

KOBAS 2.0: a web server for annotation and identification of enriched pathways and diseases

Chen Xie¹, Xizeng Mao², Jiaju Huang¹, Yang Ding¹, Jianmin Wu³, Shan Dong¹, Lei Kong¹, Ge Gao¹, Chuan-Yun Li⁴ and Liping Wei^{1,*}

¹Center for Bioinformatics, State Key Laboratory of Protein and Plant Gene Research, College of Life Sciences, Peking University, Beijing, China, ²Computational Systems Biology Lab, Department of Biochemistry and Molecular Biology, and Institute of Bioinformatics, University of Georgia, USA, ³Cancer Research Program, Garvan Institute of Medical Research, 384 Victoria Street, Darlinghurst, NSW, Australia and ⁴Laboratory of Bioinformatics and Genomic Medicine, Institute of Molecular Medicine, Peking University, Beijing, China

Received March 24, 2011; Revised May 11, 2011; Accepted May 25, 2011

ABSTRACT

High-throughput experimental technologies often identify dozens to hundreds of genes related to, or changed in, a biological or pathological process. From these genes one wants to identify biological pathways that may be involved and diseases that may be implicated. Here, we report a web server, KOBAS 2.0, which annotates an input set of genes with putative pathways and disease relationships based on mapping to genes with known annotations. It allows for both ID mapping and cross-species sequence similarity mapping. It then performs statistical tests to identify statistically significantly enriched pathways and diseases. KOBAS 2.0 incorporates knowledge across 1327 species from 5 pathway databases (KEGG PATHWAY, PID, BioCyc, Reactome and Panther) and 5 human disease databases (OMIM, KEGG DISEASE, FunDO, GAD and NHGRI GWAS Catalog). KOBAS 2.0 can be accessed at <http://kobas.cbi.pku.edu.cn>.

INTRODUCTION

High-throughput experimental technologies such as next generation sequencing, microarray profiling and proteomics profiling are widely used in current biological research and often identify dozens to hundreds of genes related to a biological or pathological process. Given such a set of genes, one wants to ask which metabolic and signaling pathways may be involved and which diseases may be implicated. As the number of genes is often large, it is desirable to have a computational tool to provide initial answers to these questions. However, *ab initio* prediction of pathways and diseases is challenging. One feasible

approach is to use existing databases of known metabolic and signaling pathways and databases of known disease-associated genes as the starting point for annotation of a new set of genes.

We have previously reported a standalone software and a web server KOBAS 1.0 (1,2) that annotates an input set of genes or proteins by mapping to genes with known pathways in the KEGG PATHWAY database (3). KOBAS 1.0 was the first software to identify statistically significantly enriched pathways using a hypergeometric test. It has been successfully used in pathway analysis in plants, animals and bacteria [for instance, (4–6)].

During the past decade, many other functional enrichment analysis tools have become available. Most of them focus on identification of enriched functional categories based on Gene Ontology (GO) (7), such as FuncAssociate (8), Ontologizer (9), BiNGO (10), FatiGO (11), GOToolBox (11) and GFinder (12). Although tremendously useful, functional categories are not as informative and intuitive as metabolic and signaling pathways and human diseases. A growing number of tools have been developed for pathway and disease identification, including, but not limited to, MAPPFinder (13), EASE (14), DAVID (15,16), ArrayXPath (17), WebGestalt (18), FuncCluster (19), PageMan (20), GENECODIS (21,22), GeneTrail (23), g:Profiler (24), FunNet (25) and PaLS (26). Except for DAVID, all these tools integrate limited pathway and disease databases (for a comparison, see Supplementary Table S1). Furthermore, none of these tools support sequence similarity mapping, an important feature that allows the user to take advantage of data from other species. It is necessary and important to develop a web server tool which incorporates comprehensive pathway and disease databases and supports both ID mapping and sequence similarity mapping.

