

EXPERT REVIEW

Integrating the roles of long and small non-coding RNA in brain function and disease

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Regulatory RNA is emerging as the major architect of cognitive evolution and innovation in the mammalian brain. While the protein machinery has remained largely constant throughout animal evolution, the non protein-coding transcriptome has expanded considerably to provide essential and widespread cellular regulation, partly through directing generic protein function. Both long (long non-coding RNA) and small non-coding RNAs (for example, microRNA) have been demonstrated to be essential for brain development and higher cognitive abilities, and to be involved in psychiatric disease. Long non-coding RNAs, highly expressed in the brain and expanded in mammalian genomes, provide tissue- and activity-specific epigenetic and transcriptional regulation, partly through functional control of evolutionary conserved effector small RNA activity. However, increased cognitive sophistication has likely introduced concomitant psychiatric vulnerabilities, predisposing to conditions such as autism and schizophrenia, and cooperation between regulatory and effector RNAs may underlie neural complexity and concomitant fragility in the human brain.

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INTRODUCTION

Our understanding of mechanisms underlying the complex functioning of the human brain is experiencing a transformation. Less than 2% of the transcribed human genome (currently estimated to be around 80%) consists of protein-coding transcripts while the overwhelming proportion is non-protein-coding.¹ Furthermore, while the protein-coding genome has undergone only modest alterations throughout evolution, the non-coding genome has scaled with biodiversity and organismal complexity² (Figure 1). Importantly, evidence is now rapidly accumulating to suggest that most cellular transcription is functional thereby exposing our current lack of knowledge regarding the protein regulatory system that governs cellular processes. We have hypothesized that the evolution of a more sophisticated regulatory system, afforded by the expansion of non-coding RNA, has allowed the emergence of cellular complexity and abilities such as higher-order cognition as is illustrated through the striking and rapid expansion of the primate brain³ (Figure 1). For such precipitous changes to occur in a relatively abrupt evolutionary window there must exist mechanisms that permit, but tightly regulate, heritable genetic adaptations. To this end, non-coding RNA could significantly contribute by encompassing (1) evolutionarily conserved classes of genomic controllers, such as families of small RNA (sRNA) (for example, microRNA (miRNA), small interfering RNA (siRNA) and small nucleolar RNA (snoRNA)) and (2) newly emerged families, such as long non-coding RNA (lncRNA). The picture that is forming is one of evolving systems that increase genetic complexity through unique temporal and spatial regulation of existing processes. Here I examine two such systems; one present in all life forms (sRNA) and one newly evolved (lncRNA). Recent evidence suggests lncRNAs may provide

an additional layer of regulation for the refinement of sRNA function, particularly in the brain.

SMALL NON-CODING RNA: IT IS A SMALL WORLD (OF GENE REGULATION) AFTER ALL

Throughout evolution of life on earth, small non-coding RNAs have been present (Figure 1). For example, snoRNAs are present in archae, bacteria and eukaryotes where they direct chemical modifications such as methylation and pseudouridylation of other RNA classes including transfer and ribosomal RNA.⁴ Plants possess miRNAs and siRNAs and these sRNAs have evolved to play vital roles in diverse functional capacities such as development, defense against transgenes and viruses and long-range signaling.⁵ All eukaryotes possess RNA interference systems capable of repressing gene expression post-transcriptionally. The earliest evolved of these classes, siRNAs, are transcribed from repeat-containing elements in the genome or generated from double-stranded RNA, and repress gene expression through exact complementary binding of target sequences.⁶ miRNAs emerged during early metazoan evolution⁷ and function to repress gene expression via binding to sequences with imperfect complementarity⁸ (that is likely to increase target range) and interfere with protein expression or induce transcript degradation. Strong evolutionary pressure has resulted in an overall expansion in miRNA numbers, especially in mammals, with a high rate of turnover⁹ suggesting a progressively complex and adaptable set of genomic regulators. A less well understood family of sRNAs is the piwi-interacting RNAs (piRNAs) that were initially reported to only function in the germ-line as a defense against retrotransposon activity.^{10,11} However, this view has been recently overturned with piRNAs detected in functional

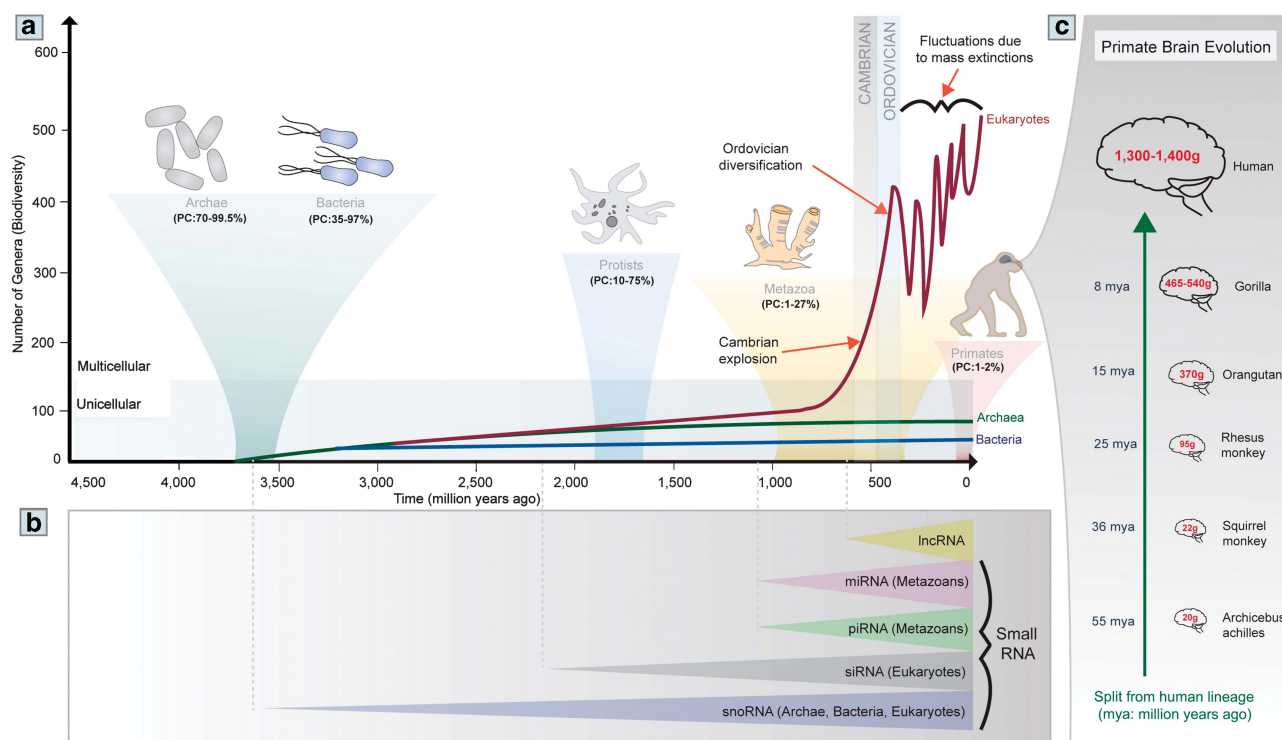


Figure 1. Evolution of organismal complexity scales with the increase in non-protein-coding size of the genome. **(a)** Life originated on earth between 3.5 and 4 billion years ago with the appearance of archae and bacteria. Their respective genomes consisted of a high percentage of protein-coding (PC) transcripts (archae 70–99.5%; bacteria 35–97%).² The emergence of protists between 1.5 and 2 billion years ago saw a significant reduction in PC percentage of the genome (10–75%) whereas metazoans first emerging around 500 million years later showed an even more striking decrease (1–27%). Primates possess only around 1–2% PC capacity in their genome. **(b)** The increase in non-protein-coding regions of the genome has seen an expansion in classes of non-coding RNA, including both small (for example, piRNA, miRNA) and long (for example, lncRNA) families. **(c)** The development of protein regulatory systems via non-coding RNA may have contributed to the considerable and rapid growth during primate brain evolution and the acquisition of higher-order cognition.³

capacities in the brain.^{12–14} Although the ancient RNAi system initially functioned mainly, or exclusively, as protectors of the genome against endogenous and exogenous threats, recent evolution has co-opted sRNA for a wide range of additional functions, notably in the brain.

sRNAs in brain development

Globally, many sRNAs are dynamically regulated during embryonic stem cell neural differentiation¹⁵ and nervous system development.¹⁶ Furthermore, disruption of common sRNA biogenesis machinery significantly impacts brain development. For example, a general loss of Dicer (resulting in absence of mature miRNA and siRNA) produces widespread neuronal alterations during development including cortical deficits and diminished differentiation,^{17,18} while conditional loss in specific cellular populations has demonstrated mechanistic deficits particular to neuronal subtypes.^{19–21} These effects may be commonly due to a critical requirement of sRNAs early in neural stem cell maintenance and expansion.²² There are many brain-specific sRNAs that clearly indicate roles in neural development and function.²³ Furthermore, distinct miRNAs have been demonstrated to influence neuronal differentiation such as brain-specific miR-9/9* and miR-124 that, when expressed in human fibroblasts, induce a neuronal fate through compositional changes of chromatin-modifying complexes.²⁴ Many additional studies supporting particular miRNAs involved in neuronal development have been documented affecting a wide variety of cellular pathways.²⁵ Prader–Willi syndrome offers insight into the importance of small interfering RNAs for early neuronal development with both SNORD115^{26,27}

and SNORD116²⁸ potentially important factors for clinical phenotypes, including neural abnormalities, observed in these patients.

sRNAs in neuronal function and dysfunction

There is an extensive body of work investigating the connection between miRNAs and neuronal activity that has been recently reviewed.²⁹ Neuronal activity reduces the expression of most neural miRNAs³⁰ emphasizing their role in fine-tuning neuronal plasticity. Indeed, Dicer gene inactivation abolishes mature miRNA formation in adult mouse forebrain and results in an enhancement of learning and memory³¹ as reduced miRNA activity permits local translation of mRNAs encoding synaptic proteins at dendritic spines and postsynaptic densities necessary for synaptic function.^{32,33} Intriguingly, a group of miRNAs, including miR-132 and miR-134, is increased in response to activity and may combine to ultimately enhance expression of proteins such as BDNF and CREB resulting in increased dendritic spine formation and maturation.³⁴ Following the recent discovery of piRNAs in the brain^{13,35} it has now been shown that this class of sRNA may have important functions in neuronal activity.¹⁴ As piRNAs essentially function through epigenetic rearrangements to block transcription of inhibitors of synaptic facilitation, and miRNAs repress translation of synaptic enhancer genes, these two classes of sRNAs when oppositely regulated by neuronal activation may synergistically combine to increase synaptic strength.¹² piRNA-mediated epigenetic changes are enduring, across many generations, and may underlie mechanisms of activity-dependent, adaptive ‘memory storage’.¹² SnoRNAs have been demonstrated to also have potential involvement in learning and memory. For example,

expression pattern studies of the brain-specific snoRNA AF357425 and Snord115 showed dynamic regulation in mouse hippocampus following contextual fear conditioning.³⁶ Additionally, an increase in the expression of Snord115, that contains complementary sequences to the serotonin 2c receptor, correlated with altered neuronal activity and autistic phenotypes *in vivo*.³⁷

The brain is highly susceptible to sRNA disturbances highlighting their critical role for correct neural function. In particular, brain disorders involving aberrant miRNA activity are extensive and well documented. For example, miRNAs are implicated in neurodevelopmental disorders (for example, Fragile X^{34,38} and Rett syndromes^{39,40}), neurodegenerative disorders (for example, Alzheimer's and Parkinson's disease^{41–43}), neuropsychiatric disorders (for example, schizophrenia^{44,45}, autism⁴⁵ and drug addiction³⁴) and brain cancers.^{46,47} Disturbances in snoRNAs may also result in deleterious neural phenotypes such as autism.^{37,48} Moreover, preliminary data suggest snoRNAs are involved in stem cell differentiation in the brain¹⁵ implying that altered activity may lead to a cancerous state and, although not yet linked to brain cancers *per se*, snoRNAs are emerging as potential contributors to tumorigenesis.⁴⁹ Vault RNAs are components of the vault ribonucleoprotein complex⁵⁰ and these small non-coding RNAs, although their exact functions are not yet clear, have recently been linked with brain cancers,⁵¹ intellectual disability⁵² and neurodegeneration.⁵³ The current escalation of research focusing on investigating sRNAs (especially those in addition to miRNA), coupled with techniques such as transcriptome-wide deep sequencing, will undoubtedly result in rapid progress to determine the full impact of their involvement in brain function and disease.

lncRNA: FINE TUNING FOR FUNCTIONAL SOPHISTICATION

In contrast to the more ancient sRNAs discussed above, lncRNAs appear later in evolution (Figure 1), where they are present in invertebrates,⁵⁴ vertebrates⁵⁵ and plants,⁵⁶ and about one-third are primate-specific.⁵⁷ lncRNAs maintain features common to protein-coding genes such as promoter regions, intron–exon boundaries and alternative splicing; however in contrast, they are mainly nuclear localized, less polyadenylated and far more tissue-specific than protein-coding genes.⁵⁷ These differences are significant as they imply that the functions of lncRNAs lie in the refinement of regulatory circuits specific to particular cells and activities, especially in the brain.⁵⁷ It is well documented that lncRNAs display strikingly unique spatial and temporal expression and are highly expressed in the specific cells where they are active.^{58,59} However, although some lncRNAs are highly expressed throughout multiple tissues, their function may not always be obvious when using a traditional knockout strategy if functional analyses are not carried out that would specifically uncover their precise, and sometimes subtle, role within the cell. For example, while mouse knockouts of the lncRNAs Hotair⁶⁰ and Xist⁶¹ result in severe phenotypes, mice with a knockout of the ubiquitously and highly expressed lncRNA Malat1^{62,63} displayed no obvious phenotype. While indications of function, such as regulation of synaptogenesis,⁶⁴ alternative splicing,⁶⁵ cell cycle control⁶⁶ and cancer⁶⁷ have been reported for Malat1 it is still unknown what the precise role is of this abundant and broadly expressed lncRNA.

lncRNAs in brain function and dysfunction

Research investigating lncRNAs is still in its relative infancy; however, great strides are rapidly being made in their functional annotation. In 2011, Guttman and colleagues used a loss-of-function strategy to extensively investigate many lncRNAs in mice and their roles in embryonic stem cell biology and differentiation.⁵⁸ Their results, and others,^{68–70} have reinforced the

importance of lncRNAs in the regulation of cellular maintenance and cell fate, especially in the brain and particularly through governing epigenetic processes. In fact, an emerging theme for lncRNAs is control of epigenetic targeting via three-dimensional modularity through their ability to bind RNA, DNA and protein.^{71–73}

Functional domains contained within lncRNAs allow fine-tuning of activity-dependent cellular regulation by way of scaffolds or molecular 'sponges'.^{65,72–74} For example, Malat1 has been shown to be involved in synapse formation⁶⁴ and act as a splicing factor 'sponge' suggesting a role in alternative splicing in neural cells.⁷⁵ Recently, the first mechanistic illustration of an lncRNA involved in neuronal activity-dependent alternative splicing was demonstrated for Gomafu.⁷⁴ This study also showed Malat1 expression was unchanged in induced pluripotent stem cell-derived neurons upon stimulation, indicating that Malat1 may either play a subtly different role in human neuronal function, or an equivalent role in different subsets of neural cells. The former hypothesis may be more likely as both Malat1 and Gomafu are present in non-overlapping nuclear-retained speckle-like structures in neurons⁷⁶ suggesting discrete nuclear speckle bodies are independently regulated. In addition to epigenetic targeting and alternative splicing, lncRNAs are involved in mRNA processing, stability, translation and decay^{66,72,77} and, taken together, places the modular functions of lncRNAs at a cellular crossroad with enormous regulatory potential.

As with sRNAs, disruptions to widespread lncRNA-mediated functions would have adverse consequences. This is especially important in the brain where most tissue-specific lncRNAs are expressed.⁵⁷ Indeed, it is emerging that lncRNAs are associated with a wide range of neurodevelopmental, neurodegenerative and psychiatric diseases^{74,78–84} and brain cancer.^{85,86} With the advent of induced pluripotent stem cell technology, coupled with next generation sequencing, it is now possible to generate tissue- and subtype-specific human neural cells from disease-affected and unaffected subjects, and comprehensively analyze the transcriptome and proteome to determine causality.⁸⁷ These techniques, focusing on distinct cell populations, will undoubtedly reveal many more long non-coding RNAs as critical regulators of normal human brain activity and associated disorders.

LONG AND sRNAs: REGULATING THE REGULATOR

The discussion to this point has revealed many overlapping functional themes between long and sRNAs in the brain with common links to neurogenesis and neuronal activity and, in dysfunction, to neurodevelopmental, psychiatric and neurodegenerative disease. Therefore, it should come as no surprise that these two classes of non-coding RNA would be intimately linked in cellular function. Furthermore, evolutionary data suggest that lncRNA, appearing after sRNAs, may have arisen as an upstream regulatory tier of sRNA-directed transcriptional control. This seems to be the case, especially for widely studied miRNAs, although feedback mechanisms may also be employed^{88,89} (Figure 2). First, lncRNAs can act as precursors from which miRNAs are generated,⁹⁰ such as in the case of miR-675 that is contained within exon 1 of the lncRNA H19.⁹¹ The role of H19, therefore, may function to regulate the release of miR-675 and its potent anti-proliferative and anti-Igf signaling properties.⁹² The process of sRNA regulation by lncRNA precursors also extends to snoRNAs⁹³ and piRNAs,¹¹ and may well exist as a common checkpoint for sRNA production. Second, lncRNAs and miRNAs may bind directly to affect the transcriptional landscape. A recent report examined deep sequencing data sets produced by photoactivatable ribonucleoside-enhanced cross-linking and immunoprecipitation (PAR-CLIP) for Argonaute proteins in HEK293 cells to elucidate direct miRNA-binding sites in the human genome.⁹⁴ The authors report finding expected protein-coding gene targets but,

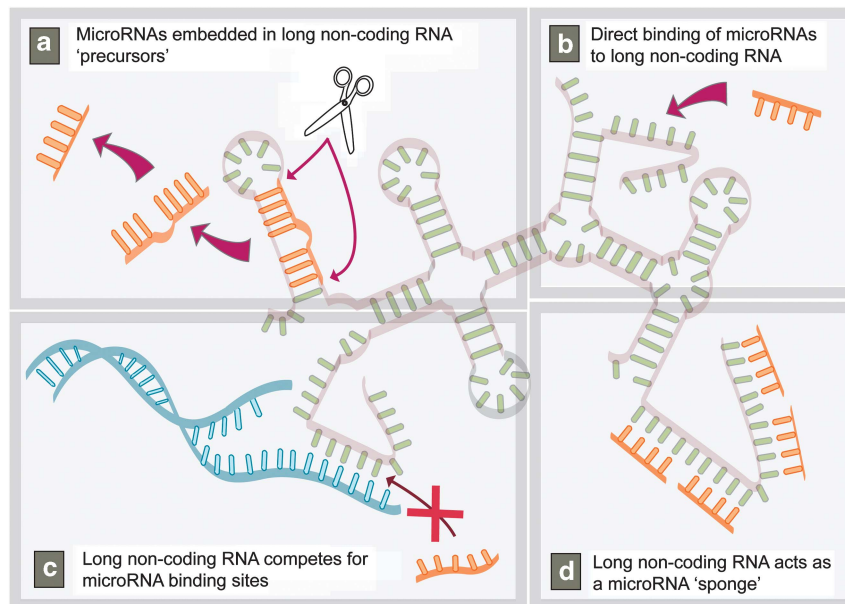


Figure 2. Regulatory interactions between long non-coding RNA (lncRNA), and microRNA (miRNA). (a) miRNAs may be embedded in lncRNA precursors from which they are generated;⁹⁰ (b) miRNAs may bind broadly to lncRNAs resulting in as yet undefined cellular effects;⁹⁴ (c) lncRNAs compete for miRNA binding sites⁹⁶ and (d) lncRNA act as miRNA 'sponges' to regulate suppressor activity.⁹⁷

surprisingly, many lncRNAs were additionally targeted by miRNAs inferring an as yet unexplored feature of small and long RNA interaction. This is illustrated in a separate study that assembled all known miRNA–lncRNA interactions and computationally predicted that the majority of interactions are yet to be discovered.⁹⁵ Third, lncRNAs can alter miRNA binding through direct competition for binding sites. For example, the lncRNA BACE1-antisense, that is upregulated in Alzheimer's disease pathology, physically blocks the binding of miR-485-5p to BACE1 target sites resulting in a reduction of miR-485-5p-induced repression of endogenous BACE1 protein translation.⁹⁶ Finally, lncRNAs may also function as miRNA 'sponges' thereby decreasing their suppressor functions. This is illustrated by an lncRNA expressed antisense to the PTEN pseudogene, PTENpg1 that regulates PTEN expression partly through its miRNA sponge activity.⁹⁷ The lncRNA lincRNA-RoR also binds miRNAs resulting in the upregulation of principal transcription factors involved in embryonic stem cell maintenance and differentiation.⁹⁸ Collectively, regulatory interactions between long and sRNAs represent an advanced layer of genome modulation, combining the widespread effects of sRNAs with the specificity of lncRNAs.

TARGETING NON-CODING RNAs FOR THERAPEUTIC INTERVENTION

Genome-wide association studies of psychiatric disease have uncovered only a small amount of genetic links to protein-coding regions.⁹⁹ It has been observed that most disease-associated haplotype blocks from genome-wide association studies occur in non-coding regions of the genome^{100–103} and combined with strong experimental evidence for lncRNAs and sRNAs underpinning neural processes (for example, lncRNAs;^{3,74,104,105} sRNAs^{14,25,34,106}) and brain disease (lncRNAs;^{74,104} sRNAs^{42,45,46}) suggest that a major proportion of disease-causing variants reside in regulatory non-coding genes.

Clearly, regulatory RNAs provide excellent opportunities for targeted therapeutic interventions in brain disease given their functional and regional specificity and an implied reduction in potential adverse side effects. The challenges are (1) uncovering

the disease-relevant signaling pathways and related druggable targets, (2) an efficient method of genetic manipulation and (3) effective delivery systems. Overcoming the first challenge is underway with specific small^{23,28,29,34,42,44–47,106–109} and long^{74,78,83,85,104,110} RNA targets, and their related signaling pathways, being discovered that may alleviate brain disorders if effectively targeted. The second challenge requires strategies for genetic manipulation and assessment and viability of multiple methods for targeted transcript knockdown. This includes sRNA mimics and antagonist oligonucleotides (for example, Antago-miRs), locked nucleic acid antisense oligonucleotides and synthetic miRNA sponges.^{111–113} An effective delivery method is the final challenge and positive progress is being made exploiting a number of different systems such as exosomes^{114–116} and nanoparticles,^{117,118} while increasing our understanding of the optimal formulation for non-viral siRNA delivery.¹¹⁹

FUTURE PERSPECTIVES

This article aims to elaborate on the significance of non-coding RNA in brain function and to unify the independent fields of small and long non-coding research to better understand protein regulatory systems that underpin evolved neurological processes. An appealing picture emerges where the ancient sRNA system, a widespread post-transcriptional genomic supervisor, is itself governed by a newly evolved layer of regulation imparted by the lncRNAs. Our knowledge of brain function will undoubtedly improve through research into all classes of sRNAs and uncovering the functional repertoire of lncRNAs, to complement the more comprehensively investigated miRNAs. Moreover, a deeper appreciation of the interactions between DNA, proteins and regulatory RNA within a three dimensional cellular architecture (or four dimensional when including time or activity) will allow deciphering of the exquisite cellular control of human neural cells and how dysregulation at multiple levels may result in similar adverse neurodevelopmental, psychiatric and neurodegenerative phenotypes.

CONFLICT OF INTEREST

The author declares no conflict of interest.

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REFERENCES

- Dunham I, Kundaje A, Aldred SF, Collins PJ, Davis CA, Doyle F *et al.* An integrated encyclopedia of DNA elements in the human genome. *Nature* 2012; **489**: 57–74.
- Liu G, Mattick JS, Taft RJ. A meta-analysis of the genomic and transcriptomic composition of complex life. *Cell Cycle* 2013; **12**: 2061–2072.
- Barry G, Mattick JS. The role of regulatory RNA in cognitive evolution. *Trends Cogn Sci* 2012; **16**: 497–503.
- Matera AG, Terns RM, Terns MP. Non-coding RNAs: lessons from the small nuclear and small nucleolar RNAs. *Nat Rev Mol Cell Biol* 2007; **8**: 209–220.
- Parent JS, Martinez de Alba AE, Vaucheret H. The origin and effect of small RNA signaling in plants. *Front Plant Sci* 2012; **3**: 179.
- Shabalina SA, Koonin EV. Origins and evolution of eukaryotic RNA interference. *Trends Ecol Evol* 2008; **23**: 578–587.
- Grimson A, Srivastava M, Fahey B, Woodcroft BJ, Chiang HR, King N *et al.* Early origins and evolution of microRNAs and Piwi-interacting RNAs in animals. *Nature* 2008; **455**: 1193–1197.
- Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; **136**: 215–233.
- Meunier J, Lemoine F, Soumillon M, Liechti A, Weier M, Guschanski K *et al.* Birth and expression evolution of mammalian microRNA genes. *Genome Res* 2013; **23**: 34–45.
- Houwing S, Kamminga LM, Berezikov E, Cronembold D, Girard A, van den Elst H *et al.* A role for Piwi and piRNAs in germ cell maintenance and transposon silencing in Zebrafish. *Cell* 2007; **129**: 69–82.
- Brenneke J, Aravin AA, Stark A, Dus M, Kellis M, Sachidanandam R *et al.* Discrete small RNA-generating loci as master regulators of transposon activity in *Drosophila*. *Cell* 2007; **128**: 1089–1103.
- Landry CD, Kandel ER, Rajasethupathy P. New mechanisms in memory storage: piRNAs and epigenetics. *Trends Neurosci* 2013; **36**: 535–542.
- Lee EJ, Banerjee S, Zhou H, Jammalamadaka A, Arcila M, Manjunath BS *et al.* Identification of piRNAs in the central nervous system. *RNA* 2011; **17**: 1090–1099.
- Rajasethupathy P, Antonov I, Sheridan R, Frey S, Sander C, Tuschl T *et al.* A role for neuronal piRNAs in the epigenetic control of memory-related synaptic plasticity. *Cell* 2012; **149**: 693–707.
- Skreka K, Schafferer S, Nat IR, Zywicki M, Salti A, Apostolova G *et al.* Identification of differentially expressed non-coding RNAs in embryonic stem cell neural differentiation. *Nucleic Acids Res* 2012; **40**: 6001–6015.
- Kapsimali M, Kloosterman WP, de Bruijn E, Rosa F, Plasterk RH, Wilson SW. MicroRNAs show a wide diversity of expression profiles in the developing and mature central nervous system. *Genome Biol* 2007; **8**: R173.
- Giraldez AJ, Cinalli RM, Glasner ME, Enright AJ, Thomson JM, Baskerville S *et al.* MicroRNAs regulate brain morphogenesis in zebrafish. *Science* 2005; **308**: 833–838.
- De Pietri Tonelli D, Pulvers JN, Haffner C, Murchison EP, Hannon GJ, Huttner WB. miRNAs are essential for survival and differentiation of newborn neurons but not for expansion of neural progenitors during early neurogenesis in the mouse embryonic neocortex. *Development* 2008; **135**: 3911–3921.
- Cuellar TL, Davis TH, Nelson PT, Loeb GB, Harfe BD, Ullian E *et al.* Dicer loss in striatal neurons produces behavioral and neuroanatomical phenotypes in the absence of neurodegeneration. *Proc Natl Acad Sci USA* 2008; **105**: 5614–5619.
- Schaefer A, O'Carroll D, Tan CL, Hillman D, Sugimori M, Llinas R *et al.* Cerebellar neurodegeneration in the absence of microRNAs. *J Exp Med* 2007; **204**: 1553–1558.
- Davis TH, Cuellar TL, Koch SM, Barker AJ, Harfe BD, McManus MT *et al.* Conditional loss of Dicer disrupts cellular and tissue morphogenesis in the cortex and hippocampus. *J Neurosci* 2008; **28**: 4322–4330.
- Kawase-Koga Y, Low R, Otaegi G, Pollock A, Deng H, Eisenhaber F *et al.* RNAase-III enzyme Dicer maintains signaling pathways for differentiation and survival in mouse cortical neural stem cells. *J Cell Sci* 2010; **123**: 586–594.
- Nowak JS, Michlewski G. miRNAs in development and pathogenesis of the nervous system. *Biochem Soc Trans* 2013; **41**: 815–820.
- Yoo AS, Sun AX, Li L, Shcheglovitov A, Portmann T, Li Y *et al.* MicroRNA-mediated conversion of human fibroblasts to neurons. *Nature* 2011; **476**: 228–231.
- Sun AX, Crabtree GR, Yoo AS. MicroRNAs: regulators of neuronal fate. *Curr Opin Cell Biol* 2013; **25**: 215–221.
- Morabito MV, Abbas AI, Hood JL, Kesterson RA, Jacobs MM, Kump DS *et al.* Mice with altered serotonin 2C receptor RNA editing display characteristics of Prader-Willi syndrome. *Neurobiol Dis* 2010; **39**: 169–180.
- Kishore S, Stamm S. The snoRNA HBII-52 regulates alternative splicing of the serotonin receptor 2C. *Science* 2006; **311**: 230–232.
- Duker AL, Ballif BC, Bawle EV, Person RE, Mahadevan S, Alliman S *et al.* Paternally inherited microdeletion at 15q11.2 confirms a significant role for the SNORD116 C/D box snoRNA cluster in Prader-Willi syndrome. *Eur J Hum Genet* 2010; **18**: 1196–1201.
- Eacker SM, Dawson TM, Dawson VL. The interplay of microRNA and neuronal activity in health and disease. *Front Cell Neurosci* 2013; **7**: 136.
- Krol J, Busskamp V, Markiewicz I, Stadler MB, Ribi S, Richter J *et al.* Characterizing light-regulated retinal microRNAs reveals rapid turnover as a common property of neuronal microRNAs. *Cell* 2010; **141**: 618–631.
- Konopka W, Kiryk A, Novak M, Herwerth M, Parkitna JR, Wawrzyniak M *et al.* MicroRNA loss enhances learning and memory in mice. *J Neurosci* 2010; **30**: 14835–14842.
- Ashraf SI, McLoon AL, Scarsic SM, Kunes S. Synaptic protein synthesis associated with memory is regulated by the RISC pathway in *Drosophila*. *Cell* 2006; **124**: 191–205.
- Banerjee S, Neveu P, Kosik KS. A coordinated local translational control point at the synapse involving relief from silencing and MOV10 degradation. *Neuron* 2009; **64**: 871–884.
- Im HI, Kenny PJ. MicroRNAs in neuronal function and dysfunction. *Trends Neurosci* 2012; **35**: 325–334.
- Dharap A, Nakka VP, Vemuganti R. Altered expression of PIWI RNA in the rat brain after transient focal ischemia. *Stroke* 2011; **42**: 1105–1109.
- Rogelj B, Hartmann CE, Yeo CH, Hunt SP, Giese KP. Contextual fear conditioning regulates the expression of brain-specific small nucleolar RNAs in hippocampus. *Eur J Neurosci* 2003; **18**: 3089–3096.
- Nakatani J, Tamada K, Hatanaka F, Ise S, Ohta H, Inoue K *et al.* Abnormal behavior in a chromosome-engineered mouse model for human 15q11-13 duplication seen in autism. *Cell* 2009; **137**: 1235–1246.
- Sellier C, Freyermuth F, Tabet R, Tran T, He F, Ruffenach F *et al.* Sequestration of DROSHA and DGCR8 by expanded CGG RNA repeats alters microRNA processing in fragile X-associated tremor/ataxia syndrome. *Cell Rep* 2013; **3**: 869–880.
- Wu H, Tao J, Chen PJ, Shahab A, Ge W, Hart RP *et al.* Genome-wide analysis reveals methyl-CpG-binding protein 2-dependent regulation of microRNAs in a mouse model of Rett syndrome. *Proc Natl Acad Sci U S A* 2010; **107**: 18161–18166.
- Urdinguio RG, Fernandez AF, Lopez-Nieva P, Rossi S, Huertas D, Kulis M *et al.* Disrupted microRNA expression caused by Mecp2 loss in a mouse model of Rett syndrome. *Epigenetics* 2010; **5**: 656–663.
- Tan L, Yu JT, Hu N, Tan L. Non-coding RNAs in Alzheimer's disease. *Mol Neurobiol* 2013; **47**: 382–393.
- Abe M, Bonini NM. MicroRNAs and neurodegeneration: role and impact. *Trends Cell Biol* 2013; **23**: 30–36.
- Salta E, De Strooper B. Non-coding RNAs with essential roles in neurodegenerative disorders. *Lancet Neurol* 2012; **11**: 189–200.
- Beveridge NJ, Cairns MJ. MicroRNA dysregulation in schizophrenia. *Neurobiol Dis* 2012; **46**: 263–271.
- Mellios N, Sur M. The emerging role of microRNAs in schizophrenia and autism spectrum disorders. *Front Psychiatry* 2012; **3**: 39.
- Zhang Y, Dutta A, Abounader R. The role of microRNAs in glioma initiation and progression. *Front Biosci (Landmark Ed)* 2012; **17**: 700–712.
- Zhi F, Wang S, Wang R, Xia X, Yang Y. From small to big: microRNAs as new players in medulloblastomas. *Tumour Biol* 2013; **34**: 9–15.
- Dykens EM, Lee E, Roof E. Prader-Willi syndrome and autism spectrum disorders: an evolving story. *J Neurodev Disord* 2011; **3**: 225–237.
- Williams GT, Farzaneh F. Are snoRNAs and snoRNA host genes new players in cancer? *Nat Rev Cancer* 2012; **12**: 84–88.
- van Zon A, Mossink MH, Scheper RJ, Sonneveld P, Wiemer EA. The vault complex. *Cell Mol Life Sci* 2003; **60**: 1828–1837.
- Li CC, Eaton SA, Young PE, Lee M, Shuttlesworth R, Humphreys DT *et al.* Glioma microvesicles carry selectively packaged coding and non-coding RNAs which alter gene expression in recipient cells. *RNA Biol* 2013; **10**: 1333–1344.
- Hussain S, Sajini AA, Blanco S, Dietmann S, Lombard P, Sugimoto Y *et al.* NSun2-mediated cytosine-5 methylation of vault noncoding RNA determines its processing into regulatory small RNAs. *Cell Rep* 2013; **4**: 255–261.
- Minones-Moyano E, Friedlander MR, Pallares J, Kagerbauer B, Porta S, Escaramis G *et al.* Upregulation of a small vault RNA (svtRNA2-1a) is an early event in Parkinson disease and induces neuronal dysfunction. *RNA Biol* 2013; **10**: 1093–1106.
- Young RS, Marques AC, Tibbit C, Haerty W, Bassett AR, Liu JL *et al.* Identification and properties of 1,119 candidate lincRNA loci in the *Drosophila melanogaster* genome. *Genome Biol Evol* 2012; **4**: 427–442.
- Pauli A, Valen E, Lin MF, Garber M, Vastenhouw NL, Levin JZ *et al.* Systematic identification of long noncoding RNAs expressed during zebrafish embryogenesis. *Genome Res* 2012; **22**: 577–591.

- 56 Zhang YC, Chen YQ. Long noncoding RNAs: new regulators in plant development. *Biochem Biophys Res Commun* 2013; **436**: 111–114.
- 57 Derrien T, Johnson R, Bussotti G, Tanzer A, Djebali S, Tilgner H *et al.* The GENCODE v7 catalog of human long noncoding RNAs: analysis of their gene structure, evolution, and expression. *Genome Res* 2012; **22**: 1775–1789.
- 58 Guttman M, Donaghey J, Carey BW, Garber M, Grenier JK, Munson G *et al.* lincRNAs act in the circuitry controlling pluripotency and differentiation. *Nature* 2011; **477**: 295–300.
- 59 Mercer TR, Dinger ME, Sunkin SM, Mehler MF, Mattick JS. Specific expression of long noncoding RNAs in the mouse brain. *Proc Natl Acad Sci U S A* 2008; **105**: 716–721.
- 60 Li L, Liu B, Wapinski OL, Tsai MC, Qu K, Zhang J *et al.* Targeted disruption of Hota1 leads to homeotic transformation and gene derepression. *Cell Rep* 2013; **5**: 3–12.
- 61 Marahrens Y, Panning B, Dausman J, Strauss W, Jaenisch R. Xist-deficient mice are defective in dosage compensation but not spermatogenesis. *Genes Dev* 1997; **11**: 156–166.
- 62 Eissmann M, Gutschner T, Hammerle M, Gunther S, Caudron-Herger M, Gross M *et al.* Loss of the abundant nuclear non-coding RNA MALAT1 is compatible with life and development. *RNA Biol* 2012; **9**: 1076–1087.
- 63 Nakagawa S, Ip JY, Shioi G, Tripathi V, Zong X, Hirose T *et al.* Malat1 is not an essential component of nuclear speckles in mice. *RNA* 2012; **18**: 1487–1499.
- 64 Bernard D, Prasanth KV, Tripathi V, Colasse S, Nakamura T, Xuan Z *et al.* A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *EMBO J* 2010; **29**: 3082–3093.
- 65 Tripathi V, Ellis JD, Shen Z, Song DY, Pan Q, Watt AT *et al.* The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell* 2010; **39**: 925–938.
- 66 Tripathi V, Shen Z, Chakraborty A, Giri S, Freier SM, Wu X *et al.* Long noncoding RNA MALAT1 controls cell cycle progression by regulating the expression of oncogenic transcription factor B-MYB. *PLoS Genet* 2013; **9**: e1003368.
- 67 Gutschner T, Hammerle M, Eissmann M, Hsu J, Kim Y, Hung G *et al.* The non-coding RNA MALAT1 is a critical regulator of the metastasis phenotype of lung cancer cells. *Cancer Res* 2013; **73**: 1180–1189.
- 68 Lv J, Cui W, Liu H, He H, Xiu Y, Guo J *et al.* Identification and characterization of long non-coding RNAs related to mouse embryonic brain development from available transcriptomic data. *PLoS ONE* 2013; **8**: e71152.
- 69 Ng SY, Johnson R, Stanton LW. Human long non-coding RNAs promote pluripotency and neuronal differentiation by association with chromatin modifiers and transcription factors. *EMBO J* 2012; **31**: 522–533.
- 70 Qureshi IA, Mehler MF. Emerging roles of non-coding RNAs in brain evolution, development, plasticity and disease. *Nat Rev Neurosci* 2012; **13**: 528–541.
- 71 Guttman M, Rinn JL. Modular regulatory principles of large non-coding RNAs. *Nature* 2012; **482**: 339–346.
- 72 Mercer TR, Mattick JS. Structure and function of long noncoding RNAs in epigenetic regulation. *Nat Struct Mol Biol* 2013; **20**: 300–307.
- 73 Tsai MC, Manor O, Wan Y, Mosammamapara N, Wang JK, Lan F *et al.* Long non-coding RNA as modular scaffold of histone modification complexes. *Science* 2010; **329**: 689–693.
- 74 Barry G, Briggs JA, Vanichkina DP, Poth EM, Beveridge NJ, Ratnu VS *et al.* The long non-coding RNA Gomafu is acutely regulated in response to neuronal activation and involved in schizophrenia-associated alternative splicing. *Mol Psychiatry* advance online publication, 30 April 2013; doi:10.1038/mp.2013.45 (e-pub ahead of print).
- 75 Anko ML, Neugebauer KM. Long noncoding RNAs add another layer to pre-mRNA splicing regulation. *Mol Cell* 2010; **39**: 833–834.
- 76 Sone M, Hayashi T, Tarui H, Agata K, Takeichi M, Nakagawa S. The mRNA-like noncoding RNA Gomafu constitutes a novel nuclear domain in a subset of neurons. *J Cell Sci* 2007; **120**: 2498–2506.
- 77 Gong C, Maquat LE. lncRNAs transactivate STAU1-mediated mRNA decay by duplexing with 3' UTRs via Alu elements. *Nature* 2011; **470**: 284–288.
- 78 Johnson R. Long non-coding RNAs in Huntington's disease neurodegeneration. *Neurobiol Dis* 2012; **46**: 245–254.
- 79 Lin M, Pedrosa E, Shah A, Hrabovsky A, Maqbool S, Zheng D *et al.* RNA-Seq of human neurons derived from iPS cells reveals candidate long non-coding RNAs involved in neurogenesis and neuropsychiatric disorders. *PLoS ONE* 2011; **6**: e23356.
- 80 Nishimoto Y, Nakagawa S, Hirose T, Okano HJ, Takao M, Shibata S *et al.* The long non-coding RNA nuclear-enriched abundant transcript 1_2 induces paraspeckle formation in the motor neuron during the early phase of amyotrophic lateral sclerosis. *Mol Brain* 2013; **6**: 31.
- 81 Petazzi P, Sandoval J, Szczesna K, Jorge OC, Roa L, Sayols S *et al.* Dysregulation of the long non-coding RNA transcriptome in a Rett syndrome mouse model. *RNA Biol* 2013; **10**: 1197–1203.
- 82 Talkowski ME, Maussion G, Crapper L, Rosenfeld JA, Blumenthal I, Hanscom C *et al.* Disruption of a large intergenic noncoding RNA in subjects with neurodevelopmental disabilities. *Am J Hum Genet* 2012; **91**: 1128–1134.
- 83 Ziats MN, Rennert OM. Aberrant expression of long noncoding RNAs in autistic brain. *J Mol Neurosci* 2013; **49**: 589–593.
- 84 Lipovich L, Dachtel F, Cai J, Bagla S, Balan K, Jia H *et al.* Activity-dependent human brain coding/noncoding gene regulatory networks. *Genetics* 2012; **192**: 1133–1148.
- 85 Han L, Zhang K, Shi Z, Zhang J, Zhu J, Zhu S *et al.* lncRNA pro fi le of glioblastoma reveals the potential role of lncRNAs in contributing to glioblastoma pathogenesis. *Int J Oncol* 2012; **40**: 2004–2012.
- 86 Zhang X, Sun S, Pu JK, Tsang AC, Lee D, Man VO *et al.* Long non-coding RNA expression profiles predict clinical phenotypes in glioma. *Neurobiol Dis* 2012; **48**: 1–8.
- 87 Bellin M, Marchetto MC, Gage FH, Mummery CL. Induced pluripotent stem cells: the new patient? *Nat Rev Mol Cell Biol* 2012; **13**: 713–726.
- 88 Juan L, Wang G, Radovich M, Schneider BP, Clare SE, Wang Y *et al.* Potential roles of microRNAs in regulating long intergenic noncoding RNAs. *BMC Med Genomics* 2013; **6**(Suppl 1): S7.
- 89 Leucci E, Patella F, Waage J, Holmstrom K, Lindow M, Porse B *et al.* MicroRNA-9 targets the long non-coding RNA MALAT1 for degradation in the nucleus. *Sci Rep* 2013; **3**: 2535.
- 90 He S, Su H, Liu C, Skogerboe G, He H, He D *et al.* MicroRNA-encoding long non-coding RNAs. *BMC Genomics* 2008; **9**: 236.
- 91 Cai X, Cullen BR. The imprinted H19 noncoding RNA is a primary microRNA precursor. *RNA* 2007; **13**: 313–316.
- 92 Keniry A, Oxley D, Monnier P, Kyba M, Dandolo L, Smits G *et al.* The H19 lincRNA is a developmental reservoir of miR-675 that suppresses growth and Igf1r. *Nat Cell Biol* 2012; **14**: 659–665.
- 93 Yin QF, Yang L, Zhang Y, Xiang JF, Wu YW, Carmichael GG *et al.* Long noncoding RNAs with snoRNA ends. *Mol Cell* 2012; **48**: 219–230.
- 94 Jalali S, Bhartiya D, Lalwani MK, Sivasubbu S, Scaria V. Systematic transcriptome wide analysis of lncRNA-miRNA interactions. *PLoS ONE* 2013; **8**: e53823.
- 95 Paraskevopoulou MD, Georgakilas G, Kostoulas N, Reczko M, Maragkakis M, Dalamagas TM *et al.* DIANA-LncBase: experimentally verified and computationally predicted microRNA targets on long non-coding RNAs. *Nucleic Acids Res* 2013; **41**: D239–D245.
- 96 Faghihi MA, Zhang M, Huang J, Modarresi F, Van der Brug MP, Nalls MA *et al.* Evidence for natural antisense transcript-mediated inhibition of microRNA function. *Genome Biol* 2010; **11**: R56.
- 97 Johnsson P, Ackley A, Vidarsdottir L, Lui WO, Corcoran M, Grandner D *et al.* A pseudogene long-noncoding-RNA network regulates PTEN transcription and translation in human cells. *Nat Struct Mol Biol* 2013; **20**: 440–446.
- 98 Wang Y, Xu Z, Jiang J, Xu C, Kang J, Xiao L *et al.* Endogenous miRNA sponge lincRNA-RoR regulates Oct4, Nanog, and Sox2 in human embryonic stem cell self-renewal. *Dev Cell* 2013; **25**: 69–80.
- 99 Hindorf LA, Sethupathy P, Junkins HA, Ramos EM, Mehta JP, Collins FS *et al.* Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc Natl Acad Sci U S A* 2009; **106**: 9362–9367.
- 100 Manolio TA. Genomewide association studies and assessment of the risk of disease. *N Engl J Med* 2010; **363**: 166–176.
- 101 Pasmant E, Sabbagh A, Vidaud M, Bieche I. ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS. *FASEB J* 2011; **25**: 444–448.
- 102 Mattick JS. The genetic signatures of noncoding RNAs. *PLoS Genet* 2009; **5**: e1000459.
- 103 Taft RJ, Pang KC, Mercer TR, Dinger M, Mattick JS. Non-coding RNAs: regulators of disease. *J Pathol* 2010; **220**: 126–139.
- 104 Qureshi IA, Mehler MF. Long non-coding RNAs: novel targets for nervous system disease diagnosis and therapy. *Neurotherapeutics* 2013; **10**: 632–646.
- 105 Qureshi IA, Mattick JS, Mehler MF. Long non-coding RNAs in nervous system function and disease. *Brain Res* 2010; **1338**: 20–35.
- 106 Wang W, Kwon EJ, Tsai LH. MicroRNAs in learning, memory, and neurological diseases. *Learn Mem* 2012; **19**: 359–368.
- 107 Miller BH, Zeier Z, Xi L, Lanz TA, Deng S, Strathmann J *et al.* MicroRNA-132 dysregulation in schizophrenia has implications for both neurodevelopment and adult brain function. *Proc Natl Acad Sci U S A* 2012; **109**: 3125–3130.
- 108 Wright C, Turner JA, Calhoun VD, Perrone-Bizzozero N. Potential Impact of miR-137 and Its Targets in Schizophrenia. *Front Genet* 2013; **4**: 58.
- 109 Lau P, Bossers K, Janky R, Salta E, Frigerio CS, Barbash S *et al.* Alteration of the microRNA network during the progression of Alzheimer's disease. *EMBO Mol Med* 2013; **5**: 1613–1634.
- 110 Wahlestedt C. Targeting long non-coding RNA to therapeutically upregulate gene expression. *Nat Rev Drug Discov* 2013; **12**: 433–446.

- 111 Ruberti F, Barbato C, Cogoni C. Targeting microRNAs in neurons: tools and perspectives. *Exp Neurol* 2012; **235**: 419–426.
- 112 Martinez T, Wright N, Lopez-Fraga M, Jimenez AI, Paneda C. Silencing human genetic diseases with oligonucleotide-based therapies. *Hum Genet* 2013; **132**: 481–493.
- 113 Kole R, Krainer AR, Altman S. RNA therapeutics: beyond RNA interference and antisense oligonucleotides. *Nat Rev Drug Discov* 2012; **11**: 125–140.
- 114 Marcus ME, Leonard JN. FedExosomes: Engineering Therapeutic Biological Nanoparticles that Truly Deliver. *Pharmaceuticals (Basel)* 2013; **6**: 659–680.
- 115 El Andaloussi S, Lakhil S, Mager I, Wood MJ. Exosomes for targeted siRNA delivery across biological barriers. *Adv Drug Deliv Rev* 2013; **65**: 391–397.
- 116 Lee Y, El Andaloussi S, Wood MJ. Exosomes and microvesicles: extracellular vesicles for genetic information transfer and gene therapy. *Hum Mol Genet* 2012; **21**: R125–R134.
- 117 Zhou J, Shum KT, Burnett JC, Rossi JJ. Nanoparticle-Based Delivery of RNAi Therapeutics: Progress and Challenges. *Pharmaceuticals (Basel)* 2013; **6**: 85–107.
- 118 Muthiah M, Park IK, Cho CS. Nanoparticle-mediated delivery of therapeutic genes: focus on miRNA therapeutics. *Expert Opin Drug Deliv* 2013; **10**: 1259–1273.
- 119 O'Mahony AM, Godinho BM, Cryan JF, O'Driscoll CM. Non-viral nanosystems for gene and small interfering RNA delivery to the central nervous system: formulating the solution. *J Pharm Sci* 2013; **102**: 3469–3484.