehydrogenase (DPD), microsatellite instability (MSI) and carcinoembryonic antigen (CEA) [26,27]. Kuremsky et al. suggested that EGFR, thymidylate synthase and p21 have potential utility while Spolverato et al. concluded that there is no single biomarker that is robust, sensitive and specific enough or that has been tested in a rigorous prospective study where treatment factors and response grading are standardised. This is perhaps not surprising given the multiple targets, feedback mechanisms and redundancy of these pathway modulators. Combinations of these biomarkers have yielded more promising results, with a study combining three elements Ki67, thymidylate synthase and Bax to have 83% sensitivity and 84% specificity in predicting tumour response [28].

Differing methodologies also pose challenges in interpreting study findings. Mutations found by gene sequencing of pre-treatment biopsies for p53 were higher in non-responders to neoadjuvant chemoradiation however, results using immunohistochemistry (IHC) were mixed and no conclusions could be drawn [29]. This review also surmised p21 to have reasonable predictive utility, with high levels post-treatment correlating with worse outcomes. Pre-treatment expression of COX-2 and EGFR appeared to correlate with poor response while high spontaneous or intrinsic apoptosis correlated with a good response.

Survivin is thought to be inversely related to the apoptotic index. Pre-treatment survivin expression correlated with tumour downstaging in a study of 54 patients, with high survivin correlating with less downstaging [30]. Bax, on the other hand, is a proapoptotic protein of the Bcl-2 family, and Bax expression was found to be significantly higher in the complete pathological response group compared to partial response [31]. Similarly, Smac is a pro-apoptotic protein, and in a study of 40 patients, high Smac correlated with tumour volume reduction [32].

It is also unclear how these biomarkers are altered by radiation, and not all studies compared pre- and post-treatment samples. Pre- and post-treatment samples were immunostained for p53 and its downstream targets of MDM2 and p21 [33]. p53 did not change with short-course hyperfractionated radiation (2 fractions delivered daily for 5 days), however MDM2 and p21 were moderately induced. The authors attributed the lack of p53 induction to loss of p53 function, which correlated with mutations in the p53 genes. Other studies demonstrated a decrease in Ki67 and p21, in conjunction with an increase in apoptosis [34–37]. A panel of molecular markers, including p53 and p21, DNA mismatch repair proteins (MLH-1 and MSH2), MIB-1, thymidylate synthase, EGFR and VEGF in 91 patients at biopsy and at time of surgery showed baseline over-expression for p53, p21, MLH-1, MSH2, MIB-1, thymidylate synthase, EGFR and VEGF. Of all these biomarkers, only VEGF decreased after treatment [38].

Chemotherapy and radiotherapy may also have differential effects. Preoperative radiation increased COX-2, however the addition of chemotherapy blunted this effect [39]. The fibro-inflammatory reaction increased after addition of chemotherapy to radiation and correlated with T-stage downstaging. Higher thymidylate synthase levels were associated with higher tumour response (as per classification by Zlobec et al. [40]) in patients treated with chemoradiotherapy, but not in the group treated with radiation alone, raising the question of whether the addition of chemotherapy is purely synergistic [41]. Thymidylate synthase plays an important role in pyrimidine nucleotide synthesis and is a target enzyme for 5-FU which is the most common backbone chemotherapy agent in rectal cancer. Low thymidylate synthase levels have been shown to confer a poorer response to adjuvant therapy in colorectal cancer [42].

Biomarker changes early on during the treatment course can potentially classify patients into responders and those who will not derive a benefit from ongoing treatment, allowing stratification of therapy. p21 and apoptosis-positive biopsy specimens obtained 7 days post commencement of chemoradiation correlated with response based on regression, size reduction and T-stage downstaging [43].

Almost every study mentioned thus far has included a marker of proliferation versus apoptosis. Most commonly, Ki67 is the cell proliferation marker however, others include the proliferating-cell nuclear antigen (PCNA) and mitotic counts [37]. There also appears to be a distinction between

Table 1

Investigational biomarkers of response to therapy in locally advanced rectal cancer.

Biomarker	Methodology	Treatment (chemotherapy, duration of radiation)	Results
Ki67, Mcm3, Bax, Bcl-2, ssDNA, Grp78, TS, DPD, CD34, VEGF, nestin and L-type amino-acid transporter [28]	IHC	Long-course chemoradiation, with infusional irinotecan and oral S-1	Pre-treatment high Ki67, Bax, TS and low Grp78 correlated with tumour response (Dworak grades 3 and 4)
COX-2, EGFR, Ki67, p21, survivin, TS and VEGF [30]	IHC	Long-course chemoradiation, with 5-FU or capecitabine	Pre-treatment low survivin correlated with histological tumour downstaging
Bcl-2, Bax, Ki-67, Ku-70 and HDAC1 and mGluR4 [31]	IHC	Long-course chemoradiation, with concurrent 5-FU	Pre-treatment Bax expression correlated with tumour response (Dworak)
Ki67, VEGF and Smac [32]	IHC	Long-course chemoradiation, with 5-FU and oxaliplatin	Smac expression decreased after treatment, high Smac and low Ki67 or VEGF correlated with tumour response (tumour volume measurements)
M30 and Ki67 [34]	IHC	Short-course radiation and non-irradiated cases	M30 expression increased after radiation, but did not correlate with cancer-specific survival. High M30 in non-irradiated cases correlated with improved survival.
p53, p21, apoptotic index, Ki67 [35]	IHC for p53, Ki67 and p21; TUNEL for apoptosis	Radiation 20 Gy in combination with intraoperative radiation 15 Gy	Pre-treatment negative p53 and positive p21 and apoptosis correlated with greater tumour shrinkage (barium enema) and regression (Rich et al.)
COX-2, Ki67 [36]	IHC	Long-course chemoradiation, with 5-FU	COX-2 increased and Ki67 decreased post radiation, no correlation with tumour downstaging
Ki67, PCNA and mitotic counts [37]	IHC and mitoses counted per 10 high-power field	Long course radiation +/- concurrent 5-FU +/- intra-operative radiation	Proliferative activity decreased post radiation, and was associated with more favourable pathological tumour stage, and improved DFS
P53, p21, MLH-1, MSH2, MIB-1, TS, EGFR and VEGF [38]	IHC	Long-course chemoradiation, with 5-FU	Low constitutive p21, absence of post-treatment EGFR were associated with improved DFS and OS; high post-treatment MIB-1 was associated with worse OS
CK18, Ki67 and COX-2, fibro-inflammatory reaction [39]	IHC and scoring of fibro-inflammatory changes as per Shia et al.	Long-course radiation +/- concurrent 5-FU	Addition of chemotherapy to radiation decreased COX-2 and Ki67 expression and increased inflammatory changes, the latter correlated with tumour downstaging
TS, p53, p21, VEGF, MLH1, MSH2 [41]	IHC	38 patients treated with radiation alone (40 Gy), 19 patients treated with radiation (45 Gy) with concurrent 5-FU and oxaliplatin	High TS levels correlated with tumour response in the chemoradiation group
P21, Ki67, apoptosis and p53 [43]	IHC	Long-course chemoradiation, with uracil-tegafur or S-1	p21 and apoptosis-positivity on biopsy specimens obtained a week post treatment correlated with tumour response (JCCC), tumour volume reduction, as assessed by barium enema +/- MRI, and tumour stage downstaging
TP, TS, VEGF, bax, p53, Nfkap B, survivin, COX-2, PCNA, CD44, CD133, MMP-2 and MMP-9 [46]	Reverse transcriptase-PCR	Long-course chemoradiation, with 5-FU	Pre-treatment low CD44 mRNA correlated with tumour regression, PCNA correlated with pathological lymph node negativity
CD133 [47]	IHC	20–45 Gy radiation with concurrent 5-FU and tegafur-uracil	CD133 expression increased from 27.5% to 70% with chemoradiation, histological responders were lower with CD133 expression, no relation to survival
E-cadherin, mir200c, beta-catenin [51]	IHC for E-cadherin and beta-catenin, ISH for mir200c	Long-course chemoradiation, with 5-FU	Reduced E-cadherin and expression of mir200c, as well as increased beta-catenin were associated with non-response. E-cadherin and mir200c associated with inferior cancer-specific survival
MSH2, MLH1, p53, p21, p27, topo II, Ki67 and Bcl-2 [55]	IHC	Long-course chemoradiation, with 5-FU and CPT-11	MSH2 and MLH1 deficiency in biopsy samples, and p21-positivity, correlated with response

