

REVIEW ARTICLES

Etiologic, Environmental and Inherited Risk Factors in Sarcomas

DAVID M. THOMAS, MD, PhD^{1,2,3*} AND MANDY L. BALLINGER, PhD^{2,3**}¹The Kinghorn Cancer Centre, Garvan Institute of Medical Research, Darlinghurst, New South Wales, Australia²Research Division, Peter MacCallum Cancer Centre, East Melbourne, Victoria, Australia³Sir Peter MacCallum Department of Oncology, University of Melbourne, Parkville, Australia

Sarcomas are a rare group of mesenchymal tumors affecting a younger population. The etiology remains unknown in most cases. Environmental factors that increase sarcoma risk include radiation exposure and chemical carcinogens. Several familial cancer syndromes confer sarcoma predisposition, such as the Li-Fraumeni Syndrome (LFS). In this increasingly genomic focussed era of medicine, it will be clinically important to understand the genetic basis of sarcoma risk.

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INTRODUCTION

Sarcomas are a complex group of rare malignant connective tissue tumors, comprised of over 50 subtypes, each of which is genetically and pathologically distinct. Although precise estimates are difficult to obtain, the annual incidence of all sarcomas approximates 30/million population [1]. Importantly, sarcomas disproportionately affect a young population comprising 20% of childhood cancers, and 10% of cancers in adolescents and young adults. In particular, bone sarcomas (Ewing sarcoma and osteosarcoma) are essentially diseases of the young, while the median age of onset of soft-tissue sarcomas is somewhat older (65 years [1]). Early age of onset is a predictor of inherited or constitutive cancer risk, suggesting the importance of genetic factors in sarcoma development in general.

There is evidence for ethnic variation in the spectrum of connective tissue tumors, with Ewing sarcoma reportedly rare in African American populations [2], and giant cell tumor of bone being more common in Asians [3]. A recent survey of racial and ethnic differences in the incidence of sarcoma subtypes amongst adolescents and young adults in the United States found that Hispanics had higher incidences of liposarcoma, African Americans had a higher incidence of fibromatous sarcomas, while Caucasians had a higher incidence of synovial sarcoma [4].

There are several issues potentially limiting sarcoma data collection and analysis. Accurate population-level ascertainment of case numbers is not always easily available, and pathological subtypes are frequently misreported. Cancer incidence is often reported by topography rather than morphology leading to an under representation of sarcoma incidence in some subtypes. For example, uterine leiomyosarcoma and breast angiosarcomas are frequently coded as gynecologic and breast cancers, respectively. Additionally, some estimates suggest that 20% of sarcoma diagnoses are revised substantially on central expert review [5,6]. Assuming each subtype has a distinct genetic and environmental etiology, the rarity of individual subtypes further exacerbates challenges in identifying causal associations. For this reason, and because the classifications of sarcoma have substantially evolved over the past 20 years, there is a tendency to pool sarcoma subtypes in retrospective epidemiologic studies. Whilst this achieves statistical power, it probably results in loss of signal for subtype-specific risk factors.

On the other hand, sarcomas are sufficiently rare that their incidence is notable clinically, a feature that has paradoxically led to the mapping of many dominant Mendelian cancer syndromes, including the Retinoblastoma and Li-Fraumeni syndromes (LFS), as well as environmental risk factors such as radiation, carcinogens, and viruses. Benign connective tissue tumors are estimated to be 100 times more common than sarcomas. A subset of benign tumors, including peripheral nerve sheath tumors, giant cell tumor of bone, and some cartilage tumors, undergo malignant degeneration to true sarcoma. Moreover, distinguishing between benign and malignant tumors can be challenging because the histologic appearances can be ambiguous, and some connective tissue tumors may have “malignant” behavior while being considered “benign.” For example, giant cell tumor of bone may metastasise to the lung—a characteristic usually considered diagnostic of malignancy. Finally, some connective tissue tumors have a truly “intermediate” phenotype exemplified by aggressive fibromatosis. Benign connective tissue tumors form part of the spectrum of tumors in inherited cancer syndromes.

We describe here the known heritable, environmental, and other genetic risk factors associated with sarcomas (Table I) and propose that in this era of genomic cancer medicine that understanding the genetic basis for cancer risk will be increasingly important clinically.

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*Correspondence to: David Thomas, MD, PhD, The Kinghorn Cancer Centre, Garvan Institute of Medical Research, 370 Victoria Street, Darlinghurst, New South Wales 2010, Australia. Fax: +612 9355 5872. E-mail: d.thomas@garvan.org.au

**Correspondence to: Mandy Ballinger, PhD, Research Division, Peter MacCallum Cancer Centre, St. Andrews Place, East Melbourne 3002, Victoria, Australia. E-mail: mandy.ballinger@petermac.org