*To whom correspondence should be addressed. Tel: +86 10 6275 5206; Fax: +86 10 6275 9001; Email: weilp@mail.cbi.pku.edu.cn

Here, we report a significantly expanded new version, KOBAS 2.0, which incorporates 5 pathway databases [KEGG PATHWAY, PID (27), BioCyc (28), Reactome (29,30) and Panther (31)] and 5 human disease databases [OMIM (<http://www.ncbi.nlm.nih.gov/omim/>), KEGG DISEASE (32), FunDO (33,34), GAD (35) and NHGRI GWAS Catalog (NHGRI) (36)]. Similar to version 1.0, KOBAS 2.0 supports not only ID mapping, but also sequence similarity mapping. KOBAS 2.0 consists of a standalone command line program written in Python which runs on most Linux systems as well as a user friendly web server developed using Java. Both command line program and web server are freely available at <http://kobas.cbi.pku.edu.cn>. KOBAS 2.0 flowchart is summarized in Figure 1 and detailed below.

MATERIALS AND METHODS

KOBAS 2.0 parses 10 pathway and disease databases and stores the data in a SQL relational database

Table 1 summarizes information about the pathway and disease databases that KOBAS 2.0 incorporates. Specifically, KEGG PATHWAY (3) and Reactome (29,30) are general pathway databases, whereas PID (27) and Panther (31) focus on signaling pathways and BioCyc (28) focuses on metabolic pathways. PID has only human data, whereas the others are multispecies databases. OMIM (<http://www.ncbi.nlm.nih.gov/omim/>) contains information on all known mendelian disorders and genes. KEGG DISEASE (32) collects knowledge on genetic and environmental factors of diseases. FunDO (33,34) is generated from GeneRIF using Disease Ontology Lite that is a condensed version of Disease Ontology. GAD (35) and NHGRI GWAS Catalog (36) both collect data from genetic association studies: GAD includes data from both candidate genes and GWAS studies, whereas NHGRI GWAS Catalog is a catalog of only GWAS studies.

KOBAS 2.0 downloaded the raw data files from each database. As shown in Table 1, the file formats include

plain text, XML and table. We have written parsers for all the data files. For each pathway or disease database, we retrieve the gene-term mapping by parsing the raw data files. We retrieve the gene annotation and gene-ID relations from KEGG Genes and BioMart (37). To integrate across different databases, we mapped the genes in all databases to KEGG GENES and KEGG ORTHOLOGY (KO). The gene-pathway and gene-disease data is stored in our backend SQL relational database. The FASTA protein sequence files were preprocessed for BLAST. KOBAS 2.0 backend data is updated every 3 months.

KOBAS 2.0 annotates input genes with pathways and diseases and identifies enriched pathways and diseases

KOBAS 2.0 has two consecutive programs 'annotate' and 'identify', which is similar to KOBAS 1.0 (1,2). The first program 'annotates' each input gene with putative pathways and diseases by mapping the gene to genes in KEGG GENES or terms in KO which are linked to pathway and disease terms in backend databases. For ID mapping, input IDs are mapped directly to genes using the cross-links we parsed from KEGG GENES. Then, if necessary, IDs are mapped to KO terms. For sequence similarity mapping, each input sequence is BLASTed against all sequences in KEGG GENES. The default cutoffs are BLAST E -value $<10^{-5}$ and rank ≤ 5 . They mean that an input sequence is assigned KO term(s) of the first BLAST hit that (i) has known KO assignments; (ii) has BLAST E -value $<10^{-5}$; and (iii) has less than five other hits with a lower E -value that do not have KO assignments (1). A new option in KOBAS 2.0 is that users can map against genes in user-specified species instead of all genes by BLASTing against only sequences of the user-specified species. In order to reduce possible false positives due to multidomain proteins, we added a new option to allow users to set a cutoff of BLAST subject coverage. Another new option allows users to restrict sequence mapping to only orthologs as defined by Ensembl Compara (38).

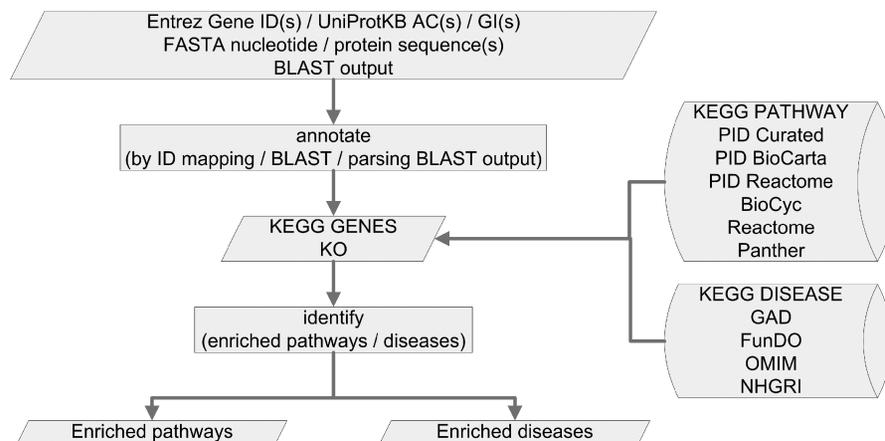


Figure 1. KOBAS 2.0 workflow. The types of input can be ID, FASTA sequence, or tabular BLAST output. KOBAS 2.0 has two programs 'annotate' and 'identify'. The first program annotates input genes with pathways and diseases by ID mapping or sequence similarity mapping. The second program identifies statistically significantly enriched pathways and diseases.

Table 1. Pathway and disease databases supported by KOBAS 2.0^a

Database name	Data content	File format	Number of species	Number of pathways or diseases in human	Number of genes mapped to KEGG GENES/all genes in human	URL
KEGG PATHWAY	Pathway	Text	1327	220	5595/5595	http://www.genome.jp/kegg/pathway.html
PID Curated	Pathway	XML	1	192	2782/3315	http://pid.nci.nih.gov/
PID BioCarta	Pathway	XML	1	254	1907/2391	http://pid.nci.nih.gov/
PID Reactome	Pathway	XML	1	996	3783/4405	http://pid.nci.nih.gov/
BioCyc	Pathway	Text and Table	6	277	1087/1120	http://biocyc.org/
Reactome	Pathway	Table	22	68	4366/4534	http://www.reactome.org/ReactomeGWT/entrypoint.html
Panther	Pathway	Table	43	154	2170/2207	http://www.pantherdb.org/
OMIM	Disease	Table	1	4990	3792/3792	http://www.ncbi.nlm.nih.gov/omim
KEGG DISEASE	Disease	Text	1	323	798/798	http://www.genome.jp/kegg/disease/
FunDO	Disease	Table	1	561	3888/4029	http://django.nubic.northwestern.edu/fundo/
GAD	Disease	Table	1	3770	3164/3238	http://geneticassociationdb.nih.gov/
NHGRI	Disease	Table	1	369	1975/2191	http://www.genome.gov/gwastudies/

^aThe numbers in this table are summarized from KOBAS 2.0 backend database updated in November 23rd, 2010. And all the analyses using KOBAS 2.0 in this article are based on this data version.

The second program ‘identifies’ statistically significantly enriched pathways and diseases by comparing results from the first program against the background (usually genes from the whole genome, or all probe sets on a microarray). Users can define their own background distribution in KOBAS 2.0 (for example, result from the first program to ‘annotate’ all probe sets on a microarray). If users do not upload a background file, KOBAS 2.0 uses the genes from whole genome as the default background distribution. Here, we consider only pathways and diseases for which there are at least two genes mapped in the input. Users can choose to perform statistical test using one of the following four methods: binomial test, chi-square test, Fisher’s exact test and hypergeometric test, and perform FDR correction. The purpose of performing FDR correction is to reduce the Type-1 errors. When a large number of pathway and disease terms are considered, multiple hypotheses tests are performed, which leads to a high overall Type-1 error even for a relatively stringent *P*-value cutoff. KOBAS 1.0 supports the FDR correction method QVALUE (39). In KOBAS 2.0, we add two more popular FDR correction methods: Benjamini-Hochberg (40) and Benjamini-Yekutieli (41).

INPUT AND OUTPUT

Input

The input to ‘annotate’ can be a list of IDs, a FASTA sequence file or a tabular BLAST output. KOBAS 2.0 currently can accept three kinds of IDs: Entrez Gene ID, UniProtKB AC and GI. FASTA sequences can be protein or nucleotide sequences. Because BLAST is computationally intensive, the number of sequences that can be run on the online web server is limited to 500 per run. A new feature in KOBAS 2.0 is that, if users want to annotate

more sequences online, they can run BLAST locally and upload the tabular BLAST output as the input to KOBAS 2.0. Or they can always run the standalone version of KOBAS 2.0 which has no limit. If users want to get the pathway and disease annotations of their genes, they only need to run ‘annotate’. If they want to find enriched pathways and diseases, they can feed the output of ‘annotate’ directly into ‘identify’ as input.