Table 1 (Continued)

Biomarker	Methodology	Treatment (chemotherapy, duration of radiation)	Results
MSI [56]	Fluorescent multiplex PCR-based method	Long-course chemoradiation, with 5-FU +/- cisplatin or oxaliplatin	8% (2/25) were MSI-high and did not respond (Dworak) to chemoradiation
COX-2, VEGF, EGFR, Ki67, CAIX and c-CK18, fibro-inflammatory changes [105]	IHC, stromal changes as described by Shia et al.	79 patients received chemoradiation 45 Gy with concurrent 5-FU; 20 patients received 30 Gy radiation	Ki67 and EGFR were downregulated, while c-CK 18 and COX-2 were upregulated with treatment; high VEGF in pre-treatment sample correlated with lower tumour response (Dworak), lower Ki-67 and c-CK18 in post-treatment samples correlated with better regression; fibrosis and COX-2 expression correlated with OS
MT isoforms, MCM3, Ki67 [106]	Real-time PCR for MT, IHC for MCM3 and Ki67	Short-course radiation	mRNA expression for MT increased post radiation, MCM3 expression decreased post radiation
COX-2 [107]	IHC	Long-course chemoradiation, with 5-FU	Pre-treatment COX-2 overexpression correlated with poor tumour response (5-point regression grade)
PLK1 [108]	IHC +/- Affymetrix HG133 microarray	Long-course chemoradiation, with capecitabine and oxaliplatin	Decreased pre-treatment PLK1 expression was associated with tumour regression (Dworak) and decreased tumour recurrence
Hypoxia-related proteins HIF-1 α , CA-IX, VEGF, GLUT-1 [109]	IHC	Long-course chemoradiation, with 5-FU	Increased pre-treatment CA-IX expression correlated with decreased OS
CA-IX [110]	IHC and confirmation by western blotting analysis	Long-course chemoradiation and short-course radiation both included	HIF-1 α , VEGF, GLUT-1 may be co-modulated. Negative or weakly positive tumour CA-IX expression correlated with improved DFS
EGFR, VEGF, HIF- α [111]	Quantitative real-time-PCR	Short-course chemoradiation with 5-FU and tegafur-uracil	Pre-treatment EGFR, VEGF, HIF- α correlated with response (JCCC) EGFR, VEGF levels correlated with improved DFS
GLUT-1, CXCR4, CA9 [112]	IHC	Long-course chemoradiation, with 5-FU	Pre-treatment CA9 expression was significantly lower in responders (Wheeler et al.)

Abbreviations: 5-FU, 5-fluorouracil, CA9, carbonic anhydrase 9, CA-IX, carbonic anhydrase-9, COX, cyclooxygenase, CXCR4, C-X-C chemokine receptor 4, DFS, disease-free survival, DPD, dihydropyrimidine dehydrogenase, EGFR, epidermal growth factor receptor, GLUT-1, glucose transporter-1, HDAC1, histone deacetylase, HIF-1 α , hypoxia-induced factor 1-alpha, IHC, immunohistochemistry, ISH, *in situ* hybridisation, JCCC, Japanese classification of colorectal cancer, mGluR4, metabotropic glutamate receptor 4, MMP, matrix metalloproteinase, MRI, magnetic resonance imaging, MSI, microsatellite instability, MT, metallothionein, OS, overall survival, PCNA, proliferating-cell nuclear antigen, PCR, polymerase chain reaction, PLK1, Human polo-like kinase 1, ssDNA, single-stranded DNA, TP, thymidylate phosphorylase, TS, thymidylate synthase, TUNEL, terminal deoxynucleotidyl transferase dUTP nick end labelling, VEGF, vascular endothelial growth factor.

intrinsic and extrinsic radiation-induced effects, with some reports demonstrating intrinsic apoptosis to be beneficial [29,34,43] while others reported radiation-induced effects correlating with better prognosis [39].

There has been increasing interest in stem cell markers in colorectal cancer, based on the cancer stem-cell theory that a sub-clone of cells has tumorigenic potential and drives disease progression [44,45]. CD44 is a stem cell marker and has been shown to be associated with resistance to radiation. Low CD44 was found to be correlated with tumour regression [46] and remained the only significant factor after multiple logistic regression with odds ratio (OR) 4.7 ($p=0.03$). Prominin-1 or CD133 is another marker thought to originate from cancer stem cells. [47]. CD133 increased after chemoradiation and correlated with lower histological response, however this did not translate into survival differences. These stem cell markers may be useful in determining radioresistance and therapeutic strategies. Another

study found higher CD133+CXCR4+ cells in rectal compared to colon cancer cases, and this correlated with liver metastases and poorer survival [48].

2.2. Biomarkers of emerging interest

2.2.1. The epithelial to mesenchymal transition (EMT)

EMT is a process whereby cancer cells lose their epithelial traits and become more mesenchymal, acquiring invasive capabilities and plasticity [49]. EMT is thought to predict for aggressive disease and metastasis. EMT is recognised as a biological hallmark in the Colorectal Cancer Subtyping Consortium (CRCSC) molecular subtypes of CRC, based on cross comparison of all the different studies inclusive of 4562 samples with collated gene expression data [50]. These were mainly stage I to III cancers, and included colon as well as rectal cancers. Colorectal molecular subtype (CMS) 4 is a tumour cohort defined as possessing mesenchymal

characteristics and carries the worst prognosis. Hence markers of EMT have been explored as biomarkers of response to neoadjuvant therapy. In a rectal cancer study, E-cadherin loss, micro-RNA-200c (mir200c) loss, increased beta-catenin and presence of tumour budding were associated with non-response [51]. Reduced E-cadherin and reduced mir200c were associated with reduced cancer-specific survival. This supports the hypothesis that EMT does play a role in rectal cancer progression and in treatment response.

2.2.2. Microsatellite instability (MSI)

MSI accounts for around 15% of colorectal cancers and around 2–8% of rectal cancers [52,53]. MSI is another hallmark in the CRCSC molecular subtyping [50,54]. MSI-high tumours consist of cells with mismatch repair deficiency that accumulate DNA errors throughout the genome and are associated with a superior outcome. However, its role in treatment response in locally advanced rectal cancer is controversial. Some studies have demonstrated MSI correlating with better response, while others have not [27,38,55]. A study found 8% of rectal cancer samples to be MSI-high and showed no response to chemoradiotherapy [56]. MSH-2 negative cell lines have shown increased death due to chromosomal damage in the S-phase of the cell cycle after ionising radiation [57]. Other studies have shown conflicting results, with baseline MLH-1 positive patients reported to achieve a higher pCR [38]. Microsatellite instability in biopsy specimens correlated with better survival in a study of 57 patients undergoing 5-FU, radiation and CPT-11, a topoisomerase I inhibitor [55].

3. Blood-based biomarkers

3.1. Carcinoembryonic antigen (CEA)

CEA is currently the only biomarker in the National Comprehensive Cancer Network (NCCN) guidelines, included as part of baseline staging and in post treatment surveillance. Studies have consistently demonstrated low pre-operative levels of CEA to be associated with more favourable tumour response, most using 5 ng/ml as the cutoff [17,58–60]. However, this has not been used to stratify treatment strategies as it lacks sensitivity and specificity. Studies that have adopted a non-operative approach post long-course chemoradiation have not included CEA in determining treatment plan [20].

3.2. Tissue-inhibitor of metalloproteinase-1 (TIMP-1)

TIMP-1 is a member of the natural matrix metalloproteinase inhibitor family, shown to predict treatment response in CRC in the metastatic setting [61]. However, data for rectal cancer is sparse. In a study of 33 patients with rectal cancer treated with neoadjuvant chemoradiotherapy (5-FU), post-treatment serum TIMP-1 (ELISA) increased by 13% [62]. A recent study demonstrated that lower post-chemoradiation

TIMP-1 levels to be related to pCR (Mandard classification) [63]. Importantly, TIMP-1 was able to stratify the good clinical responders, as assessed by complete or partial response in radiologic imaging.

3.3. Haematological and inflammatory profiles

There is increasing evidence that the clinical effect of neoadjuvant chemoradiation is dependent on host conditions. A study of 73 patients undergoing long-course therapy found that high haemoglobin levels ($p=0.001$) and low platelets ($p=0.0008$) correlated significantly with tumour size reduction as assessed by barium enema [64]. Response was also high in cases with high lymphocytes and low neutrophils whilst high C-reactive protein and fibrinogen were predictors of poor response. These findings raise the issue of whether optimising the host systemic response may sensitise the tumour to treatment, with interventions such as pre-operative blood transfusions to reverse anaemia.

Another study showed that patients with good response to chemoradiation had higher pre-treatment levels of circulating lymphocytes, higher lymphocyte to total white cell count ratio and lower neutrophil to lymphocyte ratio [65]. Multivariate analysis revealed that lymphocyte to white cell ratio significantly correlated with regression ($p<0.01$) as well as disease free survival and overall survival ($p=0.04$ and 0.02 respectively). Similar trends were found in another study which included patients who received chemoradiation or radiation alone [66]. Ratio of lymphocytes to total white cell count was higher in the pCR group, and ratio of neutrophils lower. They also reported lymphocytes to be reduced significantly during treatment, which may provide an opportunistic microenvironment for tumour cells to proliferate.

Microarray profiling of these peripheral cells may provide further insight into the biological processes underlying treatment response. Pre-treatment peripheral blood mononuclear cells in 35 cases of locally advanced rectal cancers treated with long-course therapy were subjected to microarray gene profiling [67]. Differentially expressed genes were BC 035656.1, CIR, PRDM2, CAPG, FALZ, HLA-DPB2, NUPL2 and ZFP26. The FALZ locus codes transcription factors and overexpression leads to apoptosis, while ZFP36 induces VEGF degradation. FALZ gene expression determined by qRT-PCR (as validation) showed significant differences between responders and non-responders.

3.4. Interleukins and osteopontin

There is increasing focus on the role of inflammation in CRC [68,69]. Serum interleukin-6 (IL-6) is a pro-inflammatory cytokine while osteopontin is a glycoprophosphoprotein that is secreted in the context of malignancy or inflammation. Both were investigated in thirty patients treated with long-course chemoradiation [36]. IL-6 increased while osteopontin was unchanged post-treatment. There was a trend suggesting high osteopontin levels correlating with

development of metastases. Another study examined IL-6 and osteopontin in conjunction with IL-8 [70], and found that only IL-8 was significant in predicting pCR or downstaging after chemoradiation in multivariate analysis. This may be explained by a positive correlation between osteopontin and IL-8. IL-6 polymorphisms have also been investigated, with particular polymorphisms correlating with risk of colon cancer but this was not evident in the rectal cancer cases [71].

3.5. Vascular endothelial growth factor (VEGF)

VEGF can be activated in tumour cells by cytokines including IL-1 and IL-6 as well as hypoxia resulting in secretion of proteolytic enzymes and matrix metalloproteases. The effect of treatment on VEGF levels was investigated in tissue and in blood from 32 patients who received short-course neoadjuvant radiation prior to curative surgery, compared to matched patients who did not receive radiation [72]. VEGF expression by IHC in tissue was significantly increased post-treatment, however circulating VEGF (ELISA) was decreased. The latter was hypothesised to be due to a tumour reduction effect however this has not been studied with respect to tumour recurrence.

4. Tumour DNA, RNA and cells

4.1. MicroRNA (miRNA)

It is estimated that up to 30% of the human genome is regulated by miRNAs. miRNA have been found to be overexpressed in tumour tissue in colorectal cancer [73]. The rectal cancer microRNAome was described by Gaedcke et al. who screened 57 rectal cancer samples and matched controls [74]. Patients were treated with long-course radiation with concurrent 5-FU +/- oxaliplatin. Forty-nine miRNAs were found to be differentially expressed. Rectal-cancer specific miRNA included miR-492, miR-542-5p, miR-584, miR-483-5p, miR144, miR2110, miR-652, miR-375, miR-147b, miR-148a, miR-190, miR-26a/b and miR-338-3p. Another study used 904 miRNA gene chips to analyse expression in normal and cancerous rectal tissue. One hundred and six miRNAs were found to be aberrantly expressed in rectal cancer, of which 45 miRNAs were newly found to be overexpressed, including miR-18a, miR-135b, miR-21, miR-143 and miR-145 [75]. miR-135b had been found to be overexpressed while miR-143 and miR-145 downregulated in colorectal cancer [76–78]. A recent study found miR-21-5p to predict response to chemoradiation with a high accuracy, with an

Table 2
Investigational miRNAs in response to therapy in locally advanced rectal cancer.

Study and reference	Treatment (chemotherapy, duration of radiation)	Results
38 fresh rectal biopsy samples [113]	Neoadjuvant chemotherapy (oxaliplatin and 5-FU) and radiotherapy	14 miRNAs were differentially expressed in TRG1 patients compared to TRG > 1 (Mandard); 13 were confirmed by qRT-PCR Upregulated – miR-1183, miR-483-5p, miR-622, miR-125a-3p, miR-1224-5p, miR-188-5p, miR-1471, miR-671-5p, miR-1909, miR-630, miR-765 Down-regulated – miR-1274b, miR720 miR-622 and miR-630 had 100% sensitivity and 100% specificity
12 formalin-fixed paraffin embedded biopsy samples [114]	Unknown	3 miRNA transcripts miR-16, miR-590-5p and miR-153 predicted complete versus incomplete response (Dworak). 2 transcripts miR-519c-3p and miR-561 predicted good versus poor response with a median accuracy of 100%
20 fresh rectal biopsy samples [115]	Neoadjuvant chemotherapy (capecitabine or infusional 5-FU) and radiotherapy	miR-215, miR190b and miR-29b-2 were overexpressed in non-responders while let-7e, miR-196b, miR-450a, miR-450b-5-p and miR-99a were overexpressed in responders (Mandard)
85 formalin-fixed paraffin embedded post-treatment samples [116]	Neoadjuvant chemotherapy (capecitabine) and radiotherapy	miR-21, miR-99, miR-125b, miR-125b1, let-7c and miR-490 upregulation correlated with TRG 1-2 (Mandard). miR-21 and miR-125a-3p downregulation correlated with TRG 4
40 formalin-fixed paraffin embedded biopsy and post-treatment samples [117]	Neoadjuvant chemotherapy (5-FU) and radiotherapy	miR-21 was downregulated while miR-143 and miR-145 were upregulated with treatment. miR- 145 correlated with tumour regression (Schneider et al.)
103 formalin-fixed paraffin embedded post-treatment samples [51]	Neoadjuvant chemotherapy (capecitabine or infusional 5-FU) and radiotherapy	miR-200c loss was associated with non-response (Mandard)
27 biopsy specimens in training set and 17 in validation set [79]	Neoadjuvant chemotherapy (5-FU based) and radiotherapy	miR-21-5p was overexpressed in complete responders, with sensitivity and specificity of 78% and 86% respectively

Abbreviations: 5-FU, 5-fluorouracil, miR, micro RNA, qRT-PCR, quantitative real-time reverse transcription polymerase chain reaction, TRG, tumour regression grade.