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TABLE I. Risk Factors for Sarcoma Subtypes

| Sarcoma | Risk factor | Heritable (H) Environ (E) | Gene | Quality of evidence |
|----------------------------|----------------------------------|---------------------------|---------------------|---------------------|
| | | Other (O) | | |
| Chondrosarcoma | Multiple osteochondromas | H | <i>EXT1</i> | High |
| | Maffuci syndrome | O | | High |
| | Ollier disease | O | <i>PTHRI</i> | High |
| Embryonal rhabdomyosarcoma | Beckwith–Wiedemann syndrome | H | <i>CDKN1C, IGF2</i> | High |
| GIST | Familial GIST | H | <i>KIT, PDGFRA</i> | High |
| Hepatic angiosarcoma | Anabolic steroids | E | — | Very low |
| | Arsenic | E | — | Low |
| | Thorotrast | E | — | Low |
| | Vinyl chloride | E | — | High |
| Kaposi sarcoma | KSHV (HHV8) | E | — | High |
| KS and leiomyosarcoma | Immunosuppression | O | — | Medium |
| | Organ transplant | O | — | Low |
| Leiomyosarcoma | Epstein–Barr virus | E | — | Very low |
| Lymphangiosarcoma | Lymphedema | O | — | High |
| MPNST | Neurofibromatosis type 1 | H | <i>NF1</i> | High |
| Malignant rhabdoid tumors | Rhabdoid predisposition syndrome | H | <i>SMARCB1</i> | High |
| Myogenic sarcomas | Rubinstein–Taybi syndrome | H | <i>CREBBP</i> | High |
| Osteosarcoma and STS | Li Fraumeni syndrome | H | <i>TP53</i> | High |
| | Retinoblastoma | H | <i>RB</i> | High |
| | Chlorophenols | E | — | Very low |
| | Ionising radiation | E | — | High |
| | Phenoxyherbicides | E | — | Very low |
| | Trauma | O | — | Very low |
| | Bloom syndrome | H | <i>BLM</i> | High |
| | Familial Paget disease of bone | H | <i>TNFRSF11A</i> | High |
| | Rothmund–Thomson syndrome | H | <i>RECQL4</i> | High |
| | SV-40 | E | — | Very low |
| Osteosarcoma | Paget disease of bone | O | — | High |
| | Mazabraud syndrome | O | <i>GNAS1</i> | High |
| | McCune–Albright syndrome | O | <i>GNAS1</i> | High |
| | Neurofibromatosis type 2 | H | <i>NF2</i> | High |
| | Werner syndrome | H | <i>WRN</i> | High |
| Schwannomas | | | | |
| STS and bone tumors | | | | |

GIST, gastrointestinal stromal tumor; MPNST, malignant peripheral nerve sheath tumor; STS, soft tissue sarcomas; KS, Kaposi sarcoma

ENVIRONMENTAL RISK FACTORS

The etiology of the majority of sarcomas is unknown, but include carcinogens, viruses, as well as ionising radiation. The strongest evidence for an environmental cause relates to ionising radiation. Bone-homing radio-isotopes, such as strontium-89, cause osteosarcoma in animals [7–9]. Long-term occupational exposure to radiation increased the incidence of bone tumors [10,11]. Atomic bomb survivors have an increased incidence of many cancers including sarcomas [12]. Patients who have been treated with radiotherapy show an increased incidence of in-field soft-tissue and bone sarcomas [13,14].

The Childhood Cancer Survivor Study found the risk of secondary sarcoma was ninefold higher among this group compared to the general population and was strongly associated with radiation therapy [15]. Chemotherapy, including anthracyclines or alkylating agents also increased the risk of secondary sarcomas in these childhood survivors by ~twofold [15]. Interactions between environmental and inherited factors appear important. Some familial cancer syndromes may be particularly susceptible to radiation-induced tumorigenesis [16–19].

Animal studies suggest that various chemicals or drugs may cause sarcomas. In mice, methylcholanthrene reproducibly induces fibrosarcomas [20], while therapeutic administration of parathyroid hormone is associated with osteosarcoma in Fischer 344 rats [21,22]. Importantly, animals fundamentally differ from man, and this association has not been confirmed in man [22]. Riddelline, o-nitrotoluene, and benzophenone exposures induce sarcomas in rats and mice [23,24]. A study of occupational risk factors in Europe found significantly increased risk for bone sarcoma in woodworkers, bricklayers, blacksmiths, and

toolmakers [25]. Exposure to pesticide, insecticides, and herbicides was also shown to slightly increase risk although details of chemical constituents were not included. Herbicides and pesticides containing phenoxyacetic acids may be associated with increased sarcoma rates [26,27], both contaminated [28] and free of dioxins [29], although this association has not been confirmed by other studies [30,31]. Studies of the contributions of chlorophenols and dioxins have been inconsistent [32–35]. Vinyl chloride, arsenic, anabolic steroids, and thorium may be moderately strongly associated with an increased risk of hepatic angiosarcomas [36–42]. Water fluoridation is probably not associated with an increased risk of osteosarcoma [43].

Animal studies show that viruses can cause sarcomas. The *Src* oncogene was first isolated from the Rous sarcoma virus [44], and there are many other examples. There is little evidence that viruses are associated with human sarcomas, with the exception of Kaposi sarcoma and Kaposi sarcoma-associated herpesvirus (KSHV), also known as HHV8 [45]. Kaposi sarcoma is a tumor of lymphatic endothelial cells, rather than true mesenchyme, and KSHV sequences were first isolated from AIDS-associated Kaposi sarcoma [46,47]. Perhaps most data exist for SV40 DNA sequences identified in human osteosarcoma [48,49]. In AIDS cases, leiomyosarcomas associated with Epstein–Barr virus have been reported [50,51] and have also been described in transplant recipients [52,53]. Overall, however, the evidence supporting a significant etiologic role for viruses in sarcoma development is weak.

