



## The roles of TNF in brain dysfunction and disease

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### ABSTRACT

Certain cytokines, the prototype being the highly pleiotropic TNF, have many homeostatic physiological roles, are involved in innate immunity, and cause inflammation when in excess. These cytokines have long been accepted to have central roles in the pathogenesis of systemic or local non-cerebral disease states, whether acute or chronic, and whether or not caused by infectious agents. Over the last decade they have also been appreciated to be broadly important in brain physiology. As in other organs, excessive levels in brain are harmful, and its physiological complexity leads to correspondingly complex dysfunction. This review summarizes the burgeoning literature on this topic, and how the functions of these molecules, particularly TNF, are influencing the outlook of researchers on the pathophysiology of these diseases. Basic brain physiology is thus informing knowledge of the brain dysfunction that characterizes such apparently diverse states as Alzheimer's disease, trauma (mostly, but not only, to the brain), Parkinson's disease, and severe systemic infectious states, including malaria, sepsis, viral diseases and major depression. The implication is that the anti-cytokine therapies now in use, typically directed at TNF, warrant testing in these diseases in circumstances in which the therapeutic agent enters the cerebrospinal fluid. Routinely administering such drugs to patients exhibiting the neurological changes discussed in this review would simply add another organ system to what is already a very successful strategy in the treatment of inflammatory disease at other sites, such as joints, skin and gut. Clearly, the most relevant research is focussed on Alzheimer's disease, but the principles may also apply to other encephalopathies.

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**Abbreviations:** A $\beta$ , amyloid beta; AGE, advanced glycation end products; AMPA, alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; APP, amyloid  $\beta$  (A4) precursor protein; CO, carbon monoxide; CORM-3, tricarbonylchloro(glycinato)ruthenium (II); CSF, cerebrospinal fluid; GABA, gamma-aminobutyric acid; H<sub>2</sub>S, hydrogen sulfide; HMBG1, high mobility group box 1 protein; HO-1, hemoxygenase-1; i. c. v., intracerebroventricular; IL-1, interleukin-1; iNOS, inducible nitric oxide synthase; IFN- $\gamma$ , interferon- $\gamma$ ; LPS, lipopolysaccharide; LTP, long-term potentiation; LTs, lymphotoxins; NMDA, N-methyl-D-aspartate; nNOS, neuronal nitric oxide synthase; NO, nitric oxide; mtDNA, mitochondrial DNA; RAGE, receptor for advanced glycation end products; SIRT, sirtuin; TAPI-2, TNF protease inhibitor-2; TACE, TNF converting enzyme; TGFs, transforming growth factors; TLR2, toll-like receptor 2; TNF, tumor necrosis factor.

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### 1. Introduction – why link cerebral malaria and Alzheimer's disease?

Several years ago, driven by the conflict between the traditional mechanism in the literature on human cerebral malaria and the logic of physiology (and therefore pathophysiology), we were searching the outer reaches of its literature for a way to describe the cul de sac the field had entered. This uncovered what we regarded as a particularly clear article (Castellani et al., 2008) on the same problem in Alzheimer's disease: a well-entrenched conventional approach born of an ancient histological observation being argued to have outlived its usefulness – indeed, in Castellani's words, having become

somewhat of a nuisance to what should be the real focus, invisible molecules and subcellular damage that cannot be verified by direct observation.

This sounded familiar for some of us in the human cerebral malaria world. This form of encephalopathy rivals Alzheimer's disease in the scale of the problem, either condition being much more common than other similar diseases. Untreated malaria infection in naive individuals can best be described as fitting in with the systemic infectious diseases that are liable to exhibit an associated encephalopathy. Over a hundred years ago, and for want of any other leads, pathologists were attracted to red cells containing parasites often seen in sections from brains of fatal coma cases caused by the parasite *Plasmodium falciparum* (Marchiafava & Bignami, 1894) when seeking a mechanism for the neurological aspects of malaria, and mechanical blockage of micro-cerebral vasculature soon became the accepted explanation for this condition. Thus in the disease caused by this species of malaria, which dominates most of the malarial world, parasitized red cells adhering to the walls of small cerebral blood vessels became the primary cause of abnormal brain function. This is referred to as the parasite sequestration argument of malarial disease, in which physically restricted blood flow is believed to deprive brain cells of oxygen and nutrients. It is still commonly championed as the primary cause of the encephalopathy seen in human malaria, although, as discussed later, emerging information on vivax malaria is making this allegiance difficult to sustain.

As reviewed (Castellani et al., 2008), those searching brain sections from cases that included what is now called Alzheimer's disease described neurofibrillary tangles (NFTs) (Fuller, 1907) – later found to be comprised of tau protein – and plaques, later realized to be formed of amyloid beta ( $A\beta$ ). As with adherent parasitized red cells in malaria, these obvious physical structures became the centre of attention for investigators trying to understand Alzheimer's. The earlier literature on the apoE4 association with  $A\beta$  (Section 5.2.5.) undoubtedly reinforced the case for amyloid being the primary cause of this disease. Assumed to be directly harmful, NFTs, plaques, and the molecules that formed them became the target of science-based interventions. It seems that in both fields, therefore, histological hallmarks have somehow been allowed to evolve into primary mechanisms that researchers were comfortable with, perhaps, in part, because they could point them out with the aid of a microscope.

Over 60 years ago, a malaria researcher with sepsis research experience summarized the accumulated scepticism in the malarial literature, and suggested that inflammation was a more likely explanation for the mechanism of the disease (Maegraith, 1948). The parallels with the scientific challenge at the core of cerebral malaria and Alzheimer's disease are striking, not least because the key inflammatory cytokine, tumor necrosis factor (TNF), is appearing with increasing frequency in both literatures. The link between TNF and any non-tumor disease actually began with malaria.

In the late 1970's, when TNF had a small new literature restricted to tumor killing, one of us (IAC) began collaborating with the group who first described TNF (Carswell et al., 1975). This original TNF-systemic disease link thus made has been reviewed recently (Clark, 2007). Our joint research was directed at understanding host protection against malaria parasites as well as malarial pathology. By 1995, these same inflammatory mediators were argued to be relevant to the pathogenesis of Alzheimer's and other neurodegenerative diseases (McGeer & McGeer, 1995). Some years later, tissues from malaria brains were stained for amyloid  $\beta$  ( $A\beta$ ) precursor protein (APP) (Medana et al., 2002) and cerebral malaria CSF was assayed for tau (Medana et al., 2005). In the same year it was reported that excess  $A\beta$  would not alter an important neurological function in mice lacking the gene for TNFR1 (Wang et al., 2005), and later cerebral malaria severity was correlated with CSF levels of TNF (John et al., 2008b). In the following year it was reported that TNF induces tau accumulation (Gorlovoy et al., 2009). Clearly, acquiring a decent

working knowledge of the science behind  $A\beta$  and tau, the histological hallmarks of Alzheimer's disease, is necessary for research into malarial disease, and competence in Alzheimer's research increasingly requires reasonable literacy in TNF. Only then can the considerable congruency between cerebral malaria and Alzheimer's disease be appreciated. Indeed, such an interaction was the genesis of the collaboration represented by this review, with a TNF researcher with a cerebral malaria background reading a review on the link between neurogenesis and cytokines (Abdipranoto-Cowley et al., 2009).

The major theme developed in this review is that the development of the basic literature on TNF and neurophysiology (and hence neuropathophysiology) appears to apply to cerebral malaria and Alzheimer's disease. Although these two diseases differ greatly in the age group affected and rapidity of onset and duration of clinical change, Alzheimer's might be usefully viewed, in pathophysiological terms, as a much less acute, but more insidious, version of cerebral malaria and its sequelae. This contrast should not deter us from this view, any more than should the chronicity of AIDS dementia conceptually separate it from the more acute viral encephalopathies. All encephalopathies discussed here show evidence of the same pattern of events, albeit with different severities and durations. Cytokine-mediated diseases in particular are inevitably multifactorial, and much can be lost if each is considered in isolation. Our hope in this review is that by considering these diseases together perspectives that might have otherwise lain dormant will surface, and bring about a consensus that leads to useful treatment.

## 2. Tumor necrosis factor TNF and the other cytokines, and the innate immune system

As well as discussing the inflammatory cytokines themselves, a brief overview of the innate immune system, which is evolutionarily older than adaptive immunity, is warranted here. Activation of the innate immune system leads to the production of TNF and similar cytokines, and depends on them. The past decade has witnessed considerable conceptual advances in our understanding of how innate immune responses, which dominate the CNS (Section 4.5), are initiated. New inclusive terminologies have accompanied this. PAMPs, or pathogen-related molecular patterns (Janeway, 1992), on viruses, bacteria and fungi are now joined by DAMPs, (danger-associated molecular patterns) found on proteins, released from damaged cells (Matzinger, 2002) as entities recognized, or sensed, by pattern recognition receptors (PRRs). Gram-negative bacterial cell wall lipopolysaccharide (LPS), also called endotoxin, and discussed variously throughout this review, is an example of a PAMP, and HMGB1 (Section 4.3) can be regarded as a DAMP. The most intensely studied PRRs are the Toll-like receptors (TLRs) comprising a superfamily of at least 11 members expressed by a wide range of cells. As reviewed, (Beutler & Poltorak, 2001), the *Drosophila* Toll protein has a fascinating lineage stretching from insect dorso-lateral polarity, through insect innate immunity, to its homologs being important for mammalian innate immunity. Once activated, TLRs induce signalling cascades that predominantly lead to release of inflammatory cytokines, chemokines, and anti-microbial peptides (AMPs). This is discussed in more detail in Section 5.2.3).