AUC (area under curve) of 0.94 [79]. Of note, they validated their findings in an independent group of patients. miRNA has been reported to be important in determination of radiosensitivity of rectal cancers to chemoradiotherapy. In vitro studies demonstrated the correlation between miRNA and radiosensitivity [80]. Transfection of let-7g, miR-224, miR-320a and miR-332 in colorectal cancer cell lines induced radiation sensitivity. Let-7g expression also appeared to correlate with better survival. Table 2 shows the miRNA candidates that have been investigated in response to treatment in locally advanced rectal cancer.

The advantage of miRNA expression profiling is the large number of candidate markers that can be obtained at any one time. Targeting these miRNA with oligonucleotides represents a potentially useful therapy [81,82]. However, the challenge lies in the interpretation of large amounts of data, given that most studies have little, if any, overlap in dysregulated markers.

4.2. Global gene expression

Microarrays screening a large array of genes have been increasingly utilised to develop a profile that will predict response [82–85]. Using tissue microarrays in 23 patients treated with chemoradiation, different expression levels were obtained for 54 genes with a sensitivity of 78% and specificity of 86% in responder versus non-responder (assessed by T downstaging) groups with locally advanced rectal cancer [84]. Lower levels of calmin, kinectin 1, copine III, villin-like, motilin, cdc42, mypsin IA, cyclin T1, IL12A, SMC1 and platelet-derived growth factor were found in the responders. In the responders assessed by tumour regression, four genes were differentially expressed including REGL, ACVR2B, SMARCC1 and ZNF134. Of note, many of the biomarkers mentioned previously such as p53, p21, p27, bcl2, Ki-67, MSH2, survivin, thymidylate synthase and PCNA were not differentially expressed. Another study also confirmed a gene set predicting for response with an accuracy of 87%. Thymidylate synthase and RAD23 were highly expressed in cases that had a complete response [86].

Genes that were able to discriminate between responders and non-responders with an accuracy of 82.4%, using the human U95Av2 Gene Chip, included genes involved in apoptosis [87]. Lumican, thrombospondin 2 and galectin-1, which are apoptosis inducers, were higher in responders.

4.3. KRAS mutational status and chromosomal alterations

Chromosomal copy number alterations may be associated with tumour response. Complete pathological response was associated with significantly fewer high copy gains and loss of 12p13.31 was associated with pCR [88]. KRAS mutations (codons 12, 13, 61) were reported to correlate with improved response to neoadjuvant chemoradiation in a small study [89]. Other studies investigating KRAS mutational status found

no correlation with outcomes [90,91]. However, differences were found between the G12V and G13D mutations, with the former associated with greater regression.

4.4. Circulating tumour nucleic acid (ctNA)

ctNA has been used as a surrogate for circulating tumour cells (CTCs), however it is not possible to definitively identify the cell of origin of ctNA [92]. Candidate nucleic acid biomarkers are chosen based on their assumed derivation from cancer cells, and as markers of metastatic potential. Levels of hTERT and ctRNA were found to be higher pre-chemoradiation than post-treatment in responders [93]. Another small study of 26 patients investigated CK20 and CEA ctNA, with a positive rate (defined as both biomarkers being positive) of 32% pre-treatment [94]. Sixty-three percent of responders were positive pre-treatment while 18% of non-responders were positive. There was a significant decrease post-treatment in responders. Another study found that CK20 ctRNA correlated with inferior survival [95]. The utility of ctDNA measured by real-time PCR as a therapy monitoring marker has also been explored [96]. Serial sampling revealed that the median ctDNA was 4.2 ng/ml before chemoradiation, 1.0 ng/ml at the end of chemoradiation and 4.1 ng/ml postoperatively. There was a significant difference in postoperative ctDNA, with responders (ypT0-2) having 2.2 ng/ml and non-responders (ypT3-4) 5.1 ng/ml.

4.5. Circulating tumour cells (CTCs)

There is increasing evidence for the clinical utility of CTCs in colorectal cancer, with most studies confirming high CTC counts correlating with poorer prognosis [97]. CTCs are thought to be released from the primary tumour into the bloodstream and considered the initiating seed for metastasis at distant organ sites [98]. However, the majority of these studies do not focus on rectal cancers as a separate entity. Rectal cancers receiving neoadjuvant chemoradiotherapy comprised a small subgroup (12 patients) of one study, which enumerated CTCs based on EpCAM (epithelial cell adhesion marker) positivity using flow cytometry [99]. No differences were found in CTC counts between the colonic and rectal cancers, and no effect was found with neoadjuvant chemoradiotherapy. However, the small numbers and the isolation of CTCs based on EpCAM alone, may have affected the results, due to inability to capture cells that have undergone EMT and hence display more mesenchymal surface marker characteristics [100].

5. Constitutional patient factors

There is increasing interest in the potential for treatment-associated toxicities or symptom burden to function as a biomarker for treatment response or tumour burden. In a study of 54 patients, lower levels of fatigue at baseline and

at completion of chemoradiation were found to be significant predictors of pathological response, as gauged by tumour downstaging [101].

Obesity in rectal cancer has also been reported to be associated with inferior or similar survival outcomes [102]. The measure of visceral to subcutaneous fat areas was thought to be a more accurate indicator of abdominal or visceral adiposity and quantitative measures of visceral adiposity were correlated with clinicopathological variables as well as survival outcomes in a retrospective study of patients treated with chemoradiation [103]. Patients who had higher visceral fat had less T-stage downstaging while patients with no or partial response and high adiposity comprised the worst group in terms of disease-free survival.

6. Conclusion

The approach of selecting and validating biomarkers relies on pre-existing knowledge of those same candidate biomarkers. The drawbacks of this approach include the limited pool of candidate biomarkers, given the known multiplicity and redundancy of carcinogenic pathways. More recently, microarrays and miRNA expression profiling have allowed the analyses of hundreds of potential biomarkers and selection of promising candidates [104]. This will have even greater potential if applied to circulating tumour cells or nucleic acids.

With regard to sampling time-point strategies, biomarkers measured post-treatment may be useful to decide on subsequent therapy, but the information it affords in addition to the current standard as defined on the surgical specimen, is uncertain. Baseline pre-treatment biomarkers will likely have higher clinical utility as they aid in the initial treatment decision making, prior to obtaining the surgical sample.

It is clear that currently a single biomarker will not be robust enough to have prognostic and predictive utility. Models that combine blood biomarkers, functional imaging with positron emission tomography (PET) and clinical data have shown an AUC of 0.81 for pCR, and 0.78 for tumour response based on downstaging. This is superior to the AUC of the individual markers [70]. With this information, nomograms can be constructed to incorporate all potentially useful biomarkers that on their own are not robust enough to have clinical relevance. Further, microarrays that can interrogate a tumour for multiple promising candidate markers at any one time can increase this yield. Moving forward, a non-static sample such as circulating tumour cells or circulating nucleic acids, has the potential to provide insight into tumour heterogeneity and treatment related mutational changes.

Conflict of interest

None declared.