Other risk factors for sarcoma have been reported, with varying levels of evidence. Stewart and Treves first described lymphedema, as a factor in the development of lymphangiosarcoma [54]. Whether prior trauma predisposes to sarcomas is unproven. Patients undergoing arthroplasties

have not shown an elevated incidence of sarcoma [55,56]. In elite athletes, an increased incidence of sarcoma was observed with the suggested contributory factor being prior sporting injury [57]. An early study linked high body mass index with increased sarcoma risk [58], and both diabetes and obesity are associated with an increased risk of uterine sarcomas [59]. Osteosarcoma is markedly more common during the adolescent growth spurt, suggesting a strong effect of pubertal bone growth [60].

HERITABLE RISK FACTORS

To date the heritable aspects of sarcoma have been studied to a lesser degree than other cancers with most genetic research focused on breast, bowel, and ovarian cancers, where 22–35% of the etiology is thought to be heritable [61]. Sarcomas are more common in certain familial cancer syndromes, including LFS, Retinoblastoma, Gardner's and Werner's syndromes, NF1, and some immunodeficiency conditions [62]. A third of pediatric sarcomas are associated with a significant incidence of cancer in relatives [63,64], while sarcomas are sixfold more common in relatives of children with sarcoma than age-matched controls [65]. A population-based study found that one in five sarcoma survivors developed a second cancer within 10 years, and the risk of a second sarcoma was increased 18- to 30-fold [66], with a 5.3-fold increased risk of sarcomas in parents of patients with leiomyosarcoma [67]. A recent Swiss study found that excess secondary sarcoma risk was observed after skin melanoma, breast cancer, uterine cancer, testicular cancer, thyroid cancer, Hodgkin lymphoma, and leukemias [68]. Taken together, these data strongly support a genetic component to sarcomas in particular.

KNOWN SARCOMA PREDISPOSITION SYNDROMES

LFS is a rare, dominant Mendelian cancer syndrome characterized by early onset of a variety of tumor types, most commonly sarcomas (both soft-tissue and bone), early-onset breast cancer, brain tumors, leukemia, and adrenocortical carcinoma [69,70]. The majority of mutations identified to date in families with LFS involve the *TP53* gene. Mutation carriers within LFS families are reported to have a 50% chance of developing cancer by the age of 30 years [71]. Approximately 2–4% of individuals affected by sarcoma will carry mutations in *TP53*, regardless of family history [72].

The IARC database now catalogues over 20,000 reported mutations and variants in *TP53*, which comprise over 350 pathogenic types [73]. Most mutations arise in the DNA-binding domains, but more subtle consequences may arise from variants located outside these critical domains. Specific mutations in *TP53* are associated with different penetrance and cancer types [74]. An interesting specific case is the relatively high frequency of the p.R337H mutation in Southern Brazil, where it is associated with increased risks of breast cancer and adrenocortical cancer [75].

Carriers of mutations in *TP53* experience an estimated second cancer risk of 57% at 30 years after a first cancer diagnosis [18,76], up to 83 times higher than the population at large. Two recent studies suggest that screening may be useful in early cancer detection in LFS patients, including children [77,78]. Breast screening with MRI is now recommended from 25 years for female *TP53* mutation carriers [79]. Avoidance of unnecessary ionising radiation in mutation carriers is important if possible without compromising care. Minimising unnecessary computed tomographic or X-ray surveillance is also sensible [80].

Fully 30% of LFS and up to 80% of Li Fraumeni like kindreds are due to hereditary events not involving *TP53* itself [73]. *CHK2* variant 1100delC has been reported in 2/44 families with LFS/LFL, but interestingly these families were considered atypical because of lack of sarcomas [81], and the role of *CHK2* is controversial [82,83].

The Retinoblastoma syndrome (RB) is a rare autosomal dominant familial cancer syndrome. RB is associated with the childhood tumor, retinoblastoma. Survival rates following retinoblastoma are excellent (90% at 10 years [17]). In survivors of childhood retinoblastoma, the second most commonly reported cancer type is osteosarcoma, with a standardized incidence ratio of 360 compared to the population at large [84,85]. There is also an increased risk of soft-tissue sarcomas, with a cumulative risk of 13.1% at 50 years follow up. Both chemotherapy and radiotherapy exposure are associated with an increased risk [85].

There are two forms of neurofibromatosis, NF1 and NF2. NF1 is a relatively common autosomal dominant disease, occurring in up to 1/2500 live births [86]. Clinical manifestations include café au lait pigmentation, Lisch nodules, neurofibromas, optic pathway gliomas, and bony dysplasias in the context of a family history [86]. The neurofibromas are prone to malignant degeneration, with a 1/10 lifetime chance of developing malignant peripheral nerve sheath tumors [87]. Overall, patients with NF1 have a 34-fold increased risk of dying with a malignant connective tissue of soft-tissue neoplasm, exacerbated by exposure to radiotherapy [19,88]. Malignant transformation is associated with a poor prognosis, and close surveillance is recommended to facilitate effective surgical treatment of these tumors [89].

NF2 is less common, affecting 1/25,000 live births [86]. The incidence of clinical disease is less common, because some individuals do not develop symptoms until adulthood, and many die in childhood. The clinical features of NF2 include a family history, bilateral vestibular schwannomas, meningiomas, gliomas, or neurofibromas. Most cases present clinically in early adulthood, but there is great variability [90]. Treatment for these tumors is essentially surgical, but radiotherapy may be considered. There appears to be increased risk for radiation-induced malignancy in NF2 patients, although the risk of malignant degeneration appears less common than that seen with NF1 [87]. The increased mortality associated with NF2 is mostly due to progressive, benign intracranial disease, or complications of treatment. There are guidelines for surveillance of patients with NF2 [91].