Output

The example of the output of ‘annotate’ is shown in Figure 2. Each row corresponds to one input gene. The first column contains the input gene IDs. The second and third columns contain the mapped KEGG GENE IDs, hyperlinked to detailed descriptions in KEGG and the mapped KEGG GENE names. A user can click on ‘details’ next to the input gene ID to see details about the query and related pathways and diseases.

The examples of the output of ‘identify’ is shown in Figure 3. KOBAS 2.0 separates the results of pathways and diseases into two tables. In the pathway identification result, the first three columns show the pathway name, pathway database and pathway ID, hyperlinked to detailed description in the corresponding database. The fourth column lists two numbers of the input: the first one is the number of input genes mapped to the particular pathway and the second one is the total number of input genes mapped to any pathway in the pathway database. Users can click on the first number in the fourth column to see the list of input genes mapped to the particular pathway. The fifth column lists two numbers of the background: the first one is the number of background genes mapped to the particular pathway and the second one is the total number of background genes mapped to any pathway in the pathway database. The last two columns

Result of file: upCA.a

DOWNLOAD... HELP USE THIS FILE AS IDENTIFY'S SAMPLE INPUT

RAW CONTENT TABLE VIEW

130 succeed, 241 fail

Query	Gene ID	Gene Name
MmugDNA.36383.1.S1_at (details)	hsa:55890	GPRC5C, MGC131820, RAIG-3, RAIG3
MmugDNA.8781.1.S1_at (details)	hsa:1297	COL9A1, DJ149L1.1.2, EDM6, FLJ40263, MED
MmugDNA.15232.1.S1_at (details)	None	None
Mmu.12320.1.S1_at (details)	hsa:2109	ETFB, MADD
MmugDNA.36986.1.S1_at (details)	None	None
MmugDNA.1883.1.S1_at (details)	None	None
MmuSTS.1124.1.S1_at (details)	None	None
MmugDNA.12821.1.S1_at (details)	None	None
MmugDNA.30787.1.S1_at (details)	None	None
MmugDNA.39136.1.S1_at (details)	hsa:2109	ETFB, MADD
MmuSTS.4242.1.S1_at (details)	None	None
MmuSTS.4335.1.S1_at (details)	hsa:84893	FBXO18, FBH1, FLJ14590, Fbx18, MGC131916, MGC141935, MGC141937
MmugDNA.14249.1.S1_s_at (details)	hsa:746	C11orf10
MmugDNA.14828.1.S1_at (details)	None	None
MmugDNA.37451.1.S1_at (details)	hsa:51076	CUTC, RP11-483F11.3
MmuSTS.3730.1.S1_at (details)	None	None
MmugDNA.3015.1.S1_at (details)	hsa:5436	POLR2G, MGC138367, MGC138369, RPB19, RPB7, hRPB19, hsRPB7
MmugDNA.29430.1.S1_at (details)	None	None
MmugDNA.9338.1.S1_at (details)	hsa:28989	METTL11A, AD-003, C9orf32
MmugDNA.29915.1.S1_at (details)	hsa:5705	PSMC5, S8, SUG-1, SUG1, TBP10, TRIP1, p45, p45/SUG

Displaying 1 ~ 20 of 371 Page 1 of 19

Figure 2. Screenshot of the output of 'annotate'. 371 upregulated probe sets in CA are assigned to KEGG human genes by sequence similarity mapping. Users can view the result in table format (by default) or raw format (which can be downloaded to local disks). Users can also directly use the result as the input of 'identify' to do further analysis.

list the P -value and corrected P -value of the statistical test. In the disease identification result, the seven columns show the disease name, disease database, disease ID, numbers of the input, numbers of the background, P -value and corrected P -value similar to the pathway identification result. KOBAS 2.0 merges redundant pathway and disease terms from different databases.

BENEFIT OF CROSS-SPECIES SEQUENCE SIMILARITY MAPPING OVER ID MAPPING

Other existing pathway analysis tools accept only gene IDs as input and use only ID mapping to annotate their pathways. A benefit of KOBAS 2.0 is that it can use sequence similarity mapping to annotate input genes from species that are not yet well-represented in existing pathway databases. It can also map the genes from other species to human diseases to predict whether these genes may be good candidates to study any human diseases, an important question in the model organism research. To illustrate, we analyzed the microarray expression profiles in rhesus monkeys in two major hippocampal subdivisions critical for memory/cognitive function: cornu ammonis (CA) and dentate gyrus (DG) using data from Blalock *et al.* (42). We reanalyzed their raw data on six samples

from CA and six samples from DG of young rhesus monkeys and identified 371 upregulated probe sets in CA using standard protocol [germa and limma through R and Bioconductor (43)]. We then used both DAVID (15,16) and KOBAS 2.0 to annotate these probe sets and identify enriched pathways and diseases by using the entire probe sets on the chip as background. DAVID can perform only ID mapping to rhesus genes in its two pathway databases (KEGG PATHWAY and Panther) and as a result, identified no statistically significantly enriched pathways or diseases (with default options and corrected $P \leq 0.05$). On the other hand, KOBAS 2.0 supports sequence similarity mapping by BLAST to annotate the rhesus gene set and can thus take full advantage of the abundant data on human pathways and diseases. We used 'annotate' to map sequences of upregulated probe sets in CA as well as the entire probe sets to KEGG human genes with default cutoffs and then used 'identify' to perform hypergeometric test and Benjamini-Hochberg FDR correction to find significantly enriched pathways and diseases by using the two results of 'annotate' as input and background, respectively. Figure 3 shows significantly enriched pathways and diseases identified by KOBAS 2.0. The results are consistent with known functional differences between the two regions. For example,

Result of file: upCA.BH.i

DOWNLOAD... HELP

RAW CONTENT **TABLE VIEW (FOR PATHWAY IDENTIFICATION RESULT)** TABLE VIEW (FOR DISEASE IDENTIFICATION RESULT)

Term	Database	ID	Sample Number (click to sort; click again to togg	Background Number	PValue (click to sort; click again to toggle so	Corrected PValue (click to sort; click again to toggle s Cannot sort when the
Respiratory electron transport, ATP synth	Reactome	REACT:6305	8 / 41	129 / 4092	0.0000312347300064	0.00284236043059
Alzheimer's disease	KEGG PATHWAY	hsa05010	8 / 47	189 / 5014	0.000312305371425	0.00947326293322
Huntington's disease	KEGG PATHWAY	hsa05016	8 / 47	220 / 5014	0.000864088709773	0.0196580181473
no2-dependent il-12 pathway in nk cells	PID BioCarta	100093	2 / 6	12 / 1151	0.00146145188136	0.0221653535339
Gene Expression	Reactome	REACT:71	12 / 41	501 / 4092	0.00274836840139	0.0284735432448
il12 and stat4 dependent signaling pathw	PID BioCarta	100133	2 / 6	17 / 1151	0.00297654241314	0.0284735432448
Oxidative phosphorylation	KEGG PATHWAY	hsa00190	6 / 47	159 / 5014	0.00338578203532	0.0284735432448
Respiratory electron transport	PID Reactome	500282	3 / 21	30 / 1909	0.00385051732382	0.0284735432448
Parkinson's disease	KEGG PATHWAY	hsa05012	6 / 47	165 / 5014	0.00406764903497	0.0284735432448
Diabetes pathways	Reactome	REACT:15380	9 / 41	349 / 4092	0.00647169563672	0.0420660216387
Metabolism of proteins	Reactome	REACT:17015	9 / 41	355 / 4092	0.00722985049746	0.0438610930179

Displaying 1 - 20 of 61 Page 1 of 4

Result of file: upCA.BH.i

DOWNLOAD... HELP

RAW CONTENT **TABLE VIEW (FOR PATHWAY IDENTIFICATION RESULT)** **TABLE VIEW (FOR DISEASE IDENTIFICATION RESULT)**

Term	Database	ID	Sample Number (click to sort; click again to togg	Background Number	PValue (click to sort; click again to toggle sc	Corrected PValue (click to sort; click again to toggle s Cannot sort when the
Glutaricaciduria, type IIB	OMIM	None	2 / 23	2 / 2313	0.000094620895585	0.00430525074912
Glutaric acidemia	KEGG DISEASE	H00178	2 / 8	6 / 814	0.00124446233735	0.0221653535339
menarche (age at onset)	GAD	None	2 / 23	8 / 2795	0.00176042663165	0.0228855462115
Autoimmune disease	FunDO	None	3 / 13	91 / 3507	0.00400335198312	0.0284735432448

Displaying 1 - 20 of 30 Page 1 of 2

Figure 3. Screenshot of the output of 'identify'. Statistically significantly enriched pathways and diseases of 371 upregulated probe sets in CA identified are sorted by increasing corrected *P*-value. Only those with corrected $P \leq 0.05$ are shown. Similar to the output of 'annotate', users can view the result in table format (by default) or raw format (which can be downloaded to local disks).

'respiratory electron transport, ATP synthesis by chemiosmotic coupling and heat production by uncoupling proteins' pathway and 'glutaricaciduria, type IIB' and 'Glutaric acidemia' diseases are consistent with the known knowledge that the CA region showed greater expression than DG for genes associated with mitochondrial activity (42); while 'no2-dependent il-12 pathway in nk cells', 'il12 and stat4 dependent signaling pathway in th1 development' and 'autoimmune disease' are consistent with the known knowledge that CA region showed greater expression than DG for genes associated with inflammatory responses (42).

We also compared KOBAS 2.0 with popular GO enrichment analysis tools, FuncAssociate 2.0 (8), Ontologizer 2.0 (9), BiNGO (10) and EASE (14) using the same data set. Because these other tools can only take IDs as input, we first mapped the rhesus probe sets to human genes using sequence similarity. Then we ran the four GO enrichment analysis tools, the results of which

are shown in Supplementary Table S2. The list of enriched pathways identified by KOBAS 2.0 is more specific and informative than the lists of functional categories identified by the GO enrichment analysis tools, and offers more insights into the biological processes.

CONCLUSIONS

KOBAS 2.0 has an expanded reservoir of underlying pathway databases and statistical tests, and the addition of disease databases. In future research, we aim to improve the graphical representation of the output pathways. We will continue to update KOBAS 2.0 with new pathway and disease data.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

FUNDING

'National Outstanding Young Investigator' from Natural Science Foundation of China (31025014); Johnson and Johnson (scholarship); China Ministry of Science and Technology 863 Hi-Tech Research and Development Programs (2007AA02Z165) and 973 Basic Research Program (2011CBA01102, 2007CB946904). Funding for open access charge: 973 Basic Research Program (2011CBA01102).

Conflict of interest statement. None declared.

REFERENCES

- Mao, X., Cai, T., Olyarchuk, J.G. and Wei, L. (2005) Automated genome annotation and pathway identification using the KEGG Orthology (KO) as a controlled vocabulary. *Bioinformatics*, **21**, 3787–3793.
- Wu, J., Mao, X., Cai, T., Luo, J. and Wei, L. (2006) KOBAS server: a web-based platform for automated annotation and pathway identification. *Nucleic Acids Res.*, **34**, W720–W724.
- Kanehisa, M. and Goto, S. (2000) KEGG: kyoto encyclopedia of genes and genomes. *Nucleic Acids Res.*, **28**, 27–30.
- Shi, Y.H., Zhu, S.W., Mao, X.Z., Feng, J.X., Qin, Y.M., Zhang, L., Cheng, J., Wei, L.P., Wang, Z.Y. and Zhu, Y.X. (2006) Transcriptome profiling, molecular biological, and physiological studies reveal a major role for ethylene in cotton fiber cell elongation. *Plant Cell*, **18**, 651–664.
- Huang, J., Chen, T., Liu, X., Jiang, J., Li, J., Li, D., Liu, X.S., Li, W., Kang, J. and Pei, G. (2009) More synergetic cooperation of Yamanaka factors in induced pluripotent stem cells than in embryonic stem cells. *Cell Res.*, **19**, 1127–1138.
- Sridhar, J. and Rafi, Z.A. (2008) Functional annotations in bacterial genomes based on small RNA signatures. *Bioinformatics*, **2**, 284–295.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T. et al. (2000) Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat. Genet.*, **25**, 25–29.
- Berriz, G.F., Beaver, J.E., Cenik, C., Tasan, M. and Roth, F.P. (2009) Next generation software for functional trend analysis. *Bioinformatics*, **25**, 3043–3044.
- Bauer, S., Grossmann, S., Vingron, M. and Robinson, P.N. (2008) Ontologizer 2.0—a multifunctional tool for GO term enrichment analysis and data exploration. *Bioinformatics*, **24**, 1650–1651.
- Maere, S., Heymans, K. and Kuiper, M. (2005) BiNGO: a Cytoscape plugin to assess overrepresentation of gene ontology categories in biological networks. *Bioinformatics*, **21**, 3448–3449.
- Al-Shahrour, F., Diaz-Urriarte, R. and Dopazo, J. (2004) FatiGO: a web tool for finding significant associations of Gene Ontology terms with groups of genes. *Bioinformatics*, **20**, 578–580.
- Masseroli, M., Martucci, D. and Pinciroli, F. (2004) GFINDER: Genome Function INtegrated Discoverer through dynamic annotation, statistical analysis, and mining. *Nucleic Acids Res.*, **32**, W293–W300.
- Salomonis, N., Hanspers, K., Zamboni, A.C., Vranizan, K., Lawlor, S.C., Dahlquist, K.D., Doniger, S.W., Stuart, J., Conklin, B.R. and Pico, A.R. (2007) GenMAPP 2: new features and resources for pathway analysis. *BMC Bioinformatics*, **8**, 217.
- Hosack, D.A., Dennis, G. Jr, Sherman, B.T., Lane, H.C. and Lempicki, R.A. (2003) Identifying biological themes within lists of genes with EASE. *Genome Biol.*, **4**, R70.
- Huang da, W., Sherman, B.T. and Lempicki, R.A. (2009) Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res.*, **37**, 1–13.
- Huang da, W., Sherman, B.T. and Lempicki, R.A. (2009) Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat. Protoc.*, **4**, 44–57.
- Chung, H.J., Park, C.H., Han, M.R., Lee, S., Ohn, J.H., Kim, J. and Kim, J.H. (2005) ArrayXPath II: mapping and visualizing microarray gene-expression data with biomedical ontologies and integrated biological pathway resources using Scalable Vector Graphics. *Nucleic Acids Res.*, **33**, W621–W626.
- Zhang, B., Kirov, S. and Snoddy, J. (2005) WebGestalt: an integrated system for exploring gene sets in various biological contexts. *Nucleic Acids Res.*, **33**, W741–W748.
- Henegar, C., Cancelli, R., Rome, S., Vidal, H., Clement, K. and Zucker, J.D. (2006) Clustering biological annotations and gene expression data to identify putatively co-regulated biological processes. *J. Bioinform. Comput. Biol.*, **4**, 833–852.
- Usadel, B., Nagel, A., Steinhäuser, D., Gibon, Y., Blasing, O.E., Redestig, H., Sreenivasulu, N., Krall, L., Hannah, M.A., Poree, F. et al. (2006) PageMan: an interactive ontology tool to generate, display, and annotate overview graphs for profiling experiments. *BMC Bioinformatics*, **7**, 535.
- Carmona-Saez, P., Chagoyen, M., Tirado, F., Carazo, J.M. and Pascual-Montano, A. (2007) GENECODIS: a web-based tool for finding significant concurrent annotations in gene lists. *Genome Biol.*, **8**, R3.
- Nogales-Cadenas, R., Carmona-Saez, P., Vazquez, M., Vicente, C., Yang, X., Tirado, F., Carazo, J.M. and Pascual-Montano, A. (2009) GeneCodis: interpreting gene lists through enrichment analysis and integration of diverse biological information. *Nucleic Acids Res.*, **37**, W317–W322.
- Backes, C., Keller, A., Kuentzer, J., Kneissl, B., Comtesse, N., Elnakady, Y.A., Müller, R., Meese, E. and Lenhof, H.P. (2007) GeneTrail—advanced gene set enrichment analysis. *Nucleic Acids Res.*, **35**, W186–W192.
- Reimand, J., Kull, M., Peterson, H., Hansen, J. and Vilo, J. (2007) g:Profiler—a web-based toolset for functional profiling of gene lists from large-scale experiments. *Nucleic Acids Res.*, **35**, W193–W200.
- Prifti, E., Zucker, J.D., Clement, K. and Henegar, C. (2008) FunNet: an integrative tool for exploring transcriptional interactions. *Bioinformatics*, **24**, 2636–2638.
- Alibes, A., Canada, A. and Diaz-Urriarte, R. (2008) PaLS: filtering common literature, biological terms and pathway information. *Nucleic Acids Res.*, **36**, W364–W367.
- Schaefer, C.F., Anthony, K., Krupa, S., Buchoff, J., Day, M., Hannay, T. and Buetow, K.H. (2009) PID: the Pathway Interaction Database. *Nucleic Acids Res.*, **37**, D674–D679.
- Karp, P.D., Ouzounis, C.A., Moore-Kochlacs, C., Goldovsky, L., Kaipa, P., Ahren, D., Tsoka, S., Darzentas, N., Kunin, V. and Lopez-Bigas, N. (2005) Expansion of the BioCyc collection of pathway/genome databases to 160 genomes. *Nucleic Acids Res.*, **33**, 6083–6089.
- Matthews, L., Gopinath, G., Gillespie, M., Caudy, M., Croft, D., de Bono, B., Garapati, P., Hemish, J., Hermjakob, H., Jassal, B. et al. (2009) Reactome knowledgebase of human biological pathways and processes. *Nucleic Acids Res.*, **37**, D619–D622.
- Croft, D., O'Kelly, G., Wu, G., Haw, R., Gillespie, M., Matthews, L., Caudy, M., Garapati, P., Gopinath, G., Jassal, B. et al. (2011) Reactome: a database of reactions, pathways and biological processes. *Nucleic Acids Res.*, **39**, D691–D697.
- Thomas, P.D., Campbell, M.J., Kejariwal, A., Mi, H., Karlak, B., Daverman, R., Diemer, K., Muruganujan, A. and Narechania, A. (2003) PANTHER: a library of protein families and subfamilies indexed by function. *Genome Res.*, **13**, 2129–2141.
- Kanehisa, M., Goto, S., Furumichi, M., Tanabe, M. and Hirakawa, M. (2010) KEGG for representation and analysis of molecular networks involving diseases and drugs. *Nucleic Acids Res.*, **38**, D355–D360.
- Osborne, J.D., Flatow, J., Holko, M., Lin, S.M., Kibbe, W.A., Zhu, L.J., Danila, M.I., Feng, G. and Chisholm, R.L. (2009) Annotating the human genome with Disease Ontology. *BMC Genomics*, **10**(Suppl 1), S6.
- Du, P., Feng, G., Flatow, J., Song, J., Holko, M., Kibbe, W.A. and Lin, S.M. (2009) From disease ontology to disease-ontology lite: statistical methods to adapt a general-purpose ontology for the test of gene-ontology associations. *Bioinformatics*, **25**, i63–i68.
- Becker, K.G., Barnes, K.C., Bright, T.J. and Wang, S.A. (2004) The genetic association database. *Nat. Genet.*, **36**, 431–432.

36. Hindorf,L.A., Sethupathy,P., Junkins,H.A., Ramos,E.M., Mehta,J.P., Collins,F.S. and Manolio,T.A. (2009) Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc. Natl Acad. Sci. USA*, **106**, 9362–9367.
37. Haider,S., Ballester,B., Smedley,D., Zhang,J., Rice,P. and Kasprzyk,A. (2009) BioMart Central Portal—unified access to biological data. *Nucleic Acids Res.*, **37**, W23–W27.
38. Kersey,P.J., Lawson,D., Birney,E., Derwent,P.S., Haimel,M., Herrero,J., Keenan,S., Kerhornou,A., Koscielny,G., Kahari,A. *et al.* (2010) Ensembl Genomes: extending Ensembl across the taxonomic space. *Nucleic Acids Res.*, **38**, D563–D569.
39. Storey,J.D. (2002) A direct approach to false discovery rates. *J. R. Statist. Soc. B.*, **64**, 479–498.
40. Benjamini,Y. and Hochberg,Y. (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J. R. Statist. Soc. B.*, **57**, 289–300.
41. Benjamini,Y. and Yekutieli,D. (2001) The control of the false discovery rate in multiple testing under dependency. *Ann. Statist.*, **29**, 1165–1188.
42. Blalock,E.M., Grondin,R., Chen,K.C., Thibault,O., Thibault,V., Pandya,J.D., Dowling,A., Zhang,Z., Sullivan,P., Porter,N.M. *et al.* (2010) Aging-related gene expression in hippocampus proper compared with dentate gyrus is selectively associated with metabolic syndrome variables in rhesus monkeys. *J. Neurosci.*, **30**, 6058–6071.
43. Gentleman,R.C., Carey,V.J., Bates,D.M., Bolstad,B., Dettling,M., Dudoit,S., Ellis,B., Gautier,L., Ge,Y., Gentry,J. *et al.* (2004) Bioconductor: open software development for computational biology and bioinformatics. *Genome Biol.*, **5**, R80.