References

- [1] Sauer R, Liersch T, Merkel S, Fietkau R, Hohenberger W, Hess C, et al. Preoperative versus postoperative chemoradiotherapy for locally advanced rectal cancer: results of the German CAO/ARO/AIO-94 randomized phase III trial after a median follow-up of 11 years. *J Clin Oncol* 2012;30(16):1926–33.
- [2] Ngan SY, Burmeister B, Fisher RJ, Solomon M, Goldstein D, Joseph D, et al. Randomized trial of short-course radiotherapy versus long-course chemoradiation comparing rates of local recurrence in patients with T3 rectal cancer: Trans-Tasman Radiation Oncology Group trial 01.04. *J Clin Oncol* 2012;30(31):3827–33.
- [3] Bujko K, Nowacki MP, Nasierowska-Guttmejer A, Michalski W, Bebenek M, Kryj M. Long-term results of a randomized trial comparing preoperative short-course radiotherapy with preoperative conventionally fractionated chemoradiation for rectal cancer. *Br J Surg* 2006;93(10):1215–23.
- [4] Park JJ, You YN, Agarwal A, Skibber JM, Rodriguez-Bigas MA, Eng C, et al. Neoadjuvant treatment response as an early response indicator for patients with rectal cancer. *J Clin Oncol* 2012;30(15):1770–6.
- [5] Mohiuddin M, Hayne M, Regine WF, Hanna N, Hagihara PF, McGrath P, et al. Prognostic significance of postchemoradiation stage following preoperative chemotherapy and radiation for advanced/recurrent rectal cancers. *Int J Radiat Oncol Biol Phys* 2000;48(4):1075–80.
- [6] Rodel C, Martus P, Papadopoulos T, Fuzesi L, Klimpfinger M, Fietkau R, et al. Prognostic significance of tumor regression after preoperative chemoradiotherapy for rectal cancer. *J Clin Oncol* 2005;23(34):8688–96.
- [7] Guillem JG, Chessin DB, Cohen AM, Shia J, Mazumdar M, Enker W, et al. Long-term oncologic outcome following preoperative combined modality therapy and total mesorectal excision of locally advanced rectal cancer. *Ann Surg* 2005;241(5):829–36, discussion 836–8.
- [8] Fokas E, Liersch T, Fietkau R, Hohenberger W, Beissbarth T, Hess C, et al. Tumor regression grading after preoperative chemoradiotherapy for locally advanced rectal carcinoma revisited: updated results of the CAO/ARO/AIO-94 trial. *J Clin Oncol* 2014;32(15):1554–62.
- [9] Dworak O, Keilholz L, Hoffmann A. Pathological features of rectal cancer after preoperative radiochemotherapy. *Int J Colorectal Dis* 1997;12(1):19–23.
- [10] Mandard AM, Dalibard F, Mandard JC, Marnay J, Henry-Amar M, Petiot JF, et al. Pathologic assessment of tumor regression after preoperative chemoradiotherapy of esophageal carcinoma. Clinicopathologic correlations. *Cancer* 1994;73(11):2680–6.
- [11] Ryan R, Gibbons D, Hyland JM, Treanor D, White A, Mulcahy HE, et al. Pathological response following long-course neoadjuvant chemoradiotherapy for locally advanced rectal cancer. *Histopathology* 2005;47(2):141–6.
- [12] Rullier A, Laurent C, Capdepon M, Vendrely V, Bioulac-Sage P, Rullier E. Impact of tumor response on survival after radiochemotherapy in locally advanced rectal carcinoma. *Am J Surg Pathol* 2010;34(4):562–8.
- [13] Edge SB, Compton CC. The American Joint Committee on Cancer: the 7th edition of the AJCC cancer staging manual and the future of TNM. *Ann Surg Oncol* 2010;17(6):1471–4.
- [14] Capirci C, Rampin L, Erba PA, Galeotti F, Crepaldi G, Banti E, et al. Sequential FDG-PET/CT reliably predicts response of locally advanced rectal cancer to neo-adjuvant chemo-radiation therapy. *Eur J Nucl Med Mol Imaging* 2007;34(10):1583–93.

- [15] Kim YH, Kim DY, Kim TH, Jung KH, Chang HJ, Jeong SY, et al. Usefulness of magnetic resonance volumetric evaluation in predicting response to preoperative concurrent chemoradiotherapy in patients with resectable rectal cancer. *Int J Radiat Oncol Biol Phys* 2005;62(3):761–8.
- [16] Berger C, de Muret A, Garaud P, Chapet S, Bourlier P, Reynaud-Bougnoix A, et al. Preoperative radiotherapy (RT) for rectal cancer: predictive factors of tumor downstaging and residual tumor cell density (RTCD): prognostic implications. *Int J Radiat Oncol Biol Phys* 1997;37(3):619–27.
- [17] Yan H, Wang R, Zhu K, Zhao W, Jiang S, Feng R, et al. Predictors of sensitivity to preoperative chemoradiotherapy of rectal adenocarcinoma. *Tumori* 2011;97(6):717–23.
- [18] Das P, Skibber JM, Rodriguez-Bigas MA, Feig BW, Chang GJ, Wolff RA, et al. Predictors of tumor response and downstaging in patients who receive preoperative chemoradiation for rectal cancer. *Cancer* 2007;109(9):1750–5.
- [19] Garcia-Aguilar J, Smith DD, Avila K, Bergsland EK, Chu P, Krieg RM, et al. Optimal timing of surgery after chemoradiation for advanced rectal cancer: preliminary results of a multicenter, nonrandomized phase II prospective trial. *Ann Surg* 2011;254(1):97–102.
- [20] Habr-Gama A, Perez RO, Nadalin W, Sabbaga J, Ribeiro Jr U, Silva e Sousa Jr AH, et al. Operative versus nonoperative treatment for stage 0 distal rectal cancer following chemoradiation therapy: long-term results. *Ann Surg* 2004;240(4):711–7, discussion 717–8.
- [21] Gantt GA, Chen Y, Dejulus K, Mace AG, Barnholtz-Sloan J, Kalady MF. Gene expression profile is associated with chemoradiation resistance in rectal cancer. *Colorectal Dis* 2014;16(1):57–66.
- [22] Ramzan Z, Nassri AB, Huerta S. Genotypic characteristics of resistant tumors to pre-operative ionizing radiation in rectal cancer. *World J Gastrointest Oncol* 2014;6(7):194–210.
- [23] Spindler KL, Nielsen JN, Lindebjerg J, Brandslund I, Jakobsen A. Prediction of response to chemoradiation in rectal cancer by a gene polymorphism in the epidermal growth factor receptor promoter region. *Int J Radiat Oncol Biol Phys* 2006;66(2):500–4.
- [24] Stegeman H, Span PN, Kaanders JH, Bussink J. Improving chemoradiation efficacy by PI3-K/AKT inhibition. *Cancer Treat Rev* 2014;40(10):1182–91.
- [25] Glimelius B, Gronberg H, Jarhult J, Wallgren A, Cavallin-Stahl E. A systematic overview of radiation therapy effects in rectal cancer. *Acta Oncol* 2003;42(5/6):476–92.
- [26] Kuremsky JG, Tepper JE, McLeod HL. Biomarkers for response to neoadjuvant chemoradiation for rectal cancer. *Int J Radiat Oncol Biol Phys* 2009;74(3):673–88.
- [27] Spolverato G, Pucciarelli S, Bertorelle R, De Rossi A, Nitti D. Predictive factors of the response of rectal cancer to neoadjuvant radiochemotherapy. *Cancers (Basel)* 2011;3(2):2176–94.
- [28] Kikuchi M, Mikami T, Sato T, Tokuyama W, Araki K, Watanabe M, et al. High Ki67, Bax, and thymidylate synthase expression well correlates with response to chemoradiation therapy in locally advanced rectal cancers: proposal of a logistic model for prediction. *Br J Cancer* 2009;101(1):116–23.
- [29] Smith FM, Reynolds JV, Miller N, Stephens RB, Kennedy MJ. Pathological and molecular predictors of the response of rectal cancer to neoadjuvant radiochemotherapy. *Eur J Surg Oncol* 2006;32(1):55–64.
- [30] Kim K, Chie EK, Wu HG, Kim SG, Lee SH, Kang GH, et al. High survivin expression as a predictor of poor response to preoperative chemoradiotherapy in locally advanced rectal cancer. *Int J Colorectal Dis* 2011;26(8):1019–23.
- [31] Chang HJ, Jung KH, Kim DY, Jeong SY, Choi HS, Kim YH, et al. Bax: a predictive marker for therapeutic response to preoperative chemoradiotherapy in patients with rectal carcinoma. *Hum Pathol* 2005;36(4):364–71.
- [32] Yan H, Wang R, Yu J, Jiang S, Zhu K, Mu D, et al. Predictive value of Smac: VEGF and Ki-67 in rectal cancer treated with neoadjuvant therapy. *Oncol Lett* 2010;1(4):641–7.
- [33] Stift A, Prager G, Selzer E, Widder J, Kandioler D, Friedl J, et al. The early response of p53-dependent proteins during radiotherapy in human rectal carcinoma and in adjacent normal tissue. *Int J Oncol* 2003;23(5):1269–75.
- [34] Marijnen CA, Nagtegaal ID, Mulder-Stapel AA, Schrier PI, van de Velde CJ, van Krieken JH, et al. High intrinsic apoptosis: but not radiation-induced apoptosis, predicts better survival in rectal carcinoma patients. *Int J Radiat Oncol Biol Phys* 2003;57(2):434–43.
- [35] Suzuki T, Sadahiro S, Fukasawa M, Ishikawa K, Kamijo A, Yasuda S, et al. Predictive factors of tumor shrinkage and histological regression in patients who received preoperative radiotherapy for rectal cancer. *Jpn J Clin Oncol* 2004;34(12):740–6.
- [36] Debuquoy A, Goethals L, Geboes K, Roels S, Mc Bride WH, Haustermans K. Molecular responses of rectal cancer to preoperative chemoradiation. *Radiother Oncol* 2006;80(2):172–7.
- [37] Willett CG, Warland G, Hagan MP, Daly WJ, Coen J, Shellito PC, et al. Tumor proliferation in rectal cancer following preoperative irradiation. *J Clin Oncol* 1995;13(6):1417–24.
- [38] Bertolini F, Bengala C, Losi L, Pagano M, Iachetta F, Dealis C, et al. Prognostic and predictive value of baseline and posttreatment molecular marker expression in locally advanced rectal cancer treated with neoadjuvant chemoradiotherapy. *Int J Radiat Oncol Biol Phys* 2007;68(5):1455–61.
- [39] Debuquoy A, Libbrecht L, Roobrouck V, Goethals L, McBride W, Haustermans K. Morphological features and molecular markers in rectal cancer from 95 patients included in the European Organisation for Research and Treatment of Cancer 22921 trial: prognostic value and effects of preoperative radio (chemo) therapy. *Eur J Cancer* 2008;44(6):791–7.
- [40] Zlobec I, Steele R, Nigam N, Compton CC. A predictive model of rectal tumor response to preoperative radiotherapy using classification and regression tree methods. *Clin Cancer Res* 2005;11(15):5440–3.
- [41] Negri FV, Campanini N, Camisa R, Pucci F, Bui S, Cecon G, et al. Biological predictive factors in rectal cancer treated with preoperative radiotherapy or radiochemotherapy. *Br J Cancer* 2008;98(1):143–7.
- [42] Edler D, Glimelius B, Hallstrom M, Jakobsen A, Johnston PG, Magnusson I, et al. Thymidylate synthase expression in colorectal cancer: a prognostic and predictive marker of benefit from adjuvant fluorouracil-based chemotherapy. *J Clin Oncol* 2002;20(7):1721–8.
- [43] Suzuki T, Sadahiro S, Tanaka A, Okada K, Kamata H, Kamijo A, et al. Biopsy specimens obtained 7 days after starting chemoradiotherapy (CRT) provide reliable predictors of response to CRT for rectal cancer. *Int J Radiat Oncol Biol Phys* 2013;85(5):1232–8.
- [44] Dalerba P, Dylla SJ, Park IK, Liu R, Wang X, Cho RW, et al. Phenotypic characterization of human colorectal cancer stem cells. *Proc Natl Acad Sci U S A* 2007;104(24):10158–63.
- [45] Jordan CT, Guzman ML, Noble M. Cancer stem cells. *N Engl J Med* 2006;355(12):1253–61.
- [46] Huh JW, Lee JH, Kim HR. Pretreatment expression of 13 molecular markers as a predictor of tumor responses after neoadjuvant chemoradiation in rectal cancer. *Ann Surg* 2014;259(3 (Mar)):508–15.
- [47] Saigusa S, Tanaka K, Toiyama Y, Yokoe T, Okugawa Y, Koike Y, et al. Clinical significance of CD133 and hypoxia inducible factor-1alpha gene expression in rectal cancer after preoperative chemoradiotherapy. *Clin Oncol (R Coll Radiol)* 2011;23(5):323–32.
- [48] Zhang SS, Han ZP, Jing YY, Tao SF, Li TJ, Wang H, et al. CD133(+)CXCR4(+) colon cancer cells exhibit metastatic potential and predict poor prognosis of patients. *BMC Med* 2012;10:85.
- [49] Nieto MA, Cano A. The epithelial–mesenchymal transition under control: global programs to regulate epithelial plasticity. *Semin Cancer Biol* 2012;22(5/6):361–8.
- [50] Dienstmann R, Guinney J, Delorenzi M, De Reynies A, Roepman P, Sadanandam A. Colorectal cancer subtyping consortium (CRCSC)

- identification of a consensus of molecular subtypes. *J Clin Oncol* 2014;32(5s). p. suppl; abstr 3511.
- [51] Bhangu A, Wood G, Brown G, Darzi A, Tekkis P, Goldin R. The role of epithelial mesenchymal transition and resistance to neoadjuvant therapy in locally advanced rectal cancer. *Colorectal Dis* 2014;16(4):O133–43.
 - [52] Ishikubo T, Nishimura Y, Yamaguchi K, Khansuwan U, Arai Y, Kobayashi T, et al. The clinical features of rectal cancers with high-frequency microsatellite instability (MSI-H) in Japanese males. *Cancer Lett* 2004;216(1):55–62.
 - [53] Shin JS, Tut TG, Yang T, Lee CS. Radiotherapy response in microsatellite instability related rectal cancer. *Korean J Pathol* 2013;47(1):1–8.
 - [54] Roepman P, Schlicker A, Tabernero J, Majewski I, Tian S, Moreno V, et al. Colorectal cancer intrinsic subtypes predict chemotherapy benefit: deficient mismatch repair and epithelial-to-mesenchymal transition. *Int J Cancer* 2014;134(3):552–62.
 - [55] Charara M, Edmonston TB, Burkholder S, Walters R, Anne P, Mitchell E, et al. Microsatellite status and cell cycle associated markers in rectal cancer patients undergoing a combined regimen of 5-FU and CPT-11 chemotherapy and radiotherapy. *Anticancer Res* 2004;24(5B):3161–7.
 - [56] Demes M, Scheil-Bertram S, Bartsch H, Fisseler-Eckhoff A. Signature of microsatellite instability: KRAS and BRAF gene mutations in German patients with locally advanced rectal adenocarcinoma before and after neoadjuvant 5-FU radiochemotherapy. *J Gastrointest Oncol* 2013;4(2):182–92.
 - [57] Franchitto A, Pichierri P, Piergentili R, Crescenzi M, Bignami M, Palitti F. The mammalian mismatch repair protein MSH2 is required for correct MRE11 and RAD51 relocalization and for efficient cell cycle arrest induced by ionizing radiation in G2 phase. *Oncogene* 2003;22(14):2110–20.
 - [58] Park YA, Sohn SK, Seong J, Baik SH, Lee KY, Kim NK, et al. Serum CEA as a predictor for the response to preoperative chemoradiation in rectal cancer. *J Surg Oncol* 2006;93(2):145–50.
 - [59] Moureau-Zabotto L, Farnault B, de Chaisemartin C, Esterni B, Lelong B, Viret F, et al. Predictive factors of tumor response after neoadjuvant chemoradiation for locally advanced rectal cancer. *Int J Radiat Oncol Biol Phys* 2011;80(2):483–91.
 - [60] Tural D, Selcukbiricik F, Dztferk MA, Yildiz D, Turna H, Erdamar S, et al. The relation between pathologic complete response and clinical outcome in patients with rectal cancer. *Hepatogastroenterology* 2013;60(128).
 - [61] Sorensen NM, Bystrom P, Christensen IJ, Berglund A, Nielsen HJ, Brunner N, et al. TIMP-1 is significantly associated with objective response and survival in metastatic colorectal cancer patients receiving combination of irinotecan: 5-fluorouracil, and folinic acid. *Clin Cancer Res* 2007;13(14):4117–22.
 - [62] Aldulaymi B, Christensen IJ, Soletormos G, Jess P, Nielsen SE, Laurberg S, et al. Chemoradiation-induced changes in serum CEA and plasma TIMP-1 in patients with locally advanced rectal cancer. *Anticancer Res* 2010;30(11):4755–9.
 - [63] Yoon HI, Koom WS, Kim YB, Min BS, Lee KY, Kim NK, et al. Predicting the pathologic response of locally advanced rectal cancer to neoadjuvant concurrent chemoradiation using enzyme-linked immunosorbent assays (ELISAs) for biomarkers. *J Cancer Res Clin Oncol* 2014;140(3):399–409.
 - [64] Yasuda K, Sunami E, Kawai K, Nagawa H, Kitayama J. Laboratory blood data have a significant impact on tumor response and outcome in preoperative chemoradiotherapy for advanced rectal cancer. *J Gastrointest Cancer* 2012;43(2):236–43.
 - [65] Dou X, Wang RB, Yan HJ, Jiang SM, Meng XJ, Zhu KL, et al. Circulating lymphocytes as predictors of sensitivity to preoperative chemoradiotherapy in rectal cancer cases. *Asian Pac J Cancer Prev* 2013;14(6):3881–5.
 - [66] Kitayama J, Yasuda K, Kawai K, Sunami E, Nagawa H. Circulating lymphocyte is an important determinant of the effectiveness of preoperative radiotherapy in advanced rectal cancer. *BMC Cancer* 2011;11:64.
 - [67] Palma P, Cuadros M, Conde-Muino R, Olmedo C, Cano C, Segura-Jimenez I, et al. Microarray profiling of mononuclear peripheral blood cells identifies novel candidate genes related to chemoradiation response in rectal cancer. *PLOS ONE* 2013;8(9):e74034.
 - [68] Galon J, Mlecnik B, Bindea G, Angell HK, Berger A, Lagorce C, et al. Towards the introduction of the 'Immunoscore' in the classification of malignant tumours. *J Pathol* 2014;232(2):199–209.
 - [69] Galon J, Pages F, Marincola FM, Angell HK, Thurin M, Lugli A, et al. Cancer classification using the Immunoscore: a worldwide task force. *J Transl Med* 2012;10:205.
 - [70] Buijsen J, van Stiphout RG, Menheere PP, Lammering G, Lambin P. Blood biomarkers are helpful in the prediction of response to chemoradiation in rectal cancer: A prospective, hypothesis driven study on patients with locally advanced rectal cancer. *Radiother Oncol*; 2014.
 - [71] Slattery ML, Wolff RK, Herrick JS, Caan BJ, Potter JD. IL6 genotypes and colon and rectal cancer. *Cancer Causes Control* 2007;18(10):1095–105.
 - [72] Inoue Y, Ojima E, Watanabe H, Hiro J, Toiyama Y, Kobayashi M, et al. Does preoperative chemo-radiotherapy enhance the expression of vascular endothelial growth factor in patients with rectal cancer? *Oncol Rep* 2007;18(2):369–75.
 - [73] Motoyama K, Inoue H, Takatsuno Y, Tanaka F, Mimori K, Uetake H, et al. Over- and under-expressed microRNAs in human colorectal cancer. *Int J Oncol* 2009;34(4):1069–75.
 - [74] Gaedcke J, Grade M, Camps J, Sokilde R, Kaczowski B, Schetter AJ, et al. The rectal cancer microRNAome – microRNA expression in rectal cancer and matched normal mucosa. *Clin Cancer Res* 2012;18(18):4919–30.
 - [75] Li X, Zhang G, Luo F, Ruan J, Huang D, Feng D, et al. Identification of aberrantly expressed miRNAs in rectal cancer. *Oncol Rep* 2012;28(1):77–84.
 - [76] Arndt GM, Dossey L, Cullen LM, Lai A, Druker R, Eisbacher M, et al. Characterization of global microRNA expression reveals oncogenic potential of miR-145 in metastatic colorectal cancer. *BMC Cancer* 2009;9:374.
 - [77] Ng EK, Chong WW, Jin H, Lam EK, Shin VY, Yu J, et al. Differential expression of microRNAs in plasma of patients with colorectal cancer: a potential marker for colorectal cancer screening. *Gut* 2009;58(10):1375–81.
 - [78] Michael MZ, O'Connor SM, van Holst Pellekaan NG, Young GP, James RJ. Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol Cancer Res* 2003;1(12):882–91.
 - [79] Lopes-Ramos C, Habr-Gama A, Quevedo B, Felicio N, Bettoni F, Koyama F, et al. Overexpression of miR-21-5p as a predictive marker for complete tumor regression to neoadjuvant chemoradiotherapy in rectal cancer patients. *BMC Med Genomics* 2014;7(1):68.
 - [80] Salendo J, Spitzner M, Kramer F, Zhang X, Jo P, Wolff HA, et al. Identification of a microRNA expression signature for chemoradiosensitivity of colorectal cancer cells: involving miRNAs-320a, -224, -132 and let7g. *Radiother Oncol* 2013;108(3):451–7.
 - [81] Valeri N, Braconi C, Gasparini P, Murgia C, Lampis A, Paulus-Hock V, et al. MicroRNA-135b promotes cancer progression by acting as a downstream effector of oncogenic pathways in colon cancer. *Cancer Cell* 2014;25(4):469–83.
 - [82] McDermott AM, Heneghan HM, Miller N, Kerin MJ. The therapeutic potential of microRNAs: disease modulators and drug targets. *Pharm Res* 2011;28(12):3016–29.
 - [83] Garcia-Aguilar J, Chen Z, Smith DD, Li W, Madoff RD, Cataldo P, et al. Identification of a biomarker profile associated with resistance to neoadjuvant chemoradiation therapy in rectal cancer. *Ann Surg* 2011;254(3):486–92, discussion 492–3.
 - [84] Ghadimi BM, Grade M, Difilippantonio MJ, Varma S, Simon R, Montagna C, et al. Effectiveness of gene expression profiling for response

- prediction of rectal adenocarcinomas to preoperative chemoradiotherapy. *J Clin Oncol* 2005;23(9):1826–38.
- [85] Nishioka MSM, Kurita N, Iwata T, Morimoto S, Yoshikawa K, Higashijima J, Miyatani T. Gene expression profile can predict pathological response to preoperative chemoradiotherapy in rectal cancer. *Cancer Genomics Proteomics* 2011;8:87–92.
- [86] Kim IJ, Lim SB, Kang HC, Chang HJ, Ahn SA, Park HW, et al. Microarray gene expression profiling for predicting complete response to preoperative chemoradiotherapy in patients with advanced rectal cancer. *Dis Colon Rectum* 2007;50(9):1342–53.
- [87] Watanabe T, Komuro Y, Kiyomatsu T, Kanazawa T, Kazama Y, Tanaka J, et al. Prediction of sensitivity of rectal cancer cells in response to preoperative radiotherapy by DNA microarray analysis of gene expression profiles. *Cancer Res* 2006;66(7):3370–4.
- [88] Chen Z, Liu Z, Li W, Qu K, Deng X, Varma MG, et al. Chromosomal copy number alterations are associated with tumor response to chemoradiation in locally advanced rectal cancer. *Genes Chromosomes Cancer* 2011;50(9):689–99.
- [89] Luna-Perez P, Segura J, Alvarado I, Labastida S, Santiago-Payan H, Quintero A. Specific c-K-ras gene mutations as a tumor-response marker in locally advanced rectal cancer treated with preoperative chemoradiotherapy. *Ann Surg Oncol* 2000;7(10):727–31.
- [90] Gaedcke J, Grade M, Jung K, Schirmer M, Jo P, Obermeyer C, et al. KRAS and BRAF mutations in patients with rectal cancer treated with preoperative chemoradiotherapy. *Radiother Oncol* 2010;94(1):76–81.
- [91] Zauber NP, Marotta SP, Berman E, Grann A, Rao M, Komati N, et al. Molecular genetic changes associated with colorectal carcinogenesis are not prognostic for tumor regression following preoperative chemoradiation of rectal carcinoma. *Int J Radiat Oncol Biol Phys* 2009;74(2):472–6.
- [92] Schwarzenbach H, Hoon DS, Pantel K. Cell-free nucleic acids as biomarkers in cancer patients. *Nat Rev Cancer* 2011;11(6):426–37.
- [93] Pucciarelli S, Rampazzo E, Briarava M, Maretto I, Agostini M, Digito M, et al. Telomere-specific reverse transcriptase (hTERT) and cell-free RNA in plasma as predictors of pathologic tumor response in rectal cancer patients receiving neoadjuvant chemoradiotherapy. *Ann Surg Oncol* 2012;19(9):3089–96.
- [94] Zitt M, Muller HM, Dinnewitzer AJ, Schwendinger V, Goebel G, De Vries A, et al. Disseminated tumor cells in peripheral blood: a novel marker for therapy response in locally advanced rectal cancer patients undergoing preoperative chemoradiation. *Dis Colon Rectum* 2006;49(10):1484–91.
- [95] Kienle P, Koch M, Autschbach F, Benner A, Treiber M, Wannemacher M, et al. Decreased detection rate of disseminated tumor cells of rectal cancer patients after preoperative chemoradiation: a first step towards a molecular surrogate marker for neoadjuvant treatment in colorectal cancer. *Ann Surg* 2003;238(3):324–30, discussion 330–1.
- [96] Zitt M, Muller HM, Rochel M, Schwendinger V, Goebel G, Devries A, et al. Circulating cell-free DNA in plasma of locally advanced rectal cancer patients undergoing preoperative chemoradiation: a potential diagnostic tool for therapy monitoring. *Dis Markers* 2008;25(3):159–65.
- [97] Lim SH, Becker TM, Chua W, Caixeiro NJ, Ng WL, Kienle N, et al. Circulating tumour cells and circulating free nucleic acid as prognostic and predictive biomarkers in colorectal cancer. *Cancer Lett* 2014;346(1):24–33.
- [98] Caixeiro NJ, Kienle N, Lim SH, Spring KJ, Tognola A, Scott KF, et al. Circulating tumour cells—a bona fide cause of metastatic cancer. *Cancer Metastasis Rev* 2014.
- [99] Galizia G, Gemei M, Orditura M, Romano C, Zamboli A, Castellano P, et al. Postoperative detection of circulating tumor cells predicts tumor recurrence in colorectal cancer patients. *J Gastrointest Surg* 2013;17(10):1809–18.
- [100] Lim SH, Becker TM, Chua W, Ng WL, de Souza P, Spring KJ. Circulating tumour cells and the epithelial mesenchymal transition in colorectal cancer. *J Clin Pathol*; 2014.
- [101] Park HC, Janjan NA, Mendoza TR, Lin EH, Vadhan-Raj S, Hundal M, et al. Temporal patterns of fatigue predict pathologic response in patients treated with preoperative chemoradiation therapy for rectal cancer. *Int J Radiat Oncol Biol Phys* 2009;75(3):775–81.
- [102] Ballian N, Yamane B, Levenson G, Harms B, Heise CP, Foley EF, et al. Body mass index does not affect postoperative morbidity and oncologic outcomes of total mesorectal excision for rectal adenocarcinoma. *Ann Surg Oncol* 2010;17(6):1606–13.
- [103] Clark W, Siegel EM, Chen YA, Zhao X, Parsons CM, Hernandez JM, et al. Quantitative measures of visceral adiposity and body mass index in predicting rectal cancer outcomes after neoadjuvant chemoradiation. *J Am Coll Surg* 2013;216(6):1070–81.
- [104] Milgrom S, Garcia-Aguilar J. Molecular biomarkers as predictors of response to neoadjuvant chemoradiation therapy in rectal cancer. *Semin Colon Rectal Surg* 2013;24:119–24.
- [105] Debuquoy A, Goethals L, Libbrecht L, Perneel C, Geboes K, Ectors N, et al. Molecular and clinico-pathological markers in rectal cancer: a tissue micro-array study. *Int J Colorectal Dis* 2009;24(2):129–38.
- [106] Szelachowska J, Dziegiel P, Tarkowski R, Gomulkiewicz A, Bebenek M, Halon A, et al. Therapeutic radiation induces different changes in expression profiles of metallothionein (MT) mRNA: MT protein, Ki 67 and minichromosome maintenance protein 3 in human rectal adenocarcinoma. *Anticancer Res* 2012;32(12):5291–7.
- [107] Smith FM, Reynolds JV, Kay EW, Crotty P, Murphy JO, Hollywood D, et al. COX-2 overexpression in pretreatment biopsies predicts response of rectal cancers to neoadjuvant radiochemotherapy. *Int J Radiat Oncol Biol Phys* 2006;64(2):466–72.
- [108] Rodel F, Keppner S, Capalbo G, Bashary R, Kaufmann M, Rodel C, et al. Polo-like kinase 1 as predictive marker and therapeutic target for radiotherapy in rectal cancer. *Am J Pathol* 2010;177(2):918–29.
- [109] Lee-Kong SA, Ruby JA, Chessin DB, Pucciarelli S, Shia J, Riedel ER, et al. Hypoxia-related proteins in patients with rectal cancer undergoing neoadjuvant combined modality therapy. *Dis Colon Rectum* 2012;55(9):990–5.
- [110] Korkeila E, Talvinen K, Jaakkola PM, Minn H, Syrjänen K, Sundström J, et al. Expression of carbonic anhydrase IX suggests poor outcome in rectal cancer. *Br J Cancer* 2009;100(6):874–80.
- [111] Toiyama Y, Inoue Y, Saigusa S, Okugawa Y, Yokoe T, Tanaka K, et al. Gene expression profiles of epidermal growth factor receptor: vascular endothelial growth factor and hypoxia-inducible factor-1 with special reference to local responsiveness to neoadjuvant chemoradiotherapy and disease recurrence after rectal cancer surgery. *Clin Oncol (R Coll Radiol)* 2010;22(4):272–80.
- [112] Guedj N, Bretagnol F, Rautou PE, Deschamps L, Cazals-Hatem D, Bedossa P, et al. Predictors of tumor response after preoperative chemoradiotherapy for rectal adenocarcinomas. *Hum Pathol* 2011;42(11):1702–9.
- [113] Della Vittoria Scarpato G, Falchetta F, Carlmagno C, Ubezio P, Marchini S, De Stefano A, et al. A specific miRNA signature correlates with complete pathological response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer. *Int J Radiat Oncol Biol Phys* 2012;83(4):1113–9.
- [114] Kheirlesei EA, Miller N, Chang KH, Curran C, Hennessey E, Sheehan M, et al. miRNA expressions in rectal cancer as predictors of response to neoadjuvant chemoradiation therapy. *Int J Colorectal Dis* 2013;28(2):247–60.
- [115] Svoboda M, Sana J, Fabian P, Kocakova I, Gombosova J, Nekvindova J, et al. MicroRNA expression profile associated with response

to neoadjuvant chemoradiotherapy in locally advanced rectal cancer patients. *Radiat Oncol* 2012;7:195.

- [116] Eva Bandres A, Guerrero D, Lopez I, Gonzalez-Huarriz M, Gomez Dorronsoro ML, Montes M, Monzon F, Torrea N, Armendariz P, Balen E, Viudez A, Asin G, Chicata V, Hernandez I, Eito C, Garcia-Foncillas J. Association between a specific miRNA signature and pathological response to neoadjuvant chemoradiotherapy (CRT) in locally advanced rectal cancer (LARC) patients. In: ASCO. 2012.
- [117] Drebbler U, Lay M, Wedemeyer I, Vallbohmer D, Bollschweiler E, Brabender J, et al. Altered levels of the onco-microRNA 21 and the tumor-suppressor microRNAs 143 and 145 in advanced rectal cancer indicate successful neoadjuvant chemoradiotherapy. *Int J Oncol* 2011;39(2):409–15.

Biography

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