Gastrointestinal stromal tumors (GIST) are rare connective tissue tumors arising within the gastrointestinal tract, and may occur in a syndromic fashion. The causal mutation in GIST most commonly activates KIT, a receptor tyrosine kinase [92] and less commonly, activating mutations occur in *PDGFRA*. Germ-line KIT mutations have been reported in several families and are characterised by GISTs, hyperpigmentation, urticaria pigmentosa, and dysphagia [93–95]. Interestingly, there seems to be overlap with heritable intestinal NF, which has been associated with *PDGFRA* mutations [96]. It has been proposed that intestinal NF may be a form of GIST. Intestinal nerve sheath tumors have been reported concurrently with GIST in four patients. Three of these cases had NF1 mutations, suggesting phenotypic similarities resulting from inherited *KIT*, *PDGFR*, and *NF1* mutations [97].

Carney triad is a syndrome of GIST, pulmonary chondroma, and paraganglioma for which a genetic cause has not been identified [98]. On the other hand, Carney Stratakis syndrome is an autosomally dominant inherited syndrome characterised by GIST and paragangliomas thought to be due in some cases to mutations in *SDHB*, *SDHC*, and *SDHD* [99,100].

Several other syndromes provide substantial evidence for genetic drivers of sarcoma [1]. Werner's, Bloom's and Rothmund-Thompson syndromes are due to heritable defects in helicases and are associated with an increased risk of sarcomas. Multiple exostosis syndromes due mostly to *EXT1*, *EXT2*, or *EXT3* mutations have a risk of malignant transformation estimated at 0.5–50% of cases [101,102]. An inherited predisposition to benign tumors, such as Ollier's or Mafucci's disease, is associated with an increased risk of secondary bone sarcomas although the genetic causes are unknown. Whilst most cases appear sporadic, familial patterns have been reported [103]. Paget's disease is associated

with an increased risk of osteosarcoma estimated to be below 1% but interestingly osteosarcoma patients over the age of 60 years have an up to 50% incidence of Paget's disease [104].

Understanding inherited risk is clinically important for several reasons. Many sarcoma patients are in their reproductive years, and effective strategies now exist for antenatal and pre-gestational diagnosis if a cancer predisposition gene is identified [105]. Approximately 60% of patients diagnosed with sarcoma are cured, even in *TP53* [72] mutation carriers. Surveillance programs for LFS and other multi-organ cancer prone syndromes may modify subsequent cancer risk. Finally, ionizing radiation increases cancer risks synergistically in the presence of mutations in sarcoma genes such as *Rb* and *TP53* [16,76,106]. Diagnostic and therapeutic radiation exposure is modifiable in high-risk individuals, provided such mutations are identified. Taken together, the pace of recent developments in genomic screening and screening programs suggests that the issue of genetic testing and management will become an increasingly important area for research.

OTHER GENETIC RISK FACTORS IN SARCOMA

The advent of massively parallel sequencing is ushering in a new era of cancer genetics, for several reasons. The first is that, outside of well-designed germline genetic studies, somatic mutation testing is increasingly being applied with a view to personalized therapies. In the course of these assays, germline genetic variation is being identified in the absence of a familial context, raising important questions as to the meaning of known variants in this setting. As significantly, the parallel testing of multiple genes is uncovering polygenic influences on individual cancer risk, for which the clinical genetics community are not well prepared. It is likely that polygenic contributions to cancer risk represent a significant component of cancer risk, both conceptually and quantitatively.

Genome-wide association studies (GWAS) represent an important advance over the study of Mendelian dominant syndromes in mapping the architecture of cancer risk in the population at large. The power of GWAS is in identifying novel biological contributors to cancer biology, relevant to human cancer. The largest osteosarcoma GWAS performed to date recently identified two novel susceptibility loci [107], *GRM4* and a locus on chromosome 2. *GRM4* encodes the glutamate metabotropic receptor 4, which involves the cyclic AMP pathway. The cAMP-dependent protein kinase 1A was recently shown to have tumor suppressor functions in osteosarcoma [108], while parathyroid hormone, which mediates its activity via intracellular cAMP levels, causes osteosarcoma in Fisher 344 rats [109]. *GRM4* is associated with increased tumor recurrence in colorectal cancer [110]. A second set of signals implicated a locus in a gene desert on chromosome 2, about which nothing more is known [107]. GWAS loci typically identify regions associated with modest increases in cancer risk, and the region within *GRM4* conferred an odds ratio of 1.57, while the region on chromosome 2 had an odds ratio of 1.39 [107].

SUMMARY

Sarcomas are a group of rare tumors affecting a young population and displaying ethnic variation in subtype incidence. The rarity of the disease and the multiple subtypes presents challenges in understanding the etiologies. Environmental risk factors include radiation exposure and chemical carcinogens. Genetic predisposition to sarcoma has been well characterized in some familial cancer syndromes. It increasingly appears that inherited cancer risk is important in sarcoma families. In this era of increasing sequencing capacity it is important that we continue to elucidate these heritable mechanisms. This will lead to improved outcomes for sarcoma families by increasing options for reproductive decision making, treatment, surveillance, and clinical management.

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REFERENCES

1. International Agency for Research on Cancer: WHO Classification of Tumours of Soft Tissue and Bone. 4th edition. France: International Agency for Research on Cancer (IARC); 2013. pp. 281–295.
2. Worch J, Matthay KK, Neuhaus J, et al.: Ethnic and racial differences in patients with Ewing sarcoma. *Cancer* 2010;116:983–988.
3. Niu X, Zhang Q, Hao L, et al.: Giant cell tumor of the extremity: Retrospective analysis of 621 Chinese patients from one institution. *J Bone Joint Surg Am* 2012;94:461–467.
4. Hsieh MC, Wu XC, Andrews PA, et al.: Racial and ethnic disparities in the incidence and trends of soft tissue sarcoma among adolescents and young adults in the United States, 1995–2008. *J Adolesc Young Adult Oncol* 2013;2:89–94.
5. Alvegard TA, Berg NO: Histopathology peer review of high-grade soft tissue sarcoma: The Scandinavian Sarcoma Group experience. *J Clin Oncol* 1989;7:1845–1851.
6. Presant CA, Russell WO, Alexander RW, et al.: Soft-tissue and bone sarcoma histopathology peer review: The frequency of disagreement in diagnosis and the need for second pathology opinions. The Southeastern Cancer Study Group experience. *J Clin Oncol* 1986;4:1658–1661.
7. Bijwaard H: Mechanistic models of bone cancer induction by radium and plutonium in animals compared to humans. *Radiat Prot Dosimetry* 2006;122:340–344.
8. Ellender M, Harrison JD, Pottinger H, et al.: Induction of osteosarcoma and acute myeloid leukaemia in CBA/H mice by the alpha-emitting nuclides, uranium-233, plutonium-239 and americium-241. *Int J Radiat Biol* 2001;77:41–52.
9. Miller SC, Lloyd RD, Bruenger FW, et al.: Comparisons of the skeletal locations of putative plutonium-induced osteosarcomas in humans with those in beagle dogs and with naturally occurring tumors in both species. *Radiat Res* 2003;160:517–523.
10. Carnes BA, Groer PG, Kotek TJ: Radium dial workers: Issues concerning dose response and modeling. *Radiat Res* 1997;147:707–714.
11. Wakeford R: Radiation in the workplace—a review of studies of the risks of occupational exposure to ionising radiation. *J Radiol Prot* 2009;29:A61–A79.
12. Preston DL, Ron E, Tokuoka S, et al.: Solid cancer incidence in atomic bomb survivors: 1958–1998. *Radiat Res* 2007;168:1–64.
13. Rubino C, Shamsaldin A, Le MG, et al.: Radiation dose and risk of soft tissue and bone sarcoma after breast cancer treatment. *Breast Cancer Res Treat* 2005;89:277–288.
14. Mery CM, George S, Bertagnolli MM, et al.: Secondary sarcomas after radiotherapy for breast cancer: Sustained risk and poor survival. *Cancer* 2009;115:4055–4063.
15. Henderson TO, Whitton J, Stovall M, et al.: Secondary sarcomas in childhood cancer survivors: A report from the Childhood Cancer Survivor Study. *J Natl Cancer Inst* 2007;99:300–308.
16. Kleinerman RA, Tucker MA, Abramson DH, et al.: Risk of soft tissue sarcomas by individual subtype in survivors of hereditary retinoblastoma. *J Natl Cancer Inst* 2007;99:24–31.
17. Kleinerman RA: Radiation-sensitive genetically susceptible pediatric sub-populations. *Pediatr Radiol* 2009;39:S27–S31.
18. Hisada M, Garber JE, Fung CY, et al.: Multiple primary cancers in families with Li-Fraumeni syndrome. *J Natl Cancer Inst* 1998;90:606–611.
19. Sharif S, Ferner R, Birch JM, et al.: Second primary tumors in NF1 patients treated for optic glioma: Substantial risks after radiotherapy. *J Clin Oncol* 2006;24:2570–2575.

20. Elboim M, Gazit R, Gur C, et al.: Tumor immunoediting by NKp46. *J Immunol* 184:5637–5644.
21. Barbehenn EK, Lurie P, Wolfe SM: Osteosarcoma risk in rats using PTH 1–34. *Trends Endocrinol Metab* 2001;12:383.
22. Neer RM, Arnaud CD, Zanchetta JR, et al.: Effect of parathyroid hormone (1–34) on fractures and bone mineral density in postmenopausal women with osteoporosis. *N Engl J Med* 2001;344:1434–1441.
23. Rhodes MC, Bucher JR, Peckham JC, et al.: Carcinogenesis studies of benzophenone in rats and mice. *Food Chem Toxicol* 2007;45:843–851.
24. Hong HL, Ton TV, Devereux TR, et al.: Chemical-specific alterations in ras, p53, and beta-catenin genes in hemangiosarcomas from B6C3F1 mice exposed to o-nitrotoluene or riddelliine for 2 years. *Toxicol Appl Pharmacol* 2003;191:227–234.
25. Merletti F, Richiardi L, Bertoni F, et al.: Occupational factors and risk of adult bone sarcomas: A multicentric case-control study in Europe. *Int J Cancer* 2006;118:721–727.
26. Smith AH, Pearce NE, Fisher DO, et al.: Soft tissue sarcoma and exposure to phenoxyherbicides and chlorophenols in New Zealand. *J Natl Cancer Inst* 1984;73:1111–1117.
27. Hoppin JA, Tolbert PE, Flanders WD, et al.: Occupational risk factors for sarcoma subtypes. *Epidemiology* 1999;10:300–306.
28. Eriksson M, Hardell L, Adami HO: Exposure to dioxins as a risk factor for STS: A population-based case-control study. *J Natl Cancer Inst* 1990;82:486–490.
29. Eriksson M, Hardell L, Berg NO, et al.: Soft-tissue sarcomas and exposure to chemical substances: A case-referent study. *Br J Ind Med* 1981;38:27–33.
30. Kang HK, Weatherbee L, Breslin PP, et al.: Soft tissue sarcomas and military service in Vietnam: A case comparison group analysis of hospital patients. *J Occup Med* 1986;28:1215–1218.
31. Greenwald P, Kovasznay B, Collins DN, et al.: Sarcomas of soft tissues after Vietnam service. *J Natl Cancer Inst* 1984;73:1107–1109.
32. Hoppin JA, Tolbert PE, Herrick RF, et al.: Occupational chlorophenol exposure and STS risk among men aged 30–60 years. *Am J Epidemiol* 1998;148:693–703.
33. The association of selected cancers with service in the US military in Vietnam. II. Soft-tissue and other sarcomas. The Selected Cancers Cooperative Study Group. *Arch Intern Med* 1990;150:2485–2492.
34. Tuomisto JT, Pekkanen J, Kiviranta H, et al.: Soft-tissue sarcoma and dioxin: A case-control study. *Int J Cancer* 2004;108:893–900.
35. Bertazzi PA, Consonni D, Bachetti S, et al.: Health effects of dioxin exposure: A 20-year mortality study. *Am J Epidemiol* 2001;153:1031–1044.
36. Gennaro V, Ceppi M, Crosignani P, et al.: Reanalysis of updated mortality among vinyl and polyvinyl chloride workers: Confirmation of historical evidence and new findings. *BMC Public Health* 2008;8:21.
37. Boffetta P, Matisane L, Mundt KA, et al.: Meta-analysis of studies of occupational exposure to vinyl chloride in relation to cancer mortality. *Scand J Work Environ Health* 2003;29:220–229.
38. Mori T, Kido C, Fukutomi K, et al.: Summary of entire Japanese thorotrast follow-up study: Updated 1998. *Radiat Res* 1999;152: S84–S87.
39. dos Santos Silva I, Malveiro F, Jones ME, et al.: Mortality after radiological investigation with radioactive Thorotrast: A follow-up study of up to fifty years in Portugal. *Radiat Res* 2003;159:521–534.
40. Falk H, Caldwell GG, Ishak KG, et al.: Arsenic-related hepatic angiosarcoma. *Am J Ind Med* 1981;2:43–50.
41. Falk H, Thomas LB, Popper H, et al.: Hepatic angiosarcoma associated with androgenic-anabolic steroids. *Lancet* 1979;2: 1120–1123.
42. Kasper ML, Schoenfield L, Strom RL, et al.: Hepatic angiosarcoma and bronchioloalveolar carcinoma induced by Fowler's solution. *Jama* 1984;252:3407–3408.