More recently, other families of PRRs have been described. Unlike most TLRs, which are membrane-bound, the newer PRR families consist of soluble proteins that provide cytoplasmic surveillance in order to detect intracellular invaders. They include NLRs, or NOD-like receptors. NLRs, and other proteins such as NALP3, form cytoplasmic complexes termed inflammasomes that activate caspase-1, thus generating IL-1 $\beta$  and IL-18 from their precursor proteins (Martinon et al., 2002, 2009). IL-18 is a well-known inducer of TNF (Tsutsui et al., 1997; Dinarello, 1999). TNF in turn promotes activation of caspase-1 induced by other agents, such as ATP (Franchi et al., 2009), logically enhancing IL-1 $\beta$  and IL-18 production, and hence reinforcing itself, even in the absence of microbial infection.

Cytokines have many functions, and are generated by many cell types. Many are classified under the general term of interleukins (Aarden & others, 1979), which currently extend from interleukin-1 (IL-1) to IL-34. Some, but not all, earlier cytokines, such as the interferons (Isaacs & Lindenmann, 1957) the lymphotoxins (LTs) (Ruddle & Waksman, 1967), TNF (Carswell et al., 1975), and the transforming growth factors (TGFs) (Moses et al., 1981) did not conform to this terminology. Cytokines have primary roles in physiology, including neurophysiology, innate and acquired immune responses and wound healing, as well as disease pathology. They operate in networks and cascades, and regulate cellular activity in an autocrine, paracrine and hormonal manner. For example IL-1 is induced by TNF, and shares many of its activities, including the capacity to induce IL-6. This cytokine is often used as a marker for systemic inflammatory reactions because it appears in the circulation conveniently later, when illness is undeniable, and stays high for longer. As another example of the complexities, both TNF and IL-1 induce IL-8, the prototype chemotactic cytokine, or chemokine, terms given to cytokines that attract cells along their concentration gradients. Many cytokines are now arranged in families with similar structures and functions. Some cytokines, such as TNF, LTs and IL-1 are pro-inflammatory, whereas others such as IL-4, IL-10 and the TGF family, are anti-inflammatory, being subsequently induced to inhibit these changes. These anti-inflammatory cytokines are established, from the work of Schwartz's group (see Section 6.2.2.), to be central in understanding how macrophages of systemic origin control brain injury and degeneration.

A number of more newly discovered interleukins, such as IL-12, IL-17 and IL-22 are increasingly linked with TNF (Yamaguchi et al., 2007). This group of interleukins need to have their roles in innate immunity and inflammation explored further, but since they are as yet little studied in the brain they largely remain outside the scope of this review. In passing, we note that the original term, TNF (Carswell et al., 1975), is identical to TNF- $\alpha$ , the commonly-seen suffix being a now meaningless relic from when LT, now LT $\alpha$ , was, for a limited period some years ago, referred to as TNF- $\beta$ .

Some cytokines, the prime example being TNF, are highly pleiotropic. It is, for example, an *in vitro* tumor killer, a common regulator of normal physiology, including of synapses, a homeostatic agent in cell proliferation (see Section 3.3), a key mediator of innate immunity (Clark et al., 1981), and the master cytokine that precipitates the inflammatory response (Charles et al., 1999). Our proposal that the pathology of malaria and salmonella infections is caused by over-exuberant production of the mediators of innate immunity (Clark et al., 1981), coupled with its recent linkage to the role of TNF in neurogenesis following neurodegeneration (Abdipranoto-Cowley et al., 2009) led this review to be largely about this cytokine. Not only does TNF possess the multiple roles outlined above, but the hugely successful biotechnology industry based on antagonizing it (some \$20 billion in sales in 2010) provides excellent experimental tools to explore diseases further, as well as to treat patients. Also TNF has by far the largest literature of the cytokines, allowing more comprehensive sets of useful implications. Several negative feedback loops of cytokines and their products have been shown to limit TNF generation. An example is hemoxygenase-1 (HO-1), which generates carbon monoxide (CO), which in small amounts is anti-inflammatory (Lee et al., 2003), because it inhibits TNF. This, and a functionally similar molecule, hydrogen sulfide (H<sub>2</sub>S), are discussed in more detail in Section 3.4.

### 3. Roles of tumor necrosis factor in the brain

Is TNF an initiator, or merely a marker, of brain disease? In practical terms, does intrathecal inflammation, of which TNF is the prototype mediator, precede the onset of Alzheimer's, or is it a reaction to its presence? To approach this question, TNF levels were assayed in CSF from 56 individuals who had mild cognitive

impairment (MCI), a condition that can develop into frank Alzheimer's disease, and 25 age-matched controls. When levels were tracked over a period, MCI cases were much more likely to develop into Alzheimer's if they had higher TNF levels (Tarkowski et al., 2003a). Newer studies continue to reaffirm this principle, with markers of inflammation showing in serum and CSF before any indications of increased A $\beta$  or tau (Laurin et al., 2009; Schuitemaker et al., 2009). This is consistent with data in Sections 5.2.1 and 5.2.2. Another group took advantage of the increased sensitivity of assaying for soluble TNF receptors rather than TNF itself. They found good evidence for levels of these receptors in serum and CSF predicting, over a 4–6 year period, conversion to clinical Alzheimer's disease (Buchhave et al., 2009).

Others have studied plasma levels of acute phase proteins, which are widely used as general indicators of inflammation that are upregulated by TNF or IL-1. For example C-reactive protein (CRP) and  $\alpha$ 1-antichymotrypsin (ACT) (Engelhart et al., 2004), or CRP alone (Laurin et al., 2009; Schuitemaker et al., 2009), are present in serum and CSF before any indications of increased A $\beta$  or tau. These studies have now been extended and confirmed by an impressive new report of plasma levels of another acute phase protein, clusterin (apolipoprotein J), being intimately associated with onset, progression, and severity of this disease (Thambisetty et al., 2010). A novel and impressive proteomic neuroimaging paradigm was employed. Unfortunately, the authors refer only to the amyloid chaperone function of clusterin, and seem to have been unaware of its role as an acute phase protein (Hardardottir et al., 1994), and therefore a marker, as surely as are CRP and ACT, of inflammation. One of their more telling findings was that clusterin was raised 10 years earlier than fibrillar A $\beta$  deposition. Taken together, these acute phase reactant arguments are an impressive link between inflammation the primary instigation of Alzheimer's disease.

Basic knowledge of the roles of cytokines such as TNF in the brain has expanded greatly in the last decade. For instance, as discussed in Section 5.2, TNF is associated at several levels in the amyloid pathway: before APP, in the conversion of APP to A $\beta$ , and after A $\beta$  has been generated. For some time TNF has been realised to be generated not just by microglia, as their macrophage-like character would predict, but also by neurons and astrocytes. In the light of its original tumor-killing capacity, brain-origin TNF was initially thought of purely in terms of causing cell death, but nowadays it is widely appreciated to be a physiological gliotransmitter concerned with normal communication, particularly between astrocytes and synapses, microglia and synapses, and thus synapse regulation. Indeed, TNF fits nicely into the tripartite view of synaptic physiology (Perea et al., 2009). It can also suppress adult neurogenesis, allowing net neuron numbers to reduce because of non-replacement (Abdipranoto-Cowley et al., 2009). TNF also influences cerebral function through acting on mitochondria. This section summarizes these functional roles of TNF.

#### 3.1. The effect of TNF on synapse function

The synapse is a particularly dynamic part of the neuron, and one of the earliest lesions noted in Alzheimer's brains is a reduction in synaptic efficacy (Selkoe, 2002). As recently reviewed (Arendt, 2009), many studies have also reported that synaptic loss is very pronounced. Indeed it is thought to be the strongest structural correlate of cognitive decline in this disease (DeKosky & Scheff, 1990). Soluble A $\beta$  oligomers have been shown to be a contributing factor (Shankar et al., 2007). Another such factor is nitric oxide (NO) from inducible nitric oxide (iNOS, or NOS2) stimulated in response to TNF that is induced by LPS (Weberpals et al., 2009), in the reasonably presumable absence of A $\beta$ . As discussed in Section 4.5, these authors were investigating the protection provided by the long-term cognitive defects caused by LPS, and found that NOS2 gene deficiency prevented loss of the structural synaptic proteins seen in normal mice. These two

observations are quite compatible when the ability of soluble A $\beta$  oligomers to induce TNF, and its inactivity if it does not do so (Meda et al., 1995; Wang et al., 2005; Rowan et al., 2007) (see below), is taken into account. Additionally, these data are consistent with A $\beta$  (Abramov et al., 2009) and TNF (Domercq et al., 2006) having similar effects on synaptic vesicle release.

The review referred to above (Selkoe, 2002) was written when it was beginning to be realized that the naturally occurring soluble human oligomeric forms (rather than monomers or visible fibrils or plaques) of beta-amyloid were best at inhibiting long-term potentiation (LTP) (Walsh et al., 2002) and *in vivo* cognitive behaviour (Cleary et al., 2005). Since A $\beta$  induces TNF (see Section 5.2.4) it is not surprising that both of these agents give the same outcomes in many functional assays, such as those done on brain slices. Paired functions of A $\beta$  and TNF extend, for instance, to LTP, with both soluble A $\beta$  dimers (Taylor et al., 2008; Ondrejcek et al., 2009) and TNF (Tancredi et al., 1992; Wang et al., 2005) shutting it down. In addition, two synthetic agents being developed for treating Alzheimer's, Ro25-6981 and ifenprodil, were recently reported to prevent the shut-down of synaptic transmission caused by soluble A $\beta$  dimers in rats (Hu et al., 2009). Two different specific anti-TNF agents, infliximab and a specific peptide antagonist were equally inhibitory as these GluN2B-specific drugs. In addition, they reported that Ro25-6981 prevents inhibition of synaptic transmission caused by TNF. So would, by definition, specific anti-TNF agents that enter the brain in sufficient quantities. These authors also added to the evidence that A $\beta$  dimers act through TNF by demonstrating that synapses in apical dendrites are susceptible to both agents, and synapses in basal dendrites are susceptible to neither.

In 2005 the capacity of TNF to rapidly upregulate calcium-permeable AMPA/kainate channels on neurons, with obvious implications for it to influence synaptic transmission, was published (Ogoshi et al., 2005). Soon after, the phenomenon known as synaptic scaling, in which the strength of all synapses on a cell are adjusted in response to prolonged changes in the cell's electrical activity, was argued to be mediated by TNF (Stellwagen & Malenka, 2006). In addition, TNF has been shown to control regulation of the type-1 inositol 1,4,5 trisphosphate receptor (IP<sub>3</sub>R), which is central to calcium homeostasis, and calcium-dependent functions, of neurons (Park et al., 2008). Subsequently, TNF was reported to modulate synaptic plasticity through inducing sphingomyelinase-2, and thus controlling the insertion of N-methyl-D-aspartate (NMDA) receptors into neuron membranes (Wheeler et al., 2009). Additionally, TNF can be expected to alter synaptic function through its striking ability to regulate their morphology. In brief, the percentage of pedunculated spines on dendrites is considerably decreased in neuronal cultures from iRNA-mediated TRAP-1 (TNF receptor-associated protein-1) knock-down mice (Kubota et al., 2009).

It has recently been more appreciated that the astrocyte, as the predominant glial cell of the central nervous system, orchestrates the whole enterprise through the processes of one astrocyte contacting tens of thousands of synapses, and controlling discrete large territories (Kuchibhotla et al., 2009). While astrocytes do not generate action potentials, they nevertheless have the capacity to communicate well with neurons (Haydon, 2001). TNF (which astrocytes as well as microglia and neurons can release) normally stimulates the release of glutamate from astrocytes – through selective activation of purinergic P2Y1 receptors on astrocyte cellular membranes (Domercq et al., 2006) – into the synapse microenvironment (see for review (Volterra & Meldolesi, 2005; Haydon & Carmignoto, 2006)).

While the research referred to directly above was performed with the goal of establishing the range of physiological roles of TNF in synapse function, it also lays the groundwork for understanding the pathological consequences of TNF excess. For instance, if the active, soluble forms of A $\beta$  act through TNF, which has been shown to alter synaptic transmission, it becomes plausible that the same synaptic

pathology (caused by TNF) explains the similarity between dementia of Alzheimer's disease and the delirium and encephalopathy in critical illness, independent of their initiating factor (Stevens & Pronovost, 2006).

It therefore seems reasonable to presume that the neurological consequences of excess brain concentrations of TNF (usually defined as cerebrospinal fluid (CSF) concentrations exceeding the upper limit of the homeostatic range required for normal physiological function) inhibit neurotransmission. Examples of this limit being exceeded include the increased brain and CSF levels of TNF reported in human (Moller et al., 2005) and experimental (Barichello et al., 2009) meningitis, cerebral malaria (John et al., 2008b), Alzheimer's disease (Tarkowski et al., 2003a, 2003b), fronto-temporal dementia (Sjogren et al., 2004) and Parkinson's disease (Mogi et al., 1994). Where examined, CSF and serum levels of TNF did not correlate, implying separate generation within each compartment. These concepts are examined further in the section on post-sepsis encephalopathy (see Section 4).

A dramatic example of altered functionality is the synchronous hyperactivity and intercellular calcium waves recorded in astrocytes in a mouse model for Alzheimer's disease (Kuchibhotla et al., 2009). These waves did not originate near A $\beta$  plaques or neurons, but within the astrocytes themselves. Another example is the reduced capacity of hippocampal astrocytes to release glutamate when exposed to TNF in another mouse model of Alzheimer's (Rossi et al., 2005). The authors point out the potential for this defect to down-regulate synaptic activity in these mice. Similar experiments are yet to be performed in meningitis and cerebral malaria models.

### 3.2. Influence of TNF on the role of zinc in synapse function

For decades zinc has been realised to be necessary for normal brain function (Wallwork, 1987). Much of the brain zinc is bound in metalloproteins, as elsewhere in the body, and the remainder stored in pre-synaptic vesicles. Ionic zinc has long been argued to have a physiological role in hippocampal synaptic neurotransmission (Xie & Smart, 1991).

TNF is very involved with zinc metabolism. Zinc-deficient animals are more sensitive to TNF (Waelpu et al., 2001), and hypozincemia is so dramatically induced by LPS, the prototype inducer of TNF, that blood zinc level has been proposed to be a useful assay for pyrogens (Boobis & Hartley, 1981). TNF induces astrocytes to secrete S100B, a zinc-binding protein (Edwards & Robinson, 2006), and TNF (Tarkowski et al., 2003a) as well as S100B (Green et al., 1997; Peskind et al., 2001), are increased in the CSF, and, perhaps as a consequence, distributed in brain regions in proportion to degree of pathology (Van Eldik & Griffin, 1994) in Alzheimer's patients. Ionic zinc is an endogenous modulator of synaptic functions controlled by alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA), NMDA and gamma-aminobutyric acid (GABA) receptors (Harrison & Gibbons, 1994; Smart et al., 1994). These synaptic functions include LTP, a synaptic phenomenon widely theorized to be a correlate of memory formation (Bliss & Collingridge, 1993), which is abnormal in zinc deficient rats (Hesse, 1979). Since S100B is a zinc-binding protein (Mbele et al., 2002), it is plausible that its induction (Edwards & Robinson, 2006) contributes to inhibition of synaptic activity by TNF. This is consistent with reports of impaired spatial learning memory after chelation of synaptic zinc (Frederickson et al., 1990) and learning impairment after dietary zinc deprivation (Takeda et al., 2000). It is also consistent with the recent report that Alzheimer's-like pathology is exacerbated in Tg2576 mice overexpressing human S100B (Mori et al., 2010).

Interactions between S100B and metallothioneines, the more widely appreciated endogenous zinc chelators, appear to have not yet been studied. The review of the roles of zinc in brain function and pathology that has recently appeared (Bitanirwe & Cunningham,

2009) is excellent, but did not include the implications of S100B in this context.

### 3.3. TNF and adult neurogenesis

A decade ago, when much scepticism still surrounded the evidence for neurogenesis in adults, the shrunken brains of Alzheimer's patients was simply interpreted as cell loss through neurotoxicity. This view will no doubt be revised by the discovery of stem cells in the adult human, primate and rodent hippocampus (Zhao et al., 2008). As recently reviewed (Abdipranoto et al., 2008), such brain shrinkage can readily be reasoned to indicate an imbalance between cell loss and neurogenesis, or even the failure of an intrinsic capacity for replacement of loss through natural attrition, much the same as in other organs. Functional studies of neurogenesis in the adult mouse hippocampus support the concept that such replacement is an important regulator of long-term memory (Rola et al., 2004; Aimone et al., 2006; Zhao et al., 2008; Kriegstein & Alvarez Buylla, 2009). In particular, arguments exist that continuous neurogenesis is required for normal retention of spatial and contextual memory (Snyder et al., 2005; Imayoshi et al., 2008) and object recognition.

Inflammation can repress neurogenesis (Abdipranoto-Cowley et al., 2009). When the requirement for A $\beta$  to induce TNF before it can influence learning and memory is considered (Wang et al., 2005; Ralay Ranaivo et al., 2006; He et al., 2007), research on A $\beta$  inhibiting neurogenesis (Haughey et al., 2002) is entirely consistent with the TNF data summarized in this section. Likewise, a series of papers on allopregnanolone and neurogenesis in Alzheimer's disease appears to have left the effects of TNF on neurogenesis out of consideration. In brief, allopregnanolone has been shown to be reduced in brains of Alzheimer's patients (Marx et al., 2006), and to be a potent proliferative agent for both rodent and human neural stem cells (Brinton & Wang, 2006). Neither of these papers, nor a more recent review (Wang et al., 2008), refer to the capacity of allopregnanolone, at levels detected in vivo, to reduce LPS-induced TNF release from macrophages (Ghezzi et al., 2000), and to reduce TNF and IL-1 $\beta$  production in cells isolated from rats with traumatic brain injury (He et al., 2004).

To our knowledge only one paper, from a group working on post-stroke neurogenesis, has examined the functional interaction of A $\beta$  and TNF (Katakowski et al., 2007). As the authors point out, stroke is a known cause of dementia, and ischemic events are thought to contribute to the progression of Alzheimer's disease. A $\beta$  deposits are associated with ischemic injury (Koistinaho & Koistinaho, 2005) as well as with Alzheimer's disease. These authors found that intraventricular infusion of TNF protease inhibitor-2 (TAPI-2), which increases TNF levels, decreased proliferation of subventricular zone cells subjected to middle cerebral artery occlusion. These results are consistent with the idea that TNF reduces the number of subventricular zone progenitors. In addition, primary cultures of the ischemic subventricular zone cells, collected before TAPI-2 infusion, over-expressed tumor necrosis factor converting enzyme (TACE) and its substrates, amyloid precursor protein (APP) and TNF. These concepts, and their therapeutic implications, have been argued to warrant examination in an Alzheimer's context (Katakowski et al., 2007).

Neurogenesis is part of the bigger picture in which pathways directly or indirectly triggered by TNF influence cellular homeostasis throughout the body, with low levels enhancing proliferation of a cell or organelle that higher levels destroy (Bernardino et al., 2008). This phenomenon extends to thymocytes (Ranges et al., 1988; Hernandez Caselles & Stutman, 1993), hepatocytes (Diehl & Rai, 1996; Bour et al., 1996), hematopoiesis (Clark & Chaudhri, 1988; Rebel et al., 1999) and mitochondria (Suliman et al., 2005; Nisoli & Carruba, 2006; Haden et al., 2007). Hence in some experimental circumstances TNF can inhibit neurogenesis (Monje et al., 2003; Cacci et al., 2008), and so too can inhibiting TNF with an anti-TNF antibody (Heldmann et al., 2005).

We propose that, in keeping with other organs, above, the effect depends on how much TNF is present. These principles extend to acute infectious disease, with a TNF-inducible enzyme, inducible nitric oxide synthase (NOS2), being implicated (Hoffmann et al., 2007) in meningitis-induced neurogenesis. TNF is also implicated in the neurogenesis of stroke (Heldmann et al., 2005) and, if A $\beta$ -induced TNF (Meda et al., 1995; Wang et al., 2005) is accepted and taken into consideration, Alzheimer's disease (Becker et al., 2007).

### 3.4. TNF, brain mitochondria, and the brain's high ATP requirements

A $\beta$  is associated with mitochondrial dysfunction, in which oxygen usage by the respiratory chain complex is reduced, leading to lowered ATP generation and functional loss in brain cells (Aleari et al., 2005; Chen et al., 2006). While much of this literature implies that A $\beta$  can impair mitochondrial function directly, newer research takes into account the observation that A $\beta$  is a ligand for RAGE, the receptor originally described for its ability to tether advanced glycation endproducts (AGE) to cell surfaces. Once occupied, RAGE activates NF- $\kappa$ B, a transcription factor that initiates the inflammatory cascades induced by TNF and related cytokines. RAGE-bearing cells include neurons and microglia (Yan et al., 1996). TNF can apparently inhibit mitochondrial function directly (Busquets et al., 2003); however, and more plausibly, does so by generating mediators, such as NO and oxygen radicals, with this capacity. This concept has expanded greatly in the basic (Sanchez-Alcazar et al., 2000) and applied literature, particularly that of sepsis and heart failure (Moe et al., 2004). Much research exists on the pathways involved in this bioenergetic aspect of systemic inflammation (Fink, 2001, 2004; Singer et al., 2004), as well as heart (Mariappan et al., 2009) and brain (Tangpong et al., 2008) pathology. Hence a reasonable explanation for the mitochondrial dysfunction caused by A $\beta$  involves TNF-initiated reactive oxygen species.

This explanation assumes that mitochondrial dysfunction in Alzheimer's disease requires A $\beta$  accumulation. However, there is now evidence, in a mouse model bearing two mutations in APP, that lowered mitochondrial membrane potential and reduced ATP levels appear three months before A $\beta$  deposits (Hauptmann et al., 2009). Attributing this ATP reduction to intracellular A $\beta$  (Hauptmann et al., 2009) fails to consider the capacity of A $\beta$  to induce TNF (Meda et al., 1995), and of TNF to inhibit mitochondrial function (see below). Moreover, evidence of inflammation in the brain has been reported earlier than are A $\beta$  or tau (Schuitemaker et al., 2009).

In 1954 Brian Maegraith (who in 1948 was the first to view malaria as an inflammatory state, as reviewed in this journal (Clark & Cowden, 2003)), published electron micrographs of damaged mitochondria from tissues of malaria-infected monkeys. In this article it was discussed, for the first time, the possibility of functional hypoxia in malaria arising from interference with oxygen acceptance by host cells, rather than the traditional concept of insufficient oxygen reaching the mitochondria (Maegraith, 1954). Later his group wrote of serum factors producing changes consistent with this (Riley & Maegraith, 1962). In 1972 altered mitochondrial structure was also reported in experimental sepsis, and linked to functional hypoxia in the presence of adequate oxygen (Mela et al., 1972).

The next advance leading to understanding how TNF influences mitochondrial function came from observations in John Hibbs' laboratory in the form of the argument that activated macrophages kill tumor cells by inhibiting their mitochondrial respiration, and thus ATP generation (Granger et al., 1980). This response was subsequently shown to require L-arginine (Hibbs et al., 1987), although at the time this had no special significance. Once it was realized that NO was derived from this amino acid, however (Moncada et al., 1989), the essential role of NO in tumor killing by macrophages was rapidly appreciated (Hibbs et al., 1988). By 1990 it was appreciated that TNF induced NO generation (Kilbourn et al., 1990). A few years later Fink

embarked on a detailed study of the biochemistry behind mitochondrial functional loss in sepsis. He proposed the concept (which he termed cytopathic hypoxia) that the effects of primary hypoxia are mimicked in a range of severe infectious diseases by the mitochondrial respiratory chain becoming dysfunctional (Fink, 1997, 2000, 2001). TNF became involved in the description of cytopathic hypoxia when his group found that incubating enterocytes with TNF, IL-1 and interferon- $\gamma$  (IFN- $\gamma$ ) reduced their oxygen consumption, and thus ATP output, via NO-mediated reduction of oxidative phosphorylation (Khan et al., 2002). Meanwhile, the phenomenon, and general reasoning, was extended to HIV dementia (Kruman et al., 1999) and influenza encephalopathy (Yokota, 2003).

With the revival of the notion of microbial origin of organelles (Margulis & Chapman, 1998), the above mechanism is consistent with evolutionary perspectives on mitochondria, which have been proposed to arise from a common origin with Rickettsiae (Emelyanov, 2001). In parallel with what is observed with mitochondria (see above paragraph) Rickettsiae, like many other intracellular organisms are also susceptible to TNF-induced NO (Turco & Winkler, 1993). This observation is consistent with evidence that much higher TNF expression occurs in mammalian cells on exposure to microbial and free mitochondrial DNA (mtDNA) than when they are exposed to mammalian nuclear DNA, which lacks a high density of unmethylated CpG sequences common to microbial and mtDNA (Zhang et al., 2009). As discussed in Section 4.3, this also rationalizes, through release of mtDNA, high TNF induction in brain trauma, as well as the encephalopathy subsequent to systemic trauma.

As well as the high energy requirement the brain has for general cellular function, it must generate additional ATP because its release by both neurons and glial cells is important in intercellular communication (Gordon et al., 2005). In addition, a novel form of synaptic plasticity that is feed-forward, expressed quickly, mediated by the synaptic activation of neighboring astrocytes, dependent on release of ATP, and acting on post-synaptic purinergic receptors, has been recently described (Gordon et al., 2009). If this ATP release were regulated by TNF actions on mitochondria in astrocytes, then it is reasonable to propose that this plasticity would in fact be indirectly regulated by TNF. While the proposal that TNF could regulate ATP release during synaptic plasticity remains to be proven, it is consistent with the data described above, and with the growing literature that suggests TNF regulates synaptic plasticity (Turrigiano, 2008). If TNF indeed regulates synaptic plasticity through an effect on mitochondrial ATP production, it would provide an additional reason why normal synaptic function would be at risk in diseases in which brain TNF levels are increased. This argument can be further developed in diseases such as Alzheimer's, which is currently generating an extensive literature on mitochondrial dysfunction (Ferrer, 2009; Carvalho et al., 2009). As discussed in Section 5.2.2., inhibited energy production, which can be caused by TNF, also increases the rate of APP cleavage, with harmful effects (Gabuzda et al., 1994; Velliquette et al., 2005).

#### 3.4.1. Carbon monoxide, TNF, and mitochondria

CO is usually discussed in medicine because of its harmful ability to out-compete with oxygen for binding to hemoglobin, and cytochrome oxidase within mitochondria (Goldbaum et al., 1976). It is now appreciated that cytochrome c oxidase I within complex IV, a mitochondrial enzyme responsible for the reduction of oxygen into water as the final step of the electron transport chain, is involved. For some time it has been realized that CO is generated physiologically by the action of hemoxygenase on heme, and has essential anti-inflammatory properties. The early observation that TNF induces hemoxygenase-1 (HO-1) (Cantoni et al., 1991) was later explained by the ability of TNF to induce IL-10, which in turn induced HO-1 (Platzer et al., 1995), thus generating CO (Lee et al., 2003). Likewise, the anti-inflammatory effect of 15 $\beta$  prostaglandin J<sub>2</sub>, shown to be present in tissues during inflammation (Gilroy et al., 1999), also

operates through induction of HO-1 and subsequent generation of CO (Leeper-Woodford et al., 1991).

The anti-inflammatory action of HO-1-induced CO has been demonstrated to operate through down-regulating protein release of several pro-inflammatory cytokines, including TNF, and similarly up-regulating the anti-inflammatory cytokine, IL-10 (Otterbein et al., 2000). These activities have been adopted experimentally to reverse a wide range of inflammatory states, including skeletal muscle contractile failure (Taille et al., 2001), experimental autoimmune encephalomyelitis (Liu et al., 2001), endotoxin-induced endothelial cell activation (Bulger et al., 2003), uveitis (Ohta et al., 2003), focal ischemia-reperfusion cardiac injury (Giannini et al., 2005), rejection of transplanted tissue (Camara & Soares, 2005), and a mouse model of cerebral encephalopathy of malarial origin (Pamplona et al., 2007a). Notably, this model, discussed further in the same year (Pamplona et al., 2007a), employed (separately) both pharmacological HO-1 induction and CO inhalation therapy.

It has been known for some time in Alzheimer's disease that HO-1 is overexpressed in neurons and astrocytes in the hippocampus and cerebral cortex relative to control brains (Schipper et al., 1995). Concern has been expressed by these authors that the excess free iron this produces could cause oxidative damage (Schipper, 2004). Accordingly, a therapeutic approach to inhibit HO-1 activation (Song et al., 2009) has been proposed. However, it is widely accepted that endogenously increased HO-1 activity is an attempt to restore homeostasis during inflammation (Otterbein et al., 2000). Indeed, the immediately above papers are a compelling part of the argument for a strong inflammatory component in Alzheimer's disease. Intriguingly, in the mouse model of malaria encephalopathy, in which HO-1 was overexpressed (Pamplona et al., 2007b) either by pharmacologically causing additional expression or by enforcing inhalation of 250 ppm of CO for 24 h produced an excellent recovery. It seems logical that, in a chronic condition, inhaled CO might render homeostatic HO-1 activation unnecessary by providing the anti-inflammatory component, CO, without excess free iron potentially setting a harmful Haber–Weiss reaction in motion.

As outlined above, the high affinity of CO for cytochrome c oxidase has, until recently, been thought to inform us only about its toxicity. In 2007, however, an additional mechanism for the anti-inflammatory action of CO has arisen from evidence that the action of lower levels of CO on cytochrome c oxidase is actually healthy for mitochondria, increasing their membrane potential through enhancing generation of reactive oxygen radicals that mediate adaptive mechanisms that counter cellular dysfunction (Zuckerbraun et al., 2007). In the same year this concept was given a practical application when CO inhalation by mice was shown to reverse the mitochondrial damage and apoptosis in myocardiocytes by doxorubicin, a frequently employed anti-tumor drug. In this CO inhalation model mitochondrial biogenesis was promoted, and apoptosis much reduced, (Suliman et al., 2007). The basic biology of this approach has been reviewed (Bilban et al., 2008).

More recently these activities of CO have been applied, using the ruthenium-based water-soluble CO carrier, tricarbonylchloro(glycinato) ruthenium (II) (CORM-3), in an in vivo murine sepsis model (Lancel et al., 2009). This agent releases CO upon injection. The authors present evidence for mitochondrial energetic metabolism being restored, and mitochondrial biogenesis being activated in septic animals. It is as yet unresolved why the CO carrier drug CORM-3 reduced TNF levels in an in vitro model (Sawle et al., 2005) but not in vivo (Lancel et al., 2009), whereas CO itself reduced it both in vitro (Otterbein et al., 2000; Zuckerbraun et al., 2007) and in vivo (Otterbein et al., 2000; Pamplona et al., 2007b). If upheld, the in vivo CORM-3 data (Lancel et al., 2009) imply a parallel route of CO acting directly on mitochondria rather than through inhibiting TNF production.

To date CO inhalation or CO-releasing drugs have not been used to treat any encephalopathy except the mouse model discussed above

(Pamplona et al., 2007b). Reservations have been expressed, regarding human malaria, in terms of what is known about human HO-1 distribution and the level of CO-bound hemoglobin in infected and non-infected individuals (Hunt & Stocker, 2007). Nevertheless there is widespread interest in using CO-releasing drugs to therapeutically managing inflammatory states in general (Foresti et al., 2008). Currently the use of CO-releasing drugs has not yet been tested on human subjects beyond volunteers breathing CO (1 h/day at 100 ppm for 5 days). This treatment regimen in skeletal muscle causes changes consistent with increased mitochondrial biogenesis (Rhodes et al., 2009). It could be relevant that galantamine, a drug in common useage because it has some activity in suppressing Alzheimer's diseases – presumably for its capacity to reversibly inhibit acetylcholine esterase – has recently been shown to induce HO-1 (Nakao et al., 2008).

#### 3.4.2. Hydrogen sulfide, TNF, and mitochondria

A similar story is unfolding for H<sub>2</sub>S. Although generated by different enzymes from unrelated substrates, CO and H<sub>2</sub>S are remarkably similar in their functions, from acute rotenone-type toxicity (i.e. directed at the mitochondrial oxidative respiration) when in excess, to anti-inflammatory activity in low doses. Like CO, H<sub>2</sub>S is part of normal physiology, including in the brain (Boehning & Snyder, 2003; Ishigami et al., 2009). Again like CO, it has mitochondrial cytochrome *c* oxidase as a target for normal physiology, as well as pathophysiology (Collman et al., 2009). This group describe the differential effects of high and low levels of H<sub>2</sub>S on mitochondrial cytochrome *c* oxidase. These studies are reminiscent of those on lower levels of CO on this enzyme actually improving mitochondrial energetic metabolism (Zuckerbraun et al., 2007). This aspect has apparently not yet been addressed for H<sub>2</sub>S.

Like CO, H<sub>2</sub>S protects (Li et al., 2005) against the LPS inflammatory response models often referred to in this review. Both gases have the capacity to inhibit LPS-induced TNF and its attendant cascade of pro-inflammatory mediators, as well as to upregulate protein of the anti-inflammatory mediator, IL-10 (Otterbein et al., 2000; Li et al., 2009; Whiteman et al., 2010). Using purpose-designed H<sub>2</sub>S-releasing agents, this cytokine-inhibiting activity has been extended to microglia and astrocytes (Lee et al., 2010). This group have confirmed that astrocytes in normal human brain contain H<sub>2</sub>S (Lee et al., 2009). Another H<sub>2</sub>S, sodium hydrosulfide (NaHS), is being investigated as a protective agent in a Parkinson's disease model (Hu et al., 2010). The observation, in their study, of H<sub>2</sub>S preventing the development of lesions induced by rotenone implies it possesses mitochondrial-improving activity, as found for CO (Lancel et al., 2009). Predictably, the pattern of investigation and possibilities for H<sub>2</sub>S, including using H<sub>2</sub>S-releasing agents, will parallel that of CO and the pharmaceutical agents that release it. New data, in which i.p. injection of an H<sub>2</sub>S donor protected rats from cognitive defects, cerebral NFκB inhibition and reduced TNF generation resulting from intracerebral LPS infusion (Gong et al., 2010), are consistent with this.

Manipulating CO and H<sub>2</sub>S in order to treat human inflammatory brain disease is logical, and may prove to be less expensive, as well as easier to administer, than specific anti-TNF agents. But they require much more developmental study before they are clinically useful. In contrast, anti-TNF agents, which go to the heart of the problem, and have an impressive history in treating chronic human inflammatory states for 16 years (see Section 6.2), are potentially applicable now. These two approaches to influencing essentially the same pathway may provide synergistic treatments for the range of brain diseases discussed in this review.

#### 3.5. TNF, sirtuins, and caloric restriction

The concept of dietary (caloric) restriction extending the healthy life spans of simple life forms, as well as mammals, is well established. Inevitably, therefore, its effects on neurophysiology and mental

capacity, a key aspect of the ageing process, have been examined (Mattson et al., 2003). Examples of outcomes of caloric restriction include enhanced learning in aged mice (Ingram et al., 1987) and rats (Stewart et al., 1989), protection against age-induced deterioration of memory in rats (Pitsikas & Algeri, 1992), and enhanced hippocampal neurogenesis in mice (Lee et al., 2002). It is of particular experimental interest that caloric restriction was not required when animals were young, with mice in which it was not instigated until mid-life retaining normal function well into old age (Means et al., 1993). Age-associated increases in serum levels of TNF are considered physiological, and much studied in the ageing literature. It was reported in 1997, when neither caloric restriction nor TNF was yet of interest to the Alzheimer's research community, that caloric restriction prevented mouse serum TNF from rising with age (Spaulding et al., 1997). A decade later it was shown in a diabetes model that caloric restriction inhibits serum TNF and other inflammatory cytokines, as well as upregulating the anti-inflammatory cytokine, IL-10 (Ugochukwu & Figgers, 2007).

A mechanism for the link between caloric restriction and suppression of circulating TNF levels has been constructed through the argument that SIRT1, a class III histone deacetylase that requires NAD<sup>+</sup> as a cofactor, causes adaptations to caloric restriction (Rodgers et al., 2005). These include inhibiting NFκB activity (Yeung et al., 2004; Salminen et al., 2008), and thus inhibiting TNF production (Shen et al., 2009). SIRT1 is the most studied member of the sirtuins (SIRT1-7) a group that share many activities. Resveratrol, a polyphenol belonging to the phytoalexin family, is found in seeds of various plant species, including grapes, and has been proposed to be an agonist of SIRT1 (Yeung et al., 2004), explaining why it inhibits TNF production (Bi et al., 2005; Shen et al., 2009). Sirtuins (Albani et al., in press) and resveratrol (Anekonda, 2006) have both been argued to have potential as treatments in Alzheimer's models, but whether through TNF modulation was not addressed by these authors.

Resveratrol is freely available and has been a self-administered agent for some time, so its literature is confused. Of relevance to Alzheimer's are the studies reporting protection in rat brain trauma (Ates et al., 2007; Sonmez et al., 2007) when administered i.p., and its reported capacity to reduce deficits in working memory as a dietary supplement (Abraham & Johnson, 2009). At the time of writing, the effects of resveratrol on TNF levels does not appear to have been contemplated in any of these studies. Nevertheless small molecule SIRT1 agonists are being actively sought (Smith et al., 2009a; Bemis et al., 2006), and another group has reported screening such agents for their anti-inflammatory effects, using reduced TNF levels as a readout (Nayagam et al., 2006). An opposing argument is developing that resveratrol functions through AMP-activated kinase, with SIRT1 activation data arising from an artefact (Tang, 2009; Beher et al., 2009). These groups do not deny resveratrol or caloric restriction their published activities, but argue that other, largely undefined, activities rather than SIRT1 activation would be needed to be harnessed in order to find other resveratrol-like agents.

The effect of both caloric restriction and TNF (TNF the more extensively, and always independently) have been studied on basic brain function, as well as in Alzheimer's models. For instance, the literature on the effects of caloric restriction on synaptic plasticity was reviewed in detail in 2008, with no mention of TNF (Fontan-Lozano et al., 2008), yet by then TNF, which caloric restriction reduces (Spaulding et al., 1997), already had a considerable synapse literature in its own right, as reviewed in that year (Turrigiano, 2008).

#### 4. Encephalopathy accompanying systemic infection, trauma or burns

Historically, encephalopathies have been viewed as the presence of diffuse multifocal cerebral functional change that is not, at least in the beginning, combined with morphological correlates (Kunze, 2002).

Typically, they are described by the nature of the systemic inflammatory state that induces them, septic encephalopathy being associated with severe bacterial sepsis. Severe viral diseases, hepatic failure, pancreatitis, trauma with anesthetized (e.g. surgery) or unanesthetized injury, or severe burns can also exhibit encephalopathy. These states can be clinically characterized by slowing of mental processes, impaired attention and cognition, disorientation, delirium or coma.

In passing, acute infections exhibiting delirium have been so common historically that they provide a fascinating anecdotal link from the Ancients to day-to-day life in modern Italy. Probably for millennia, lice have spread a rickettsial disease that, since the 18th century, has been termed typhus, to denote a fever accompanied by neuropsychiatric manifestations. The disease later shown to be caused by an organism that became known as *Salmonella typhi* was sufficiently typhus-like to be termed typhoid. The term typho-malaria, now outmoded, was, until relatively recently, an accepted diagnosis for a marked association between malaria parasites in blood smears and delirium. And today? Italians refer to deliriously enthusiastic supporters of a sport, or a commercial brand, as tifosi.

#### 4.1. Malaria

We have argued (Clark et al., 2008; Clark & Alleva, 2009) that malaria is best viewed as one of the cytokine-caused systemic infectious diseases that can be accompanied by encephalopathy. Nevertheless, we note that many malaria researchers still regard malaria pathophysiology as a special case distinct from the newer cytokine-based ideas used to explain the pathogenesis of systemic infectious disease in general. Instead, these researchers prefer the traditional argument that the primary cause of falciparum malaria is obstructed cerebral microvascular flow (Beare et al., 2009). Part of their argument has been that vivax malaria, a disease difficult to distinguish by traditional diagnostic methods, is rarely fatal, or a cause of neurological signs, because it does not obstruct vascular flow. See Section 5.1 for more detail. It transpires, however, that vivax malaria may have often been more harmful, with more encephalopathy in untreated cases, or cases in regions of chloroquine resistance, than previously realized. This may have been the result of confusion with falciparum malaria when using microscopy to identify the parasite, combined with a traditional conviction that if it was fatal it must have been falciparum. Together, the systematic diagnostic precision now available through PCR in much of India (Kochar et al., 2009) and publications arising from close examination of a large number of field cases in New Guinea (autonomously by two geographically separated groups (Genton et al., 2008; Tjitra et al., 2008)), have consolidated the view that vivax malaria encephalopathy is much more common than previously thought. In summary, each of these two research groups concluded that the frequencies of neurological disease, and the fatality rate, seen in infections with *P. vivax* were essentially the same as seen in the encephalopathy of falciparum malaria, as discussed recently (Clark & Alleva, 2009). Circulating TNF can be very high in vivax malaria (Karunaweera et al., 1992), although levels of TNF in the CSF, and whether they correlate with signs of encephalopathy, have never been published. Brain levels of TNF, assayed in CSF, and some downstream cytokines have been examined in falciparum malaria, and found to correlate with encephalopathy, including long-term functional loss (John et al., 2008b). The later, parasite sequestration-based events that complicate interpretation of the effects of late falciparum malaria on brain sections, as discussed a few years ago (Clark et al., 2006), are summarised in Section 5.1.

#### 4.2. Systemic inflammatory events and long-term cognitive defects

As well as the acute encephalopathy accompanying acute infection, as discussed above, a less severe but much more chronic version that follows systemic inflammation is important to understand. In 2005 a case-control study (with almost 10,000 individuals in

each group) showed an association between infectious disease and dementia in the elderly (Dunn et al., 2005). Long-term cognitive impairment, the nature of which is only now starting to be understood, can ensue (Stevens & Pronovost, 2006; Hopkins & Jackson, 2006). As discussed earlier, malaria is also noted for long-term cognitive impairment, including deficits in memory, attention, visuo-spatial skills, language and executive function. Prospective studies have documented these changes well (Boivin et al., 2007; John et al., 2008a). The condition has also been examined in children in other regions of tropical Africa (Carter et al., 2005; John et al., 2008b; Kihara et al., 2009), so older age is clearly not essential to the phenomenon.

In a review of apparent interactivity of systemic and cerebral inflammation, it was proposed (Perry et al., 2007) that, in diseases such as Alzheimer's, with its chronic activation of the innate immune response in the brain, successive systemic inflammatory events might be expected to exacerbate the rate of neurological degeneration. This idea is consistent with a report of chronic systemic inflammation, as measured by serum levels of IL-6, a stable cytokine induced by both TNF and IL-1 (Zhang et al., 1988), being inversely related to hippocampal grey matter size in middle-aged human volunteers (Marsland et al., 2008). Perry's group have since published mouse (Cunningham et al., 2009) and human (Holmes et al., 2009) data that fulfil this prediction (Perry et al., 2007). In brief, this human study (Holmes et al., 2009) found that increased baseline serum levels of TNF in Alzheimer's patients were of themselves associated with an increased rate of cognitive impairment. They also found that the more acute systemic inflammatory episodes associated with an increase in serum TNF the patient experienced over a six month period the more rapid was the rate of cognitive decline. The most extreme examples, those with the most frequent high TNF episodes, had a 10-fold increase in decline rate compared to those with low starting levels and no systemic inflammatory episodes.

Apart from its importance in allowing an appreciation of the relevance of TNF to move from the laboratory to the clinic, this work by Holmes and co-workers (above) has a practical consequence for testing anti-Alzheimer's agents. It rationalizes the short-term unpredictability of the rate of decline in this disease, which leads to the often-made remark that properly controlled trials are particularly essential in this disease because of its short-term clinical variability. Their work takes the argument further through implying that systemic inflammatory events should be documented and taken into account in such trials.

It does not necessarily follow, however, that because TNF of systemic origin can exacerbate Alzheimer's disease the cognitive decline and other changes in this condition originates from TNF, or TNF-secreting cells, of non-cerebral origin, and therefore can be countered by therapeutic anti-TNF agents that cannot effectively enter the CSF. In contrast, it is a reasonable assumption that the depression, and anxiety of rheumatoid arthritis and inflammatory bowel disease, both treatable with large and systemically administered specific anti-TNF agents (Uguz et al., 2009; Bercik et al., in press), arise from TNF or TNF-secreting cells of non-cerebral origin that has been neutralized before it reaches the brain.

The encephalopathies accompanying sepsis, trauma (Michaelson et al., 1984), burns (Mohnot et al., 1982) and elective surgery (Tufo et al., 1970) have been studied in parallel (Marchiori et al., 2006). IL-1 is also incriminated, as in a recent study of hepatic encephalopathy, using mice deficient in receptors for either IL-1 or TNF (Bemeur et al., 2010). This is not surprising, in view of the ability of TNF to induce IL-1, and the many functions they share (see Section 2). TNF has been identified as a key mediator in pancreatic (Yang et al., 2004) and septic encephalopathy (Alexander et al., 2008). TNF is increased in serum or plasma in all of the above predisposing conditions (Kwiatkowski et al., 1990; Molloy et al., 1993; Bhatia et al., 2000); (Van Reeth, 2000; Hildebrand et al., 2005), and is documented in the brain as well in

influenza (Ichiyama et al., 2003), malaria (John et al., 2008b) and liver failure (Jiang et al., 2009) encephalopathy. Thus the conditions that predispose to encephalopathy that can mimic Alzheimer's disease, but unlike it, dissipate over time, have been reported to generate a systemic inflammatory response syndrome, in which serum levels of TNF, and the cytokines it induces downstream, are increased.

The same concepts can be predicted to apply in circumstances when the temporary trigger for TNF generation is in the brain, rather than systemic, as above. The stimulus might be an infectious agent, such as in meningitis (Moller et al., 1991, 2005; Barichello et al., 2009), sindbis viral encephalitis (Carmen et al., 2009), influenza encephalopathy (Ichiyama et al., 2003), neuroborreliosis (Ramesh et al., 2008), and trypanosomiasis (sleeping sickness) (Kristensson et al., in press). Neurosyphilis (Halperin, 2010) also comes to mind, but remains unexamined. Understanding the encephalopathy of trypanosomiasis along these lines is well advanced. Brain TNF and IL-1 $\beta$  production is high (Quan et al., 1999), and various downstream mediators are increased in the choroid plexus (a site therefore worth examining in these human diseases) and elsewhere. Moreover, i.c.v. infusion of antagonists of receptors for IL-1 and TNF have been found to attenuate neurodegeneration (Quan et al., 2003). In addition a recent open trial reports a positive outcome in 6 out of 7 patients with neurosarcoidosis after infliximab treatment. The seventh responded to etanercept (Moravan & Segal, 2009). See Section 6.2.2. to have these drugs explained. From this reasoning, AIDS dementia (Mastroianni et al., 1990; Yeung et al., 1995) mimics Alzheimer's more closely because the source of TNF and IL-1 $\beta$  is largely unstopplable.

#### 4.3. Brain trauma

An obvious form of encephalopathy without an infectious or systemic origin occurs after brain trauma, a term we use to include closed head injury. Depending on severity and whether the injury is local or global, this condition can exhibit the full gamut of neurological dysfunction, including coma, impaired memory and executive function. In these patients CSF levels of TNF and other cytokines are increased (Hayakata et al., 2004; Goodman et al., 1990), and their manner of interacting has been compared with the same processes in Alzheimer's disease (Mattson et al., 1997). For example, synaptic plasticity is reduced in closed head injury patients (Albensi & Janigro, 2003). Two plausible triggers for TNF generation, HMGB1 (Scaffidi et al., 2002) and free mtDNA, exist in brain trauma. Like bacterial DNA, but unlike mammalian DNA, mtDNA is high in unmethylated CpG repeats, and therefore induces TNF release (Zhang et al., 2009). In systemic trauma the level of mtDNA in plasma (predictably therefore in CSF in brain trauma) correlated with the degree of clinical severity in a group of 38 patients (Lam et al., 2004). These concepts have recently been examined in more detail (Zhang et al., 2010). It would be surprising if this relationship does not also exist between CSF levels of mtDNA and the degree of brain trauma, in which intracerebral TNF inhibition has been reported to be protective (Shohami et al., 1996). Other evidence for a primary role of TNF in the pathogenesis of brain trauma comes from reports of IL-10, an anti-inflammatory cytokine (Knoblach & Faden, 1998), HU-211, a post-transcriptional inhibitor of TNF production (Shohami et al., 1997), and resveratrol (Ates et al., 2007; Sonmez et al., 2007), which also inhibits TNF production (Bi et al., 2005), being protective. It is therefore very relevant to the theme of this review that brain trauma is now generally accepted as a predisposing factor to early onset Alzheimer's disease (Mortimer et al., 1991).

It must also be appreciated that functional neurological changes can occur after trauma at sites other than the brain. The trauma associated with open heart surgery is an example (Tufo et al., 1970; Grimm et al., 2003; Newman, 2007). This indicates that cytokines, not

physical damage within the brain, are more likely to be the key to understanding pathogenesis.

#### 4.4. Experimental studies on the encephalopathies driven by systemic TNF

As described above, systemic inflammatory states, of infectious origin or otherwise, can develop an accompanying encephalopathy. It has been realised for over a decade that experimental non-cerebral inflammation influences the hypothalamo-pituitary-adrenal axis via TNF signalling within the brain (Turnbull et al., 1997). Awareness that i.p. injection of LPS, the prototype inducer of inflammatory cytokines, generates their transcripts within the brain also arose at this time (Pitossi et al., 1997). In 2006 a mouse model of liver failure subsequent to cholestasis was used to demonstrate that TNF-secreting monocytes migrate from the periphery to the brain, and somehow microglia are also triggered to release this same mediator soon afterwards (Kerfoot et al., 2006). A recent experimental example is colonic inflammation inducing cells in the rat hippocampus to generate TNF (Riazi et al., 2008). This increase in TNF level was functionally significant, since infusion of TNF antibody into the cerebral ventricles prevented the seizures associated with gut inflammation, but not the degree of gut inflammation itself. Contemporary studies have shown that migrating monocytes are attracted across to the brain by a chemokine (CCL2) originating from the microglia (D'Mello et al., 2009), and work demonstrating brain pericyte detachment from the basal lamina after i.p. LPS (Nishioku et al., 2009) suggests a likely route. As recently reviewed (Cameron & Landreth, 2010), a literature also exists on monocyte migration to the brain in Alzheimer's models, under the influence of the same chemokine. Though sometimes interpreted in terms of A $\beta$  plaque removal (not useful in a recent human trial (Holmes et al., 2008)), these cells may also seed the brain for excess TNF production (Kerfoot et al., 2006).

#### 4.5. Short-term systemic response causing a long-term cognitive defect

In 2004 it was reported that experimental myocardial infarction, which causes a systemic inflammatory milieu, also causes sustained (still increasing after 4 weeks) generation of TNF in the hypothalamus (Francis et al., 2004). Three years later others (Qin et al., 2007) found that mouse brain TNF production remained high for at least ten months after a single systemic LPS injection, whereas serum levels peaked at 9 h. These studies are consistent with earlier work in which the TNF switch-off that occurs systemically after a second LPS injection (the basis of LPS tolerance) proved to be absent in the CSF (Steinshamm & Waage, 2000). Thus TNF production evidently persists in brain for much longer than in the periphery, conceivably because the cytokines such as IL-10, which control it in the periphery, are less available within the CNS. Increasing this availability is the basis of a therapeutic approach that appears to have much potential (Koronyo-Hamaoui et al., 2009).

Collectively, these above data provide a glimpse into the profound implications, for understanding these cognitive defect diseases, of A $\beta$ <sub>42</sub> being an anti-microbial protein on the innate immune response, which dominates within the brain (see Section 5.2.3). One study (Qin et al., 2007) also found that microglia were chronically activated in the hippocampus and cortex after this single LPS injection. An intriguing difference between systemic and cerebral TNF generation is that TNF receptors (allowing extant TNF to function) are needed for brain, but not for liver, to generate TNF. Thus, systemic cytokines are critical for CNS effects in response to systemic immune activation. One observation this explains is experimental sepsis specifically and significantly reducing the activity of cytochrome *c* oxidase in brain mitochondria (d'Avila et al., 2008). This has considerable implication for meeting the brain's high requirement for ATP.

A useful step in elucidating long-term cognitive deficits triggered by severe systemic inflammatory responses (or worsened by a series of milder such events (Holmes et al., 2009)) was recently published (Weberpals et al., 2009). Using the same animal model, dose and route of LPS as above (Qin et al., 2007), these authors found, two months after treatment, that wild-type mice, but not others deficient in *NOS2* gene, committed significantly more working and reference memory errors. These mice are unable to make NOS2, the form of nitric oxide-generating enzyme normally induced by TNF and IL-1. In contrast, systemic LPS induced sustained levels of NOS2 in brains of wild-type mice, particularly in the hippocampus, for the full two months. Overt hippocampal neuronal loss or astrocyte activation could not be documented in either *NOS2* knock-out or wild-type mice, whereas microglial activation was present in wild-type, but not the knock-outs. Moreover, as evidence that NOS2-induced nitric oxide (NO) affects long-term cognition by influencing synaptic function, in these studies *NOS2* knock-out mice proved to be protected from reduction in presynaptic and postsynaptic proteins. One conserved protein of particular note was PSD-95, known to be critical for spatial memory formation (Migaud et al., 1998) in normal mice.

Taken together, these data (Weberpals et al., 2009) indicate that the absence of NOS2 induction protects mice from long-term microglial activation. In other words, NOS2-derived NO might contribute to the associated inflammatory reaction and long-term learning and memory dysfunction caused by systemic LPS administration. The authors suggest several possible mechanisms by which this NOS2-derived NO could contribute to these cognitive defects. These include reducing ATP levels by inhibiting mitochondrial respiration, altering synaptic structure, and reducing brain TNF and IL-1 $\beta$  levels (their mRNA levels were lower in post-LPS *NOS2* knock-out mice rather than controls, by an as-yet unexplained mechanism). All are plausible, and could act in concert. Clearly, reduced TNF levels are consistent with the concepts behind recent Alzheimer's studies (Tobinick, 2009; Holmes et al., 2009). It is relevant that human *NOS2* immunohistochemistry studies we performed some years ago (Clark et al., 2003) on fatal comatose malaria and sepsis cases showed identical *NOS2* staining of microglia (Fig. 1). Further studies in this field could examine whether the *NOS2* polymorphisms shown in such African populations (Hobbs et al., 2002; Burgner et al., 2003) bear any relationship to a tendency to develop long-term neurological deficit after recovery from this systemic inflammatory disease (Brewster et al., 1990; Carter et al., 2005; Kihara et al., 2006; John et al., 2008b; Kihara et al., 2009).

These new data (Weberpals et al., 2009) will undoubtedly encourage the use of *NOS2*-deficient mice and specific *NOS2* inhibitors in studying these encephalopathies. Moreover, it is difficult not to draw the conclusion, from the combined human (Holmes et al., 2009) and various animal studies, that patients with Alzheimer's and other encephalopathies above would be better off with lower serum and CSF levels of TNF, not an unattainable therapeutic outcome given the tools now available in the clinic. Different routes of administration would likely have different consequences, with subcutaneous etanercept (an anti-TNF biological) (Bohac et al., 2002) having the prospect of preventing these encephalopathies from becoming worse, and a route directed towards the CSF (Tobinick et al., 2006) possessing the prospect of improving the condition. Lower brain *NOS2* levels would be one consequence.

In summary, the above observations are consistent with any severe systemic inflammatory activation, through generating circulating TNF, and whether protozoal, bacterial, viral, traumatic (anesthetised or not) in origin, having the potential to cause long-term as well as acute loss of cognitive function. Moreover, these encephalopathies are unlikely to be caused directly by toxins unique to one of the disease variants, such as LPS or similar in sepsis, glycosylphosphatidylinositol (GPI) in malaria (Zhu et al., 2009), gp120 in AIDS (Tyring et al., 1991), but by the TNF that all of these agents induce.

Thus any TNF inducer serves the purpose, its harmfulness depending on how effectively, and for how long, it induces TNF. It follows that LPS, the prototype TNF inducer, is as relevant a model for any of these circumstances as it is to gram-negative sepsis, where it is present in the clinical disease.

#### 4.5.1. Sundowning

It would be instructive to know whether the phenomenon of "sundowning", the late afternoon and early evening exacerbation of behavioral symptoms in mild and moderate Alzheimer's disease that has been recognized by clinicians for more than 60 years (Bachman & Rabins, 2006), has been noted of the other encephalopathies discussed in this review. It appears to have no published rationale at present. We note, therefore, that a diurnal rhythm of TNF has been documented in the rat brain, with samples taken from hippocampus, hypothalamus, cerebral cortex, cerebellum, pons and midbrain at six hour intervals (Floyd & Krueger, 1997). Tissue TNF levels at light onset were about 10-fold higher than minimal night-time levels, particularly in the hippocampus. When it is taken into account that the rat is nocturnal, it would be surprising if this 10-fold increase would not produce a regular temporary exacerbation in all TNF-induced encephalopathies. In addition, it may account for the observation that long-term potentiation, a marker for synaptic activity, is diurnal in the normal hamster (Raghavan et al., 1999). This could readily be tested.

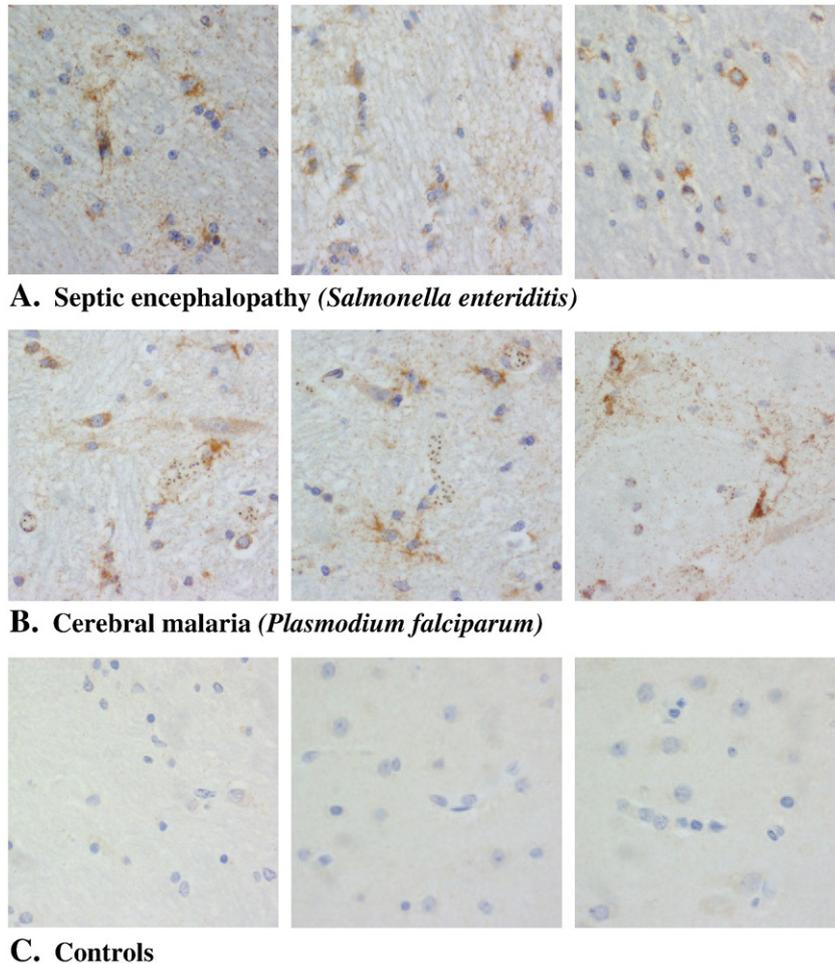
It also seems warranted to take note of what sundowning tells us about the relative importance of the possibilities for the main cause of the functional loss seen in Alzheimer's disease. It is difficult to interpret this diurnal variation in terms other than a fluctuations of a reversible change. The length of Section 3.1 on the effects of TNF on synapse function reflects both the importance others have put on synapses as the seat of this disease (DeKosky & Scheff, 1990; Selkoe, 2002; Arendt, 2009) and the size of the TNF literature on the topic. Equally, it is difficult to reconcile A $\beta$ -based primary neuronal death theories of Alzheimer's with the presence of diurnal clinical variation.

In summary, we visualize the sundowning effect seen in milder disease as consistent with a view that Alzheimer's symptoms are exquisitely sensitive to the acute effects of TNF on synapses, effects that would be rapidly reversible and therefore variable, in depending on diurnal TNF levels in the CNS. The chronic effect of relentless excess TNF, however, can be expected to be neuronal and axonal loss through TNF-driven APP occupying DR6 receptors (Section 5.2.1) and TNF reduced neurogenesis (Section 3.3). Over time these influences would result in the considerable loss of brain volume observed in terminal cases.

## 5. Malaria encephalopathy, Alzheimer's disease, depression and aggression

### 5.1. Malaria encephalopathy and TNF

As outlined in Section 4.1, the primary cause of abnormal brain function in falciparum malaria has traditionally been based on restricted cerebral blood flow caused by parasitized red cells adhering to the walls of small cerebral blood vessels (sequestration), depriving brain cells of oxygen and nutrients. Over the years such sequestration often proved difficult to align with the presence of cerebral symptoms (Maegraith & Fletcher, 1972). Cerebral hypoperfusion could not be detected (Newton et al., 1996; Clavier et al., 1999), and, given the stroke literature, it remains difficult to explain how recovery from deep coma without consistent and profound residual deficit is consistent with mechanical blockage of the cerebral vasculature being the primary mechanism. These parasites shed material that induces the patient's cells to release TNF, and thus the cascade of inflammatory cytokines that many now regard as the primary cause of the disease. This was first proposed in 1981 as one half of a single novel



**Fig. 1.** Immunohistochemical staining detecting iNOS in microglia from three representative forebrain sections of (A) cerebral malaria (B) septic encephalopathy and (C) control (no terminal coma) brains. Sections are respectively from MP 25, MP12 and MP41 in Table 1 of Clark et al. 2003, as are techniques (DAB, hematoxylin), materials and technical controls. Other cases in each category conformed to this pattern. Unpublished data.

concept, the other half being that these same mediators are the key to understanding innate immunity (Clark et al., 1981). Both arguments continue to expand, and are updated regularly (Clark & Cowden, 2003; Clark et al., 2004, 2006). By 1997 TNF had been shown by immunohistochemistry to be expressed in neurons and astrocytes in a mouse model of malarial encephalopathy (Medana et al., 1997) which, although criticized as largely irrelevant to the human disease (White et al., 2010), does manifest cognitive defects (Desruisseaux et al., 2008).

Although encephalopathy in isolation is seen in no more than a fifth of African children who are acutely ill with malaria (Marsh & Snow, 1997), its presence commonly brands all patients with neurological change as having cerebral malaria, whatever other severe pathology co-exists. Since it is this patient group in which large-scale assaying for TNF in sera from severe falciparum cases have routinely been done, the term cerebral malaria therefore tends to be used synonymously with severe falciparum malaria with coma. Hence serum TNF levels correlated well with severe malaria, but not always with what is termed cerebral malaria, i.e. a systemic disease with an encephalopathy component (Grau et al., 1989; Kwiatkowski et al., 1990; Shaffer et al., 1991).

When the problem was examined more closely, it was realised that TNF levels in the CSF, not those in the serum, correlated with the encephalopathy, as well as the residual neurological deficit that often occurs (John et al., 2008b). Moreover, CSF and serum levels did not correlate, implying independent cerebral generation of TNF. Many papers are published from East Africa on reduced cognitive function associated with a history of malarial encephalopathy. Several

functions of TNF warrant consideration in this context, including its inhibitory effects on synapses (Section 3.1), as well as its capacity to cause periventricular white matter damage (Aden et al., 2010).

As reviewed (Clark et al., 2006), the concept of malaria being a primary cytokine excess disease implies that the sequestration of parasitized red cells, the traditional primary disease mechanism, is a secondary event that begins in the brain once circulating TNF levels are high. The diseases caused by *P. falciparum* and *P. vivax* each present a TNF-induced, sepsis-like, encephalopathy, with the opportunity for the neurological dysfunction in falciparum malaria to become exacerbated by vascular obstructive hypoxia once the disease is already severe. The expanded experience of vivax malaria, a disease with negligible parasite sequestration, of recent years (Genton et al., 2008; Tjitra et al., 2008; Kochar et al., 2009) indicates that patients are sick enough, without sequestration, for a fatal outcome. Hence it has been reasoned that cerebral sequestration, severe enough to obstruct microvascular flow and cause microhemorrhages, can terminally exacerbate, but not explain, the primary pathogenesis of neurological dysfunction in falciparum malaria. This secondary phenomenon, on top of an underlying encephalopathy common, in principle, to all of the conditions covered in this review, is of immediate practical concern to those responsible for management of patients seriously ill from this infection. Reasonable in its day as a primary mechanism, it survives as a major research discipline largely because of the application of impressive technology to understand the mechanics of adhesion of infected red cells (Maier et al., 2008). Until recently, when the roles of TNF in hypoxia became appreciated (Section 5.3),

and the 2008 vivax malaria papers appeared (Genton et al., 2008; Tjitra et al., 2008), the dominance of this thinking, and the tradition of attributing all malaria pathology to it, retarded developing the cytokine approach to this disease, oddly enough the one in which the cytokine concept of disease began (Clark et al., 1981).

As summarized in Section 5.2.1, recent publications on the role of APP in fetal brain in triggering axonal pruning (Nikolaev et al., 2009), as well as the capacity of inflammatory mediators to upregulate APP (Brugg et al., 1995; Buxbaum et al., 1998; Goldgaber et al., 1989; Schmidt et al., 2008; Sommer et al., 2009), make it worth considering whether the APP stained in falciparum malaria brains (Medana et al., 2002) reflects the presence of more of this protein, due to increased levels of brain TNF (John et al., 2008b). Thus it would be the cause of axonal damage, as well as an indicator of its existence. Likewise, if the cotton wool spