



ELSEVIER

Available online at www.sciencedirect.com

ScienceDirect

Current Opinion in
Immunology

STAT3 is a central regulator of lymphocyte differentiation and function

Alisa Kane^{1,2}, Elissa K Deenick^{1,2}, Cindy S Ma^{1,2}, Matthew C Cook^{3,4}, Gulbu Uzel⁵ and Stuart G Tangye^{1,2}

Signalling in lymphocytes through cytokine receptors is critical for their development, activation and differentiation into effector cells that mediate protection against pathogens and provide the host with protective immunological memory. The essential role of cytokine signalling has been established not only by the generation and examination of gene-targeted mice, but also 'Experiments of Nature' whereby monogenic mutations cause primary immunodeficient conditions characterised by impaired immunity to infectious diseases due to compromised lymphocyte function. Mutations in *STAT3* cause autosomal dominant hyper-IgE syndrome. Here, we will review how the study of *STAT3*-deficient individuals has revealed non-redundant functions of *STAT3* and specific cytokines in human lymphocyte biology, and have delineated mechanisms underlying the distinct clinical features of autosomal dominant hyper-IgE syndrome.

Addresses

¹ Immunology and Immunodeficiency Group, Immunology Research Program, Garvan Institute of Medical Research, Darlinghurst, NSW, Australia

² St Vincent's Clinical School, University of New South Wales, Darlinghurst, NSW, Australia

³ John Curtin School of Medical Research, Australian National University, ACT, Australia

⁴ Department of Immunology, The Canberra Hospital, ACT, Australia

⁵ Laboratory of Clinical Infectious Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, MD, USA

Corresponding author: Tangye, Stuart G (s.tangye@garvan.org.au)

Current Opinion in Immunology 2014, 28:49–57

This review comes from a themed issue on **Lymphocyte activation and effector functions**

Edited by **Claude-Agnès Reynaud** and **Stuart Tangye**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 6th March 2014

0952-7915/\$ – see front matter, © 2014 Elsevier Ltd. All rights reserved.

<http://dx.doi.org/10.1016/j.coi.2014.01.015>

Introduction

The generation of lymphocytes from hematopoietic precursors, and their subsequent differentiation into effector cells, requires signals provided by surface receptors including those that bind Ag, co-stimulatory and adhesion molecules, and cytokines. The biological effects of cytokines are largely mediated by JAK/STAT signalling

pathways activated downstream of their specific receptors [1–3]. Four mammalian JAKs (JAK1, JAK2, JAK3, Tyk2) and seven STATs (STAT1, 2, 3, 4, 5a, 5b, 6) have been identified [1,2]. JAKs associate with the cytoplasmic domains of multimeric cytokine receptors and, following engagement by specific ligands, phosphorylate cytoplasmic tyrosine residues which act as docking sites for STATs. Receptor-associated STATs undergo JAK-mediated phosphorylation, which promotes formation of dimers that translocate to the nucleus and bind specific DNA sequences, promoting transcription of target genes and regulating the epigenetic state of chromatin [1–3].

Cytokines are fundamental for lymphocyte development and differentiation. This is evidenced by the immunodeficient and autoimmune states that arise from monogenic mutations in genes encoding cytokines (*IL17F*, *IL10*, *IL12B*), their receptors (*IL2RG*, *IL7RA*, *IL12RB1*, *IFNGR1/2*, *CD25/IL2RA*, *IL10R*, *IL21R*), or components of downstream signalling pathways (*JAK3*, *TYK2*, *STAT1*, *STAT2*, *STAT3*, *STAT5A*) (see [Table 1](#) for summaries) [3–6]. Here, we review the function of *STAT3* in generating efficient cellular and humoral immune responses, as revealed from analysis of *STAT3*-deficient humans and mice.

Autosomal dominant hyper-IgE syndrome (AD-HIES): human *STAT3* deficiency

In 1966, Davis and colleagues reported a new immunodeficiency in two unrelated girls with severe eczema and staphylococcal lung and skin infections [7]. Remarkably the skin infections were often abscesses that lacked significant inflammation (erythema and warmth) of the effected tissues. In 1972, Buckley reported two boys with similar presentations but emphasised their coarse facial features, chronic dermatitis, pneumatoceles and extreme hyper-IgE. These patients exhibited poor T-dependent (TD) Ag-specific Ab responses, despite normal levels of serum IgM, IgG and IgA. The children had uncomplicated infections with measles, but did have severe mucocutaneous fungal infections [8]. This condition was termed autosomal dominant hyper-IgE syndrome (AD-HIES; see [Figure 1](#)) [9,10].

These initial descriptions provide accurate accounts of the clinical phenotype of this rare disease; infection of the skin, lungs and upper airways with *S. aureus*, together with recurrent pneumonia and mucocutaneous — but not

Table 1

Gene mutations in cytokine signalling pathways that cause human immunodeficiency

Mutated gene	Mechanism of disease	Clinical features of disease and Infectious susceptibility	Cytokines involved
<i>IL2RG</i>	X-linked loss of function	<ul style="list-style-type: none"> • B⁺T⁻NK⁻ X-linked SCID 	<ul style="list-style-type: none"> • IL-2, IL-4, IL-7, IL-9, IL-15, IL-21
<i>JAK3</i>	Autosomal recessive (AR) loss of function	<ul style="list-style-type: none"> • B⁺T⁻NK⁻ AR SCID 	<ul style="list-style-type: none"> • IL-2, IL-4, IL-7, IL-9, IL-15, IL-21
<i>IL7RA</i>	AR loss of function	<ul style="list-style-type: none"> • B⁺T⁻NK⁺ combined immunodeficiency 	<ul style="list-style-type: none"> • IL-7, TSLP
<i>TYK2</i>	AR loss of function	<ul style="list-style-type: none"> • Originally considered as AR hyper-IgE syndrome, but not all patients have elevated IgE levels or chronic mucocutaneous candidiasis (CMC) • Mendelian susceptibility to mycobacterial disease (MSMD), especially disseminated BCG • Also susceptible to infection with salmonella, brucella and staphylococci • Recurrent cutaneous herpes simplex infection, VZV reactivation 	<ul style="list-style-type: none"> • Type I IFNs • IL-6, IL-10 families • IL-12, IL-23
<i>STAT1</i>	AR complete loss of function Autosomal dominant (AD) or AR partial loss of function AD gain of function	<ul style="list-style-type: none"> • MSMD (usually BCG) • Infection with herpes viruses (often fatal) • MSMD (Disseminated BCG and non-tuberculous mycobacterial [NTM] infections) • CMC • Susceptibility to infection with other pathogens including viruses (varicella, HSV, EBV), dimorphic yeast (histoplasmosis, coccidioidomycosis) and non-tuberculous mycobacteria • Autoimmune manifestations including endocrinopathy (IPEX-like disease) • Vascular anomalies • Defects in humoral immunity • Defects in NK cell function 	<ul style="list-style-type: none"> • Type I and II IFNs • IL-6, IL-10, IL-21, IL-27
<i>IL12RB1</i>	AR loss of function	<ul style="list-style-type: none"> • MSMD (BCG, NTM or <i>M. tuberculosis</i>) • Non-typhoidal salmonella infection, recurrent leishmaniasis • Some patients are also affected by other intracellular pathogens; dimorphic fungi (paracoccidioides and coccidioidomycosis) and candida infections 	<ul style="list-style-type: none"> • IL-12, IL-23
<i>IFNGR1</i>	 AR complete loss of function AR partial loss of function AD partial (dominant negative)	<ul style="list-style-type: none"> • MSMD • Defect in granuloma formation in the setting of NTM depends on the severity of the IFNγ signalling defect 	<ul style="list-style-type: none"> • Type I and II IFNs
<i>IFNGR2</i>	AR complete loss of function AR partial loss of function AD partial (dominant negative)	<ul style="list-style-type: none"> • Severe infections • Presents later in life with rather limited disease • Very rare – CMV, NTM infections 	<ul style="list-style-type: none"> • Type I and II IFNs
<i>STAT2</i>	AR loss of function	<ul style="list-style-type: none"> • Susceptible to vaccine-strain measles; otherwise healthy 	<ul style="list-style-type: none"> • Type I IFNs

Table 1 (Continued)

Mutated gene	Mechanism of disease	Clinical features of disease and Infectious susceptibility	Cytokines involved
<i>STAT3</i>	AD loss of function	<ul style="list-style-type: none"> • AD hyper-IgE syndrome (AD-HIES) • Susceptibility to infections with <i>S. aureus</i>, <i>S. pneumoniae</i>, <i>C. albicans</i> • EBV viraemia and VZV reactivation • Eczema, vascular, musculoskeletal, dental and connective tissue defects • B-cell lymphoma 	IL-6, IL-10 and IFN families, IL-21, IL-23, plus others
<i>STAT5b</i>	AR loss of function	<ul style="list-style-type: none"> • Growth hormone insensitivity • Tregs very diminished in number and dysfunctional, leading to immune dysregulation and autoimmunity, lymphocytic infiltrates in the lungs (lymphocytic interstitial pneumonitis), skin, T- and NK-cell lymphopenia, immunodeficiency with susceptibility to bacterial, viral (VZV), and fungal infections 	<ul style="list-style-type: none"> • IL-2, IL-15, TSLP • Growth hormone
<i>IL21R</i>	AR loss of function	<ul style="list-style-type: none"> • Combined immunodeficiency • Sinopulmonary infections and bronchiectasis • Disseminated cryptosporidium infection • Susceptibility to infection with <i>Pneumocystis jirovecii</i> • Impaired humoral immunity • Increased serum IgE 	<ul style="list-style-type: none"> • IL-21
<i>IL17RA</i> ; <i>IL17F</i>	AR deficiency of IL-17RA; partial deficiency of IL-17F homo- and heterodimers (with IL-17A)	<ul style="list-style-type: none"> • CMC 	<ul style="list-style-type: none"> • IL-17A • IL-17F
<i>CD25</i>	AR loss of function	<ul style="list-style-type: none"> • IPEX-like disease lymphocytic infiltration of skin, lung, liver, gut, and bone and endocrinopathy • Chronic CMV infection, susceptibility to bacterial and fungal infections 	<ul style="list-style-type: none"> • IL-2
<i>IL10</i> ; <i>IL10RA</i> / <i>IL10RB</i>	AR loss of function	<ul style="list-style-type: none"> • Early onset fistulising inflammatory bowel disease 	<ul style="list-style-type: none"> • IL-10 (also IL-22, IL-26 and IFNλ for <i>IL-10RB</i> mutations)

Abbreviations: AD, autosomal dominant; AR, autosomal recessive; BCG, *Bacillus Calmette–Guérin*; CMC, chronic mucocutaneous candidiasis; CMV, cytomegalovirus; EBV, Epstein Barr virus; IPEX, immunodysregulation polyendocrinopathy enteropathy X-linked; MSMD, Mendelian susceptibility to mycobacterial disease, SCID, severe combined immune deficiency; TSLP, thymic stromal lymphopoietin; VZV, varicella zoster virus. For more details, see Refs [4–6,26].

invasive systemic — candidiasis [9,10]. Strikingly, viral infections are not a distinguishing aspect of AD-HIES, although there may be impaired control of herpes viruses, leading to reactivation of EBV and VZV infection [11^{••},12[•]]. Eosinophilia is usually present, while a small proportion of patients develop B-cell lymphoma [9,10,11^{••}]. Interestingly dental, musculoskeletal, connective tissue and vascular features manifesting as retention of primary teeth, scoliosis, osteopenia, frequent fractures, short stature, joint hyperextensibility as well as intracranial and coronary aneurysms represent the non-immunological features of AD-HIES ([9,10]; Table 1 and Figure 1).

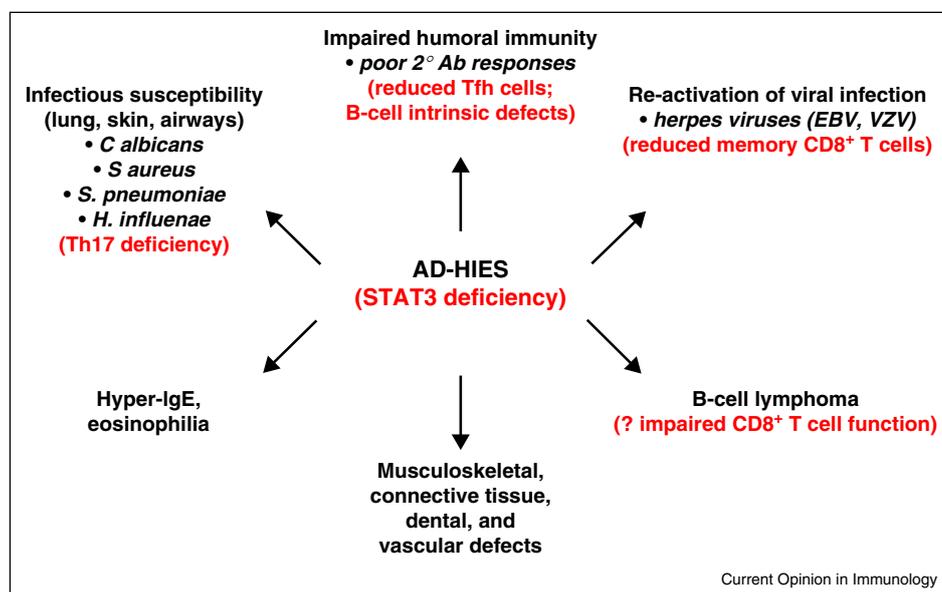
In 2007, heterozygous mutations in *STAT3* were identified as the genetic etiology of AD-HIES [13^{••},14^{••}]. This discovery underscored the importance of *STAT3* in regulating critical aspects of cellular and humoral immunity. Constitutive deletion of *Stat3* in mice is embryonically lethal due to a requirement for *STAT3*

in trophoblast invasion and placental development [15]. In contrast, the dominant negative *STAT3* alleles in AD-HIES patients reduce, but do not abolish, *STAT3* function [14^{••}]. Thus, while such mutations are not lethal, they nonetheless severely compromise immunity.

STAT3 regulates CD4⁺ T cell differentiation.

Naïve CD4⁺ T cells can differentiate into numerous effector subsets — Th1, Th2, Th9, Th17, Th22, T follicular helper (Tfh), regulatory T (Treg) cells — with unique roles in protective immunity against infectious diseases, as well as autoimmune or inflammatory disorders (Figure 2). This process is mediated by cytokines present in the microenvironment, and the signalling cascades they activate ([16–18]; Figure 2). Studies of AD-HIES patients and conditional *STAT3*-deleted mice have revealed multiple roles for *STAT3* in generating several CD4⁺ T cell subsets.

Figure 1



Clinical features and cellular defects in autosomal dominant hyper-IgE syndrome (AD-HIES) due to mutations in *STAT3*. The key clinical features of AD-HIES, and possible defects in B cells and T cells that underlie these features, are indicated. The cause of hyper-IgE, as well as the non-immunological defects, remains to be determined.

Th17 cells

Th17 cells can be induced *in vitro* from naïve precursors following stimulation with TGF- β and IL-6; the Th17 program is imprinted by additional cytokines such as IL-23 [17]. IL-6 and IL-23 activate STAT3 providing the basis for the initial discovery in mice that STAT3 plays a critical role in generating Th17 cells [19–22]. Analysis of STAT3-deficient patients revealed a lack of Th17-type cells *ex vivo* [6,23–25], as well as in response to polarising conditions *in vitro* [23] confirming the role of STAT3 in Th17 differentiation in humans as well (Figure 2). The role of IL-23 *in vivo* in this process was evident from patients with mutations in *IL12RB1* — a component of the IL-23 receptor — who also had reduced Th17 cells ([24]; Table 1). However, the reduction in Th17 cells in *IL12RB1*-deficiency was not as severe as in STAT3-deficiency, supporting a role for additional cytokines (e.g. IL-6) in Th17 generation [24]. As Th17 cells, and particularly IL-17 cytokines, are requisite for protection against fungal infections [26], the Th17 defect in AD-HIES explains the high incidence and persistence of mucocutaneous candidiasis in these individuals (Figure 1). Interestingly, *IL21R*-deficient patients do not develop candidiasis and have intact Th17 cells ([27^{••}]; Table 1), implying that — despite suggestions to the contrary from studies in mice [17,28] — IL-21 does not play a significant role in the generation and/or maintenance of human Th17 cells.

STAT3 controls Th17 differentiation by inducing genes that characterise Th17 cells, for example, transcription

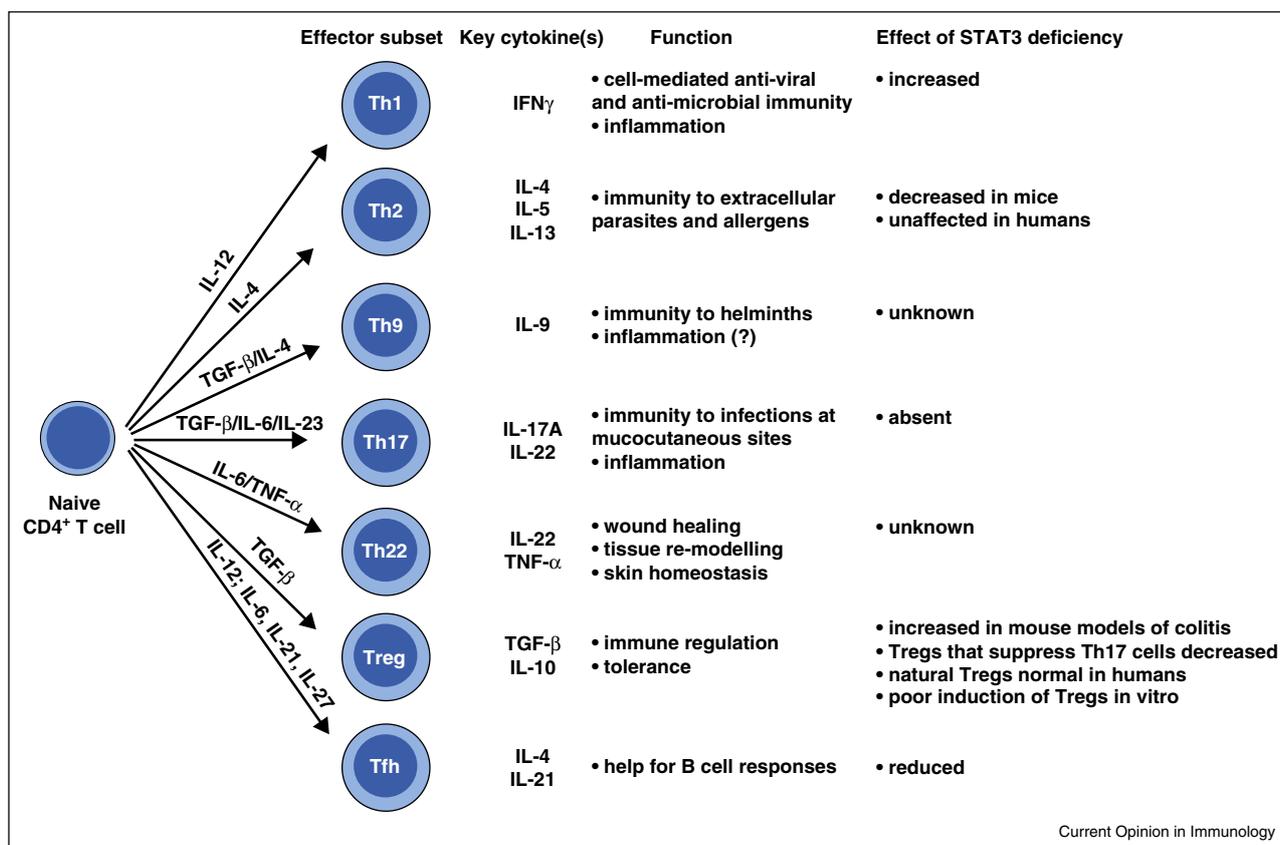
factors ROR γ t and ROR α , and effector cytokines IL-17A, IL-17F and IL-21, which can be further induced by ROR γ t [20–22,28–32]. Interestingly, STAT3 may also promote Th17 cell generation by repressing *Tbx21* (T-bet) [21], *Gata3* [33[•]] and *Foxp3* [21], thus impeding the differentiation of naïve CD4⁺ T cells to Th1, Th2 or Treg fates, respectively. Indeed, STAT3-deficient human CD4⁺ T cells generate more IFN γ ⁺ Th1-type cells *in vitro* than do normal CD4⁺ T cells ([34^{••}]; Figure 2).

Treg cells

In mice, STAT3 can either promote or restrain Treg formation by differentially regulating FoxP3 expression (Figure 2). For instance, IL-6/STAT3 signalling inhibits FoxP3 expression; thus few Tregs are generated *in vitro* under Th17 (TGF- β /IL-6) conditions [20–22]. Interestingly, IL-27 also represses TGF- β -induced FoxP3 expression by activating STAT3, which directly silences *Foxp3* [35]. Consistent with this, in a murine model of colitis, STAT3-deficient CD4⁺ T cells exhibited impaired Th17 generation, but yielded increased Tregs [30]. Conversely, STAT3 may also act in Tregs to generate a subset that specifically suppresses the function of Th17 cells as deletion of STAT3 in Tregs resulted in increased Th17-associated cytokines and pathology, without affecting Th1 or Th2 cytokines [36]. However, the cytokines that act via STAT3 to generate Tregs that control Th17 responses are unknown.

Despite these studies in mice, it is unclear whether STAT3 has an intrinsic role in the differentiation of

Figure 2



CD4⁺ T cell differentiation — effects of STAT3 deficiency. Naïve CD4⁺ T cells can differentiate into numerous subsets of effector cells with distinct roles in protection against pathogen infection. CD4⁺ T cell differentiation is regulated by cytokines provided within the stimulatory microenvironment; this requires cytokine-mediated activation of various transcription factors, including STATs, which in turn induce expression of key cytokines that underlie the effector function of each subset of CD4⁺ T cells (modified from [16]). The consequence of STAT3 deficiency on the generation of each subset in mice and humans (where known) is indicated.

human Tregs, as these cells are generated at normal frequencies in AD-HIES, and exhibit normal suppressive function *in vitro* (Figure 2; [23,37]). However, DCs from STAT3-deficient patients are unable to respond to IL-10, impairing their ability to become tolerogenic and induce Tregs from naïve CD4⁺ T cells [37]. Interestingly STAT3-deficient CD4⁺ T cells produce less IL-10 than normal CD4⁺ T cells [23]. Thus, although Tregs are generated normally in STAT3-deficient patients, the inability of STAT3-deficient DCs to convert naïve CD4⁺ T cells into Tregs, together with poor IL-10 production, opens the possibility of defective immune regulation, which could potentially contribute to atopy in AD-HIES.

Th2 cells

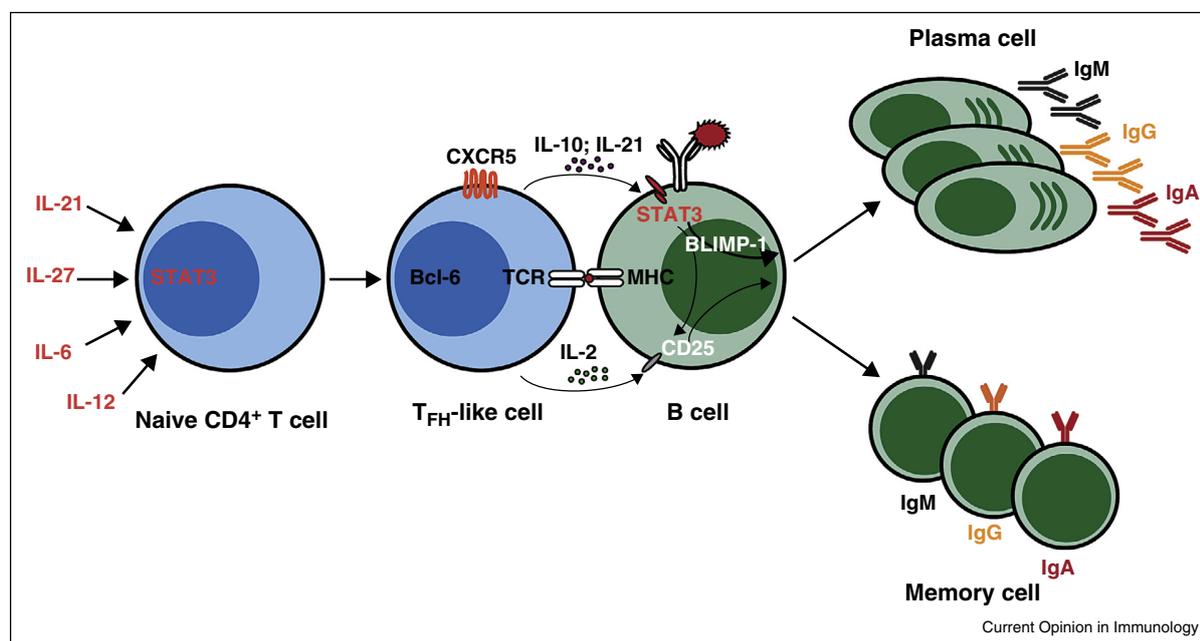
Dysregulated IgE production in AD-HIES is suggestive of excessive Th2 function. However, the differentiation of naïve human CD4⁺ T cells into Th2-type cells *in vivo* and *in vitro* appears to be unaffected by STAT3 mutations

[6,23,37]. Despite these findings, STAT3 deficiency in murine CD4⁺ T cells compromised their ability to produce the signature Th2 cytokines IL-4, IL-5 and IL-13, and express *Gata3*, under Th2 polarising culture conditions *in vitro* (Figure 2; [33]). These defects are likely due to direct regulation of Th2 transcription programs by STAT3, which was found to bind directly to *Gata3*, *cmf* and *Batf* [30,33]. Interestingly, STAT3-deficiency reduced STAT6 binding to these target genes, abolished STAT6-dependent Th2 induction *in vitro*, and ameliorated disease in a murine model of allergic inflammation [33]. Thus, STAT3, together with STAT6, is a key regulator of Th2 differentiation. But how STAT3 regulates the function of STAT6, as well as the relative role of STAT3 in human Th2 responses, remains unknown.

Tfh cells

Tfh cells provide help to B cells to differentiate into memory and plasma cells, thereby establishing long-lived

Figure 3



B-cell and T-cell intrinsic functions of STAT3 underlie successful humoral immune responses. CD4⁺ T cells differentiate into T_{FH} cells under the influence of cytokines including IL-6, IL-12, IL-21 and IL-27. The provision of IL-21 by T_{FH} cells allows for the differentiation of activated B cells into memory B cells and plasma cells. This is achieved by the induction of Blimp-1 and CD25, which promote the formation of plasma cells. The exact mechanism by which IL-21 promotes memory cell formation is unknown. Other cytokines, such as IL-10, also promote T_{FH}-mediated B-cell differentiation. This pathway of T_{FH}-dependent B-cell differentiation is compromised by STAT3-deficiency at several stages. First, fewer T_{FH} cells are generated in patients with *STAT3* mutations, thus limiting B-cell help. Second, *STAT3* mutations abolish the ability of naïve B cells to respond to the stimulatory effects of IL-21 and IL-10, thereby preventing these cytokines from inducing the formation of plasma cells from naïve precursors.

humoral immunity [38]. T_{FH} cells are typically detected in secondary lymphoid tissues, however a circulating counterpart of T_{FH} cells can be detected in peripheral blood of humans and mice [38]. AD-HIES patients have a reduction in circulating T_{FH} cells [34^{••},39,40^{••}], and *STAT3*-deficient CD4⁺ T cells failed to differentiate *in vitro* into IL-21-producing T_{FH}-like cells (Figures 2 and 3; [34^{••},41]). These findings confirm studies in mice demonstrating that *STAT3*-deficiency impedes T_{FH} formation *in vivo* by compromising signalling downstream of IL-6, IL-21 and/or IL-27 (Figures 2 and 3; [38,41–43]). However, *STAT1* may compensate for *STAT3* in some instances [43]. This deficit in T_{FH} cells would contribute to the inability of *STAT3*-deficient patients to generate normal titres of Ag-specific Ab ([8,44–46,47[•],48[•]]; Figures 1 and 3).

STAT3 and B cell function

STAT3 regulates the commitment of Ag-specific B cells to the pool of long-lived plasma cells

Although B cell development and production of basal serum IgM, IgG and IgA are normal in AD-HIES and mice that lack *STAT3* specifically in B cells, *STAT3*-deficiency causes defective TD Ag-specific Ab responses [8,44–46,47[•],48[•],49]. While *STAT3*-deficient mice only

have a reduction in IgG1-secreting plasma cells [49], AD-HIES patients have defects in both IgG and IgA Ag-specific responses [8,44–46,47[•],48[•]]. *STAT3*-deficient humans also lacked memory B cells [11^{••},40^{••},47[•],48[•]], which was not replicated in mice with *STAT3*-deficient B cells [49]. There are several possible explanations for these differences. First, they may result from species-specific differences in the regulation of B cell differentiation. Second, as memory formation in mice was assessed only at a single time point post-immunisation, the paucity of memory B cells in *STAT3*-deficient humans may stem from a defect in long-term survival rather than their generation. Third, the differences may reflect B cell-extrinsic actions of *STAT3*, since in mice only B cells lacked *STAT3*, whereas in humans all cells are *STAT3*-deficient. Defects in T_{FH} formation by *STAT3*-deficient CD4⁺ T cells provide one possible example ([34^{••},39,40^{••}], Figures 2 and 3). However, there is clearly an intrinsic defect in *STAT3*-deficient naïve B cells, as they are unable to differentiate into plasmablasts *in vitro* in response to IL-10 and IL-21 (Figure 3; [40^{••},47[•]]).

Although analysis of AD-HIES patients revealed a requirement for *STAT3* in generating Ag-specific memory and plasma cells, it did not identify the *STAT3*-activating

cytokine(s) involved in this process *in vivo*. The recent discovery of patients with *IL21R* mutations and the observation that they lack memory B cells and have defects in humoral immunity ([27^{**},40^{**}]; Table 1) established that IL-21 is likely to be the predominant cytokine upstream of STAT3 that regulates human B cell function (Figure 3). This is consistent with its superior ability to induce *PRDM1* (Blimp-1) and *XBPI* in naïve B cells, and mediate their differentiation into plasmablasts *in vitro*, compared to other STAT3 cytokines, IL-6 and IL-10 [40^{**},47^{*},50,51]. IL-21 also promotes Ab production by inducing CD25 on activated human B cells in a STAT3-dependent manner, thereby priming them to respond to IL-2 (Figure 3; [52]). Thus, IL-21R/STAT3 signalling is central to generating TD humoral immune responses in humans, and defects in this pathway underlie poor serological memory in AD-HIES and IL-21R-deficiency (Figures 1 and 3). Remarkably, the small population of memory B cells that is generated in AD-HIES is functionally normal with respect to responding to STAT3 cytokines and yielding plasmablasts [40^{**}]. This highlights differences in requirements for activating naïve and memory B cells, and suggests that STAT3-deficient memory B cells may contribute to variable levels of Ag-specific Ab titres detected in some AD-HIES patients [47^{*},48^{*}].

CD8⁺ T cells

STAT3 deficiency also affects CD8⁺ T cells in humans and mice. STAT3-deficient human CD8⁺ T cells are impaired in their ability to upregulate expression of perforin and granzyme B in response to IL-21 and IL-15, however this can be circumvented by TCR ligation [53^{*}]. Additionally, mice whose CD8⁺ T cells lack STAT3 have no defect in early viral clearance [54^{*}]. These findings suggest sufficient redundancy exists in STAT3-deficient CD8⁺ T cells to ensure acquisition of cytolytic function in response to pathogen infection, despite impaired signalling to some key cytokines. Despite this apparently intact effector function, STAT3-deficiency did impact the generation of memory CD8⁺ T cells in both species [12^{*},53^{*},54^{*}], possibly due to reduced expression of the transcription factors Bcl-6 and comsodermin, which govern the formation of some memory cell subsets [12^{*},54^{*}]. Murine studies found that combined deficiency of IL-10 and IL-21 phenocopied the memory defect in STAT3-deficient CD8⁺ T cells [54^{*}]. IL-21R-deficient humans also had fewer memory CD8⁺ T cells, suggesting that in humans IL-21 — via STAT3 — contributes to the formation and/or maintenance of CD8⁺ T cell memory [53^{*}]. STAT3-deficient CD8⁺ memory T cells failed to protect mice from secondary viral infections [54^{*}], while expression of IL-21R on murine CD8⁺ T cells is required to control chronic viral infection [55]. Interestingly, AD-HIES patients were found to have impaired control of reactivation of chronic infection by EBV and varicella [11^{**},12^{*}]. Thus, it is possible that defective CD8⁺ T cell differentiation and

function induced by IL-21 underlies poor immunity in STAT3-deficient humans to latent herpes viruses (Figure 1). However, as IL-21R-deficient individuals are not adversely affected by herpes viruses, additional STAT3-activating cytokines are also likely to be involved in this response. Given the role that CD8⁺ T cells play in anti-tumour immunity, it is also possible that defective cytotoxicity by these cells contributes to lymphoma development in AD-HIES ([9,10,11^{**}]; Figure 1).

Conclusions

The discovery of molecular lesions that cause primary immunodeficiencies provide the opportunity to identify specific functions of genes in different cell lineages, thereby defining the pathogenesis of these diseases. This is indeed the case for AD-HIES, with defined defects in CD4⁺ T cells, B cells and CD8⁺ T cells due to STAT3 deficiency potentially contributing to infection with specific pathogens, defective humoral immunity and susceptibility to lymphoma, respectively, in affected individuals. However, many issues remain unaddressed, such as the cause of hyper-IgE and the non-immune features of AD-HIES, as well as the impact of *STAT3* mutations on other human hematopoietic cell lineages such as NK cells and semi-invariant T cells, granulocytes and myeloid cells. This is particularly relevant as IL-6 and IL-23 can induce ROR γ t and IL-17 in human and murine $\gamma\delta$ T cells [56–58] and neutrophils [59], which plays important roles in host defence against fungal and bacterial infections that are common in AD-HIES [9,10]. Furthermore, as there are some clinical commonalities between *STAT3* loss-of function and *STAT1* gain-of function mutations (Table 1, Refs [5,6]), it is clear that the net outcome of lymphocyte activation is not simply the result of linear signalling through one predominant STAT but rather complex interactions between numerous STATs that not only complement but also antagonise each others function. Ongoing detailed molecular, cellular and biochemical analyses of these patients will hopefully elucidate these interactions further and yield improved therapies for these disorders.

Acknowledgement

Research performed in the authors' labs is supported by grants and fellowships awarded by the National Health and Medical Research Council of Australia.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

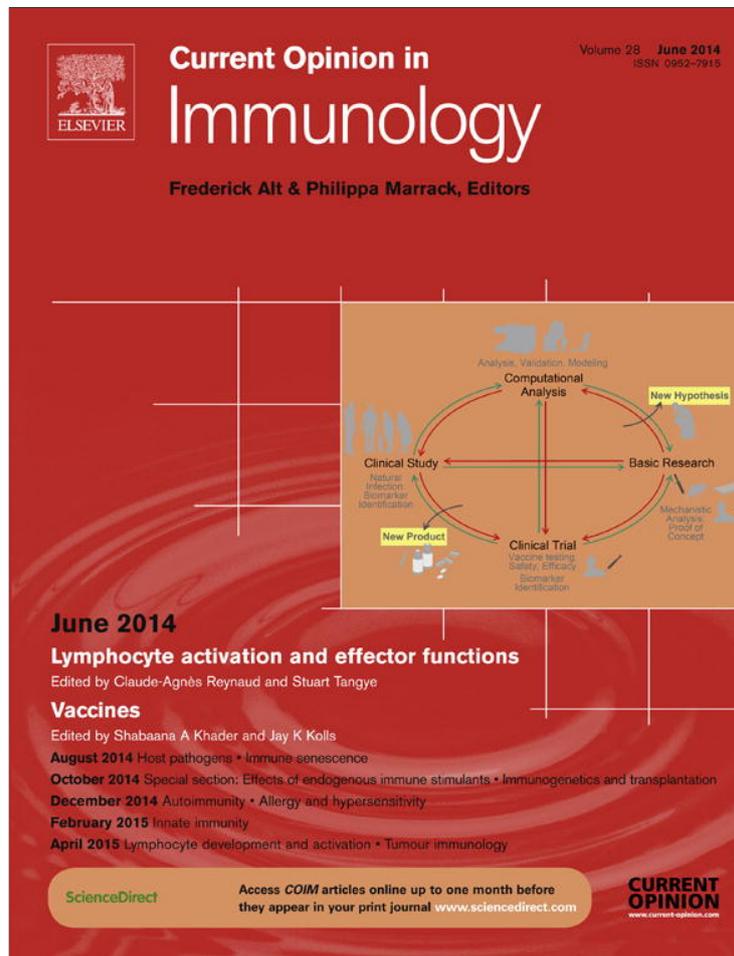
1. Levy DE, Darnell JE Jr: **Stats: transcriptional control and biological impact.** *Nat Rev Mol Cell Biol* 2002, **3**:651-662.
2. Shuai K, Liu B: **Regulation of JAK-STAT signalling in the immune system.** *Nat Rev Immunol* 2003, **3**:900-911.
3. O'Shea JJ, Holland SM, Staudt LM: **JAKs and STATs in immunity, immunodeficiency, and cancer.** *N Engl J Med* 2013, **368**:161-170.

56 Lymphocyte activation and effector functions

4. Al-Herz W, Bousfiha A, Casanova JL, Chapel H, Conley ME, Cunningham-Rundles C, Etzioni A, Fischer A, Franco JL, Geha RS *et al.*: **Primary immunodeficiency diseases: an update on the classification from the international union of immunological societies expert committee for primary immunodeficiency.** *Front Immunol* 2011, **2**:54.
5. Casanova JL, Holland SM, Notarangelo LD: **Inborn errors of human JAKs and STATs.** *Immunity* 2012, **36**:515-528.
6. Milner JD, Holland SM: **The cup runneth over: lessons from the ever-expanding pool of primary immunodeficiency diseases.** *Nat Rev Immunol* 2013, **13**:635-648.
7. Davis SD, Schaller J, Wedgwood RJ: **Job's syndrome. Recurrent, "cold", staphylococcal abscesses.** *Lancet* 1966, **1**:1013-1015.
8. Buckley RH, Wray BB, Belmaker EZ: **Extreme hyperimmunoglobulinemia E and undue susceptibility to infection.** *Pediatrics* 1972, **49**:59-70.
9. Freeman AF, Holland SM: **The hyper-IgE syndromes.** *Immunol Allergy Clin North Am* 2008, **28**:277-291 viii.
10. Grimbacher B, Holland SM, Puck JM: **Hyper-IgE syndromes.** *Immunol Rev* 2005, **203**:244-250.
11. Chandesris MO, Melki I, Natividad A, Puel A, Fieschi C, Yun L, Thumerelle C, Oksenhendler E, Boutboul D, Thomas C *et al.*: **Autosomal dominant STAT3 deficiency and hyper-IgE syndrome: molecular, cellular, and clinical features from a French national survey.** *Medicine* 2012, **91**:e1-e19.
- Provides a detailed overview of the clinical manifestations of STAT3 deficiency in humans.
12. Siegel AM, Heimall J, Freeman AF, Hsu AP, Brittain E, Brechley JM, Douek DC, Fahle GH, Cohen JL, Holland SM *et al.*: **A critical role for STAT3 transcription factor signaling in the development and maintenance of human T cell memory.** *Immunity* 2011, **35**:806-818.
- Together with Refs [53*,54*], these studies delineated defects in CD8⁺ T cell differentiation and function in humans and mice lacking STAT3.
13. Holland SM, DeLeo FR, Elloumi HZ, Hsu AP, Uzel G, Brodsky N, Freeman AF, Demidowich A, Davis J, Turner ML *et al.*: **STAT3 mutations in the hyper-IgE syndrome.** *N Engl J Med* 2007, **357**:1608-1619.
14. Minegishi Y, Saito M, Tsuchiya S, Tsuge I, Takada H, Hara T, Kawamura N, Ariga T, Pasic S, Stojkovic O *et al.*: **Dominant-negative mutations in the DNA-binding domain of STAT3 cause hyper-IgE syndrome.** *Nature* 2007, **448**:1058-1062.
- Refs [13**,14**] identified mutations in STAT3 as the molecular cause of autosomal dominant hyper IgE syndrome.
15. Takeda K, Noguchi K, Shi W, Tanaka T, Matsumoto M, Yoshida N, Kishimoto T, Akira S: **Targeted disruption of the mouse Stat3 gene leads to early embryonic lethality.** *Proc Natl Acad Sci U S A* 1997, **94**:3801-3804.
16. Deenick EK, Ma CS, Brink R, Tangye SG: **Regulation of T follicular helper cell formation and function by antigen presenting cells.** *Curr Opin Immunol* 2011, **23**:111-118.
17. O'Shea JJ, Paul WE: **Mechanisms underlying lineage commitment and plasticity of helper CD4⁺ T cells.** *Science* 2010, **327**:1098-1102.
18. Yamane H, Paul WE: **Early signaling events that underlie fate decisions of naive CD4(+) T cells toward distinct T-helper cell subsets.** *Immunol Rev* 2013, **252**:12-23.
19. Harris TJ, Grosso JF, Yen HR, Xin H, Kortylewski M, Albesiano E, Hipkiss EL, Getnet D, Goldberg MV, Maris CH *et al.*: **Cutting edge: an in vivo requirement for STAT3 signaling in TH17 development and TH17-dependent autoimmunity.** *J Immunol* 2007, **179**:4313-4317.
20. Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, Blank RB, Meylan F, Siegel R, Hennighausen L *et al.*: **Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation.** *Immunity* 2007, **26**:371-381.
21. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, Dong C: **STAT3 regulates cytokine-mediated generation of inflammatory helper T cells.** *J Biol Chem* 2007, **282**:9358-9363.
22. Nishihara M, Ogura H, Ueda N, Tsuruoka M, Kitabayashi C, Tsuji F, Aono H, Ishihara K, Huseby E, Betz UA *et al.*: **IL-6-gp130-STAT3 in T cells directs the development of IL-17+ Th with a minimum effect on that of Treg in the steady state.** *Int Immunol* 2007, **19**:695-702.
23. Ma CS, Chew GY, Simpson N, Priyadarshi A, Wong M, Grimbacher B, Fulcher DA, Tangye SG, Cook MC: **Deficiency of Th17 cells in hyper IgE syndrome due to mutations in STAT3.** *J Exp Med* 2008, **205**:1551-1557.
24. de Beaucoudrey L, Puel A, Filipe-Santos O, Cobat A, Ghandil P, Chrabieh M, Feinberg J, von Bernuth H, Samarina A, Janniére L *et al.*: **Mutations in STAT3 and IL12RB1 impair the development of human IL-17-producing T cells.** *J Exp Med* 2008, **205**:1543-1550.
25. Renner ED, Rylaarsdam S, Anover-Sombke S, Rack AL, Reichenbach J, Carey JC, Zhu Q, Jansson AF, Barboza J, Schimke LF *et al.*: **Novel signal transducer and activator of transcription 3 (STAT3) mutations, reduced T(H)17 cell numbers, and variably defective STAT3 phosphorylation in hyper-IgE syndrome.** *J Allergy Clin Immunol* 2008, **122**:181-187.
26. Cypowyj S, Picard C, Marodi L, Casanova JL, Puel A: **Immunity to infection in IL-17-deficient mice and humans.** *Eur J Immunol* 2012, **42**:2246-2254.
27. Kotlarz D, Zietara N, Uzel G, Weidemann T, Braun CJ, Diestelhorst J, Krawitz PM, Robinson PN, Hecht J, Puchalka J *et al.*: **Loss-of-function mutations in the IL-21 receptor gene cause a primary immunodeficiency syndrome.** *J Exp Med* 2013, **210**:433-443.
- The first description of human IL-21 receptor deficiency as a novel primary immunodeficiency. Study of these individuals made it possible to determine the exact role of IL-21 in immune function, and comparison to AD-HIES delineated the role of IL-21R/STAT3 signaling in host defense.
28. Wei L, Laurence A, Elias KM, O'Shea JJ: **IL-21 is produced by Th17 cells and drives IL-17 production in a STAT3-dependent manner.** *J Biol Chem* 2007, **282**:34605-34610.
29. Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, Yoshimura A, Hennighausen L, O'Shea JJ: **Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells.** *Proc Natl Acad Sci U S A* 2006, **103**:8137-8142.
30. Durant L, Watford WT, Ramos HL, Laurence A, Vahedi G, Wei L, Takahashi H, Sun HW, Kanno Y, Powrie F *et al.*: **Diverse targets of the transcription factor STAT3 contribute to T cell pathogenicity and homeostasis.** *Immunity* 2010, **32**:605-615.
31. Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, Cua DJ, Littman DR: **The orphan nuclear receptor RORgamma directs the differentiation program of proinflammatory IL-17+ T helper cells.** *Cell* 2006, **126**:1121-1133.
32. Yang XO, Pappu BP, Nurieva R, Akimzhanov A, Kang HS, Chung Y, Ma L, Shah B, Panopoulos AD, Schluns KS *et al.*: **T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR alpha and ROR gamma.** *Immunity* 2008, **28**:29-39.
33. Stritesky GL, Muthukrishnan R, Sehra S, Goswami R, Pham D, Travers J, Nguyen ET, Levy DE, Kaplan MH: **The transcription factor STAT3 is required for T helper 2 cell development.** *Immunity* 2011, **34**:39-49.
- Identified a role for STAT3 in generating Th2 cells in mice; however this is yet to be confirmed in humans.
34. Ma CS, Avery DT, Chan A, Batten M, Bustamante J, Boisson-Dupuis S, Arkwright PD, Kreins AY, Averbuch D, Engelhard D *et al.*: **Functional STAT3 deficiency compromises the generation of human T follicular helper cells.** *Blood* 2012, **119**:3997-4008.
- The first report of a Tfh defect in human STAT3 deficiency, and demonstrated that IL-12-induced differentiation of humn CD4+ T cells to Tfh-like cells required STAT3.
35. Xu L, Kitani A, Stuelten C, McGrady G, Fuss I, Strober W: **Positive and negative transcriptional regulation of the Foxp3 gene is mediated by access and binding of the Smad3 protein to enhancer I.** *Immunity* 2010, **33**:313-325.

36. Chaudhry A, Rudra D, Treuting P, Samstein RM, Liang Y, Kas A, Rudensky AY: **CD4+ regulatory T cells control TH17 responses in a Stat3-dependent manner.** *Science* 2009, **326**:986-991.
37. Saito M, Nagasawa M, Takada H, Hara T, Tsuchiya S, Agematsu K, Yamada M, Kawamura N, Ariga T, Tsuge I *et al.*: **Defective IL-10 signaling in hyper-IgE syndrome results in impaired generation of tolerogenic dendritic cells and induced regulatory T cells.** *J Exp Med* 2011, **208**:235-249.
- Reported defects in responsiveness of human DCs to IL-10 compromised their ability to induce the generation of Tregs; this impairment may contribute to immune dysregulation in AD-HIES.
38. Tangye SG, Ma CS, Brink R, Deenick EK: **The good, the bad and the ugly – TFH cells in human health and disease.** *Nat Rev Immunol* 2013, **13**:412-426.
39. Mazerolles F, Picard C, Kracker S, Fischer A, Durandy A: **Blood CD4+CD45RO+CXCR5+ T cells are decreased but partially functional in signal transducer and activator of transcription 3 deficiency.** *J Allergy Clin Immunol* 2013, **131**:1146-1156 1156 e1141-1145.
40. Deenick EK, Avery DT, Chan A, Berglund LJ, Ives ML, Moens L, Stoddard JL, Bustamante J, Boisson-Dupuis S, Tsumura M *et al.*: **Naive and memory human B cells have distinct requirements for STAT3 activation to differentiate into antibody-secreting plasma cells.** *J Exp Med* 2013, **210**:2739-2753.
- This study established that IL-21R/STAT3 signalling was required for the generation of human memory B cells *in vivo*. This also revealed that STAT3-deficient memory B cells are capable of responding to IL-10 and IL-21 as well as normal memory B cells, which completely contrasts STAT3-deficient naïve B cells.
41. Batten M, Ramamoorthi N, Kijavini NM, Ma CS, Cox JH, Dengler HS, Danilenko DM, Caplazi P, Wong M, Fulcher DA *et al.*: **IL-27 supports germinal center function by enhancing IL-21 production and the function of T follicular helper cells.** *J Exp Med* 2010, **207**:2895-2906.
42. Nurieva RI, Chung Y, Hwang D, Yang XO, Kang HS, Ma L, Wang YH, Watowich SS, Jetten AM, Tian Q *et al.*: **Generation of T follicular helper cells is mediated by interleukin-21 but independent of T helper 1, 2, or 17 cell lineages.** *Immunity* 2008, **29**:138-149.
43. Choi YS, Eto D, Yang JA, Lao C, Crotty S: **Cutting edge: STAT1 is required for IL-6-mediated Bcl6 induction for early follicular helper cell differentiation.** *J Immunol* 2013, **190**:3049-3053.
44. Leung DY, Ambrosino DM, Arbeit RD, Newton JL, Geha RS: **Impaired antibody responses in the hyperimmunoglobulin E syndrome.** *J Allergy Clin Immunol* 1988, **81**:1082-1087.
45. Sheerin KA, Buckley RH: **Antibody responses to protein, polysaccharide, and phi X174 antigens in the hyperimmunoglobulinemia E (hyper-IgE) syndrome.** *J Allergy Clin Immunol* 1991, **87**:803-811.
46. Dreskin SC, Goldsmith PK, Gallin JI: **Immunoglobulins in the hyperimmunoglobulin E and recurrent infection (Job's) syndrome. Deficiency of anti-Staphylococcus aureus immunoglobulin A.** *J Clin Invest* 1985, **75**:26-34.
47. Avery DT, Deenick EK, Ma CS, Suryani S, Simpson N, Chew GY, Chan TD, Palendira U, Bustamante J, Boisson-Dupuis S *et al.*: **B cell-intrinsic signaling through IL-21 receptor and STAT3 is required for establishing long-lived antibody responses in humans.** *J Exp Med* 2010, **207**:155-171.
48. Meyer-Bahlburg A, Renner ED, Rylaarsdam S, Reichenbach J, Schimke LF, Marks A, Tcheurekdjian H, Hostoffer R, Brahmandam A, Torgerson TR *et al.*: **Heterozygous signal transducer and activator of transcription 3 mutations in hyper-IgE syndrome result in altered B-cell maturation.** *J Allergy Clin Immunol* 2012, **129**:559-562 562 e551-552.
- Refs [47*,48*] reported defective memory B cell generation in STAT3-deficient humans. Ref [47*] also determined the requirement for STAT3 for naïve B cells to respond to IL-21 and become plasma cells by acquiring expression of Blimp-1 and XBP-1 in a STAT3-dependent manner.
49. Fornek JL, Tygrett LT, Waldschmidt TJ, Poli V, Rickert RC, Kansas GS: **Critical role for Stat3 in T-dependent terminal differentiation of IgG B cells.** *Blood* 2006, **107**:1085-1091.
50. Ettinger R, Sims GP, Fairhurst AM, Robbins R, da Silva YS, Spolski R, Leonard WJ, Lipsky PE: **IL-21 induces differentiation of human naive and memory B cells into antibody-secreting plasma cells.** *J Immunol* 2005, **175**:7867-7879.
51. Diehl SA, Schmidlin H, Nagasawa M, van Haren SD, Kwakkenbos MJ, Yasuda E, Beaumont T, Scheeren FA, Spits H: **STAT3-mediated up-regulation of BLIMP1 is coordinated with BCL6 down-regulation to control human plasma cell differentiation.** *J Immunol* 2008, **180**:4805-4815.
52. Berglund LJ, Avery DT, Ma CS, Moens L, Deenick EK, Bustamante J, Boisson-Dupuis S, Wong M, Adelstein S, Arkwright PD *et al.*: **IL-21 signalling via STAT3 primes human naive B cells to respond to IL-2 to enhance their differentiation into plasmablasts.** *Blood* 2013, **122**:3940-3950.
53. Ives ML, Ma CS, Palendira U, Chan A, Bustamante J, Boisson-Dupuis S, Arkwright PD, Engelhard D, Averbuch D, Magdorf K *et al.*: **Signal transducer and activator of transcription 3 (STAT3) mutations underlying autosomal dominant hyper-IgE syndrome impair human CD8(+) T-cell memory formation and function.** *J Allergy Clin Immunol* 2013, **132**:400-411 e409.
54. Cui W, Liu Y, Weinstein JS, Craft J, Kaech SM: **An interleukin-21-interleukin-10-STAT3 pathway is critical for functional maturation of memory CD8+ T cells.** *Immunity* 2011, **35**:792-805.
- Together with Ref [12*], these studies delineated defects in CD8+ T cell differentiation and function in humans and mice lacking STAT3.
55. Johnson LD, Jameson SC: **Immunology. A chronic need for IL-21.** *Science* 2009, **324**:1525-1526.
56. Caccamo N, La Mendola C, Orlando V, Meraviglia S, Todaro M, Stassi G, Sireci G, Fournie JJ, Dieli F: **Differentiation, phenotype, and function of interleukin-17-producing human Vgamma9Vdelta2 T cells.** *Blood* 2011, **118**:129-138.
57. Ness-Schwickerath KJ, Jin C, Morita CT: **Cytokine requirements for the differentiation and expansion of IL-17A- and IL-22-producing human Vgamma2Vdelta2 T cells.** *J Immunol* 2010, **184**:7268-7280.
58. Cho JS, Pietras EM, Garcia NC, Ramos RI, Farzam DM, Monroe HR, Magorien JE, Blauvelt A, Kolls JK, Cheung AL *et al.*: **IL-17 is essential for host defense against cutaneous Staphylococcus aureus infection in mice.** *J Clin Invest* 2010, **120**:1762-1773.
59. Taylor PR, Roy S, Leal SM Jr, Sun Y, Howell SJ, Cobb BA, Li X, Pearlman E: **Activation of neutrophils by autocrine IL-17A-IL-17RC interactions during fungal infection is regulated by IL-6, IL-23, RORgamma and dectin-2.** *Nat Immunol* (15):2013:143-151.

Provided for non-commercial research and education use.
Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

<http://www.elsevier.com/authorsrights>