43. Gelberg KH, Fitzgerald EF, Hwang SA, et al.: Fluoride exposure and childhood osteosarcoma: A case-control study. *Am J Public Health* 1995;85:1678–1683.
44. Rous P: A sarcoma of the fowl transmissible by an agent separable from the tumor cells. *J Exp Med* 1911;13:397–411.
45. Ganem D: KSHV and the pathogenesis of Kaposi sarcoma: Listening to human biology and medicine. *J Clin Invest* 120:939–949.
46. Chang Y, Cesarman E, Pessin MS, et al.: Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science* 1994;266:1865–1869.
47. Moore PS, Chang Y: Detection of herpesvirus-like DNA sequences in Kaposi's sarcoma in patients with and without HIV infection. *N Engl J Med* 1995;332:1181–1185.
48. Carbone M, Rizzo P, Procopio A, et al.: SV40-like sequences in human bone tumors. *Oncogene* 1996;13:527–535.
49. Heinssohn S, Szendroi M, Bielack S, et al.: Evaluation of SV40 in osteosarcoma and healthy population: A Hungarian-German study. *Oncol Rep* 2009;21:289–297.
50. McClain KL, Leach CT, Jensen HB, et al.: Association of Epstein-Barr virus with leiomyosarcomas in children with AIDS. *N Engl J Med* 1995;332:12–18.
51. Gupta S, Havens PL, Southern JF, et al.: Epstein-Barr virus-associated intracranial leiomyosarcoma in an HIV-positive adolescent. *J Pediatr Hematol Oncol* 32:e144–e147.
52. Deyrup AT, Lee VK, Hill CE, et al.: Epstein-Barr virus-associated smooth muscle tumors are distinctive mesenchymal tumors reflecting multiple infection events: A clinicopathologic and molecular analysis of 29 tumors from 19 patients. *Am J Surg Pathol* 2006;30:75–82.
53. Nur S, Rosenblum WD, Katta UD, et al.: Epstein-Barr virus-associated multifocal leiomyosarcomas arising in a cardiac transplant recipient: Autopsy case report and review of the literature. *J Heart Lung Transplant* 2007;26:944–952.
54. Stewart FW, Treves N: Lymphangiosarcoma in postmastectomy lymphedema: A report of six cases in elephantiasis chirurgica. *Cancer* 1948;1:64–81.
55. Paavolainen P, Pukkala E, Pulkkinen P, et al.: Cancer incidence in Finnish hip replacement patients from 1980 to 1995: A nationwide cohort study involving 31,651 patients. *J Arthroplasty* 1999;14:272–280.
56. Visuri T, Pulkkinen P, Paavolainen P: Malignant tumors at the site of total hip prosthesis. Analytic review of 46 cases. *J Arthroplasty* 2006;21:311–323.
57. Pukkala E, Kaprio J, Koskenvuo M, et al.: Cancer incidence among Finnish world class male athletes. *Int J Sports Med* 2000;21:216–220.
58. Tavani A, Soler M, La Vecchia C, et al.: Body weight and risk of soft-tissue sarcoma. *Br J Cancer* 1999;81:890–892.
59. Felix AS, Cook LS, Gaudet MM, et al.: The etiology of uterine sarcomas: A pooled analysis of the epidemiology of endometrial cancer consortium. *Br J Cancer* 2013;108:727–734.
60. Arora RS, Alston RD, Eden TO, et al.: The contrasting age-incidence patterns of bone tumours in teenagers & young adults: Implications for aetiology. *Int J Cancer* 2012;131:1678–1685.
61. Lichtenstein P, Holm NV, Verkasalo PK, et al.: Environmental and heritable factors in the causation of cancer—analyses of cohorts of twins from Sweden, Denmark, and Finland. *N Engl J Med* 2000;343:78–85.
62. Zahm SH, Fraumeni JF Jr: The epidemiology of soft tissue sarcoma. *Semin Oncol* 1997;24:504–514.
63. Toguchida J, Yamaguchi T, Dayton SH, et al.: Prevalence and spectrum of germline mutations of the p53 gene among patients with sarcoma. *N Engl J Med* 1992;326:1301–1308.
64. Hartley AL, Birch JM, Blair V, et al.: Patterns of cancer in the families of children with soft tissue sarcoma. *Cancer* 1993;72:923–930.
65. Hwang SJ, Lozano G, Amos CI, et al.: Germline p53 mutations in a cohort with childhood sarcoma: Sex differences in cancer risk. *Am J Hum Genet* 2003;72:975–983.

66. Fernebro J, Bladstrom A, Rydholm A, et al.: Increased risk of malignancies in a population-based study of 818 STS patients. *Br J Cancer* 2006;95:986–990.
67. Hemminki K, Li X: A population-based study of familial soft tissue tumors. *J Clin Epidemiol* 2001;54:411–416.
68. Levi F, Randimbison L, Maspoli-Conconi M, et al.: Incidence of second sarcomas: A cancer registry-based study. *Cancer Causes Control* 2014;25:473–477.
69. Li FP, Fraumeni JF Jr, Mulvihill JJ, et al.: A cancer family syndrome in twenty-four kindreds. *Cancer Res* 1988;48:5358–5362.
70. Tinat J, Bougeard G, Baert-Desurmont S, et al.: 2009 version of the Chompret criteria for Li Fraumeni syndrome. *J Clin Oncol* 2009;27:e108–e109;author reply e110.
71. Lustbader ED, Williams WR, Bondy ML, et al.: Segregation analysis of cancer in families of childhood soft-tissue-sarcoma patients. *Am J Hum Genet* 1992;51:344–356.
72. Mitchell G, Ballinger ML, Wong S, et al.: High frequency of germline TP53 mutations in a prospective adult-onset sarcoma cohort. *PLoS ONE* 2013;8:e69026.
73. Varley JM: Germline TP53 mutations and Li-Fraumeni syndrome. *Hum Mutat* 2003;21:313–320.
74. Olivier M, Goldgar DE, Sodha N, et al.: Li-Fraumeni and related syndromes: correlation between tumor type, family structure, and TP53 genotype. *Cancer Res* 2003;63:6643–6650.
75. Cury NM, Ferraz VE, Silva WA Jr: TP53 p. R337H prevalence in a series of Brazilian hereditary breast cancer families. *Hered Cancer Clin Pract* 2014;12:8.
76. Backlund MG, Trasti SL, Backlund DC, et al.: Impact of ionizing radiation and genetic background on mammary tumorigenesis in p53-deficient mice. *Cancer Res* 2001;61:6577–6582.
77. Villani A, Tabori U, Schiffman J, et al.: Biochemical and imaging surveillance in germline TP53 mutation carriers with Li-Fraumeni syndrome: A prospective observational study. *Lancet Oncol* 2011;12:559–567.
78. Custodio G, Parise GA, Kiesel Filho N, et al.: Impact of neonatal screening and surveillance for the TP53 R337H mutation on early detection of childhood adrenocortical tumors. *J Clin Oncol* 2013;31:2619–2626.
79. National Institute of Health and Clinical Excellence: Familial breast cancer: The classification and care of women at risk of familial breast cancer in primary, secondary and tertiary care, in partial update of NICE clinical guideline 14. London, UK: National Institute of Health and Clinical Excellence 2006.
80. Smith-Bindman R, Lipson J, Marcus R, et al.: Radiation dose associated with common computed tomography examinations and the associated lifetime attributable risk of cancer. *Arch Intern Med* 2009;169:2078–2086.
81. Vahteristo P, Tamminen A, Karvinen P, et al.: p53, CHK2, and CHK1 genes in Finnish families with Li-Fraumeni syndrome: Further evidence of CHK2 in inherited cancer predisposition. *Cancer Res* 2001;61:5718–5722.
82. Sodha N, Mantoni TS, Tavtigian SV, et al.: Rare germ line CHEK2 variants identified in breast cancer families encode proteins that show impaired activation. *Cancer Res* 2006;66:8966–8970.
83. Sodha N, Houlston RS, Bullock S, et al.: Increasing evidence that germline mutations in CHEK2 do not cause Li-Fraumeni syndrome. *Hum Mutat* 2002;20:460–462.
84. Hansen MF, Koufos A, Gallie BL, et al.: Osteosarcoma and retinoblastoma: A shared chromosomal mechanism revealing recessive predisposition. *Proc Natl Acad Sci U S A* 1985;82:6216–6220.
85. Kleinerman RA, Tucker MA, Tarone RE, et al.: Risk of new cancers after radiotherapy in long-term survivors of retinoblastoma: An extended follow-up. *J Clin Oncol* 2005;23:2272–2279.
86. Ferner RE: Neurofibromatosis 1 and neurofibromatosis 2: A twenty first century perspective. *Lancet Neurol* 2007;6:340–351.
87. Evans DG, Baser ME, McGaughan J, et al.: Malignant peripheral nerve sheath tumours in neurofibromatosis 1. *J Med Genet* 2002;39:311–314.
88. Rasmussen SA, Yang Q, and Friedman JM: Mortality in neurofibromatosis 1: An analysis using U.S. death certificates. *Am J Hum Genet* 2001;68:1110–1118.
89. Ferner RE, Gutmann DH: International consensus statement on malignant peripheral nerve sheath tumors in neurofibromatosis. *Cancer Res* 2002;62:1573–1577.
90. Evans DG, Huson SM, Donnai D, et al.: A clinical study of type 2 neurofibromatosis. *Q J Med* 1992;84:603–618.
91. Evans DG, Baser ME, O'Reilly B, et al.: Management of the patient and family with neurofibromatosis 2: A consensus conference statement. *Br J Neurosurg* 2005;19:5–12.
92. Hirota S, Isozaki K, Moriyama Y, et al.: Gain-of-function mutations of c-kit in human gastrointestinal stromal tumors. *Science* 1998;279:577–580.
93. Beghini A, Tibiletti MG, Roversi G, et al.: Germline mutation in the juxtamembrane domain of the kit gene in a family with gastrointestinal stromal tumors and urticaria pigmentosa. *Cancer* 2001;92:657–662.
94. Chen H, Hirota S, Isozaki K, et al.: Polyclonal nature of diffuse proliferation of interstitial cells of Cajal in patients with familial and multiple gastrointestinal stromal tumours. *Gut* 2002;51:793–796.
95. Li FP, Fletcher JA, Heinrich MC, et al.: Familial GIST syndrome: Phenotypic and molecular features in a kindred. *J Clin Oncol* 2005;23:2735–2743.
96. de Raedt T, Cools J, Debiec-Rychter M, et al.: Intestinal neurofibromatosis is a subtype of familial GIST and results from a dominant activating mutation in PDGFRA. *Gastroenterology* 2006;131:1907–1912.
97. Agaimy A, Markl B, Kitz J, et al.: Peripheral nerve sheath tumors of the gastrointestinal tract: A multicenter study of 58 patients including NF1-associated gastric schwannoma and unusual morphologic variants. *Virchows Arch* 456:411–422.
98. Carney JA: Gastric stromal sarcoma, pulmonary chondroma, and extra-adrenal paraganglioma (Carney Triad): Natural history, adrenocortical component, and possible familial occurrence. *Mayo Clin Proc* 1999;74:543–552.
99. McWhinney SR, Pasini B, and Stratakis CA: Familial gastrointestinal stromal tumors and germ-line mutations. *N Engl J Med* 2007;357:1054–1056.
100. Carney JA, Stratakis CA: Familial paraganglioma and gastric stromal sarcoma: A new syndrome distinct from the Carney triad. *Am J Med Genet* 2002;108:132–139.
101. Hennekam RC: Hereditary multiple exostoses. *J Med Genet* 1991;28:262–266.
102. Matsuno T, Ichioka Y, Yagi T, et al.: Spindle-cell sarcoma in patients who have osteochondromatosis. A report of two cases. *J Bone Joint Surg Am* 1988;70:137–141.
103. Halal F, Azouz EM: Generalized enchondromatosis in a boy with only platyspondyly in the father. *Am J Med Genet* 1991;38:588–592.
104. Huvos AG: Osteogenic sarcoma of bones and soft tissues in older persons. A clinicopathologic analysis of 117 patients older than 60 years. *Cancer* 1986;57:1442–1449.
105. Verlinsky Y, Kuliev A: Current status of preimplantation diagnosis for single gene disorders. *Reprod Biomed Online* 2003;7:145–150.
106. Kemp CJ, Wheldon T, Balmain A: p53-deficient mice are extremely susceptible to radiation-induced tumorigenesis. *Nat Genet* 1994;8:66–69.
107. Savage SA, Mirabello L, Wang Z, et al.: Genome-wide association study identifies two susceptibility loci for osteosarcoma. *Nat Genet* 2013;45:799–803.
108. Molyneux SD, Di Grappa MA, Bristain AG, et al.: Prkar1a is an osteosarcoma tumor suppressor that defines a molecular subclass in mice. *J Clin Invest* 2010;120:3310–3325.
109. Vahle JL, Long GG, Sandusky G, et al.: Bone neoplasms in F344 rats given teriparatide [rhPTH(1-34)] are dependent on duration of treatment and dose. *Toxicol Pathol* 2004;32:426–438.
110. Chang HJ, Yoo BC, Lim SB, et al.: Metabotropic glutamate receptor 4 expression in colorectal carcinoma and its prognostic significance. *Clin Cancer Res* 2005;11:3288–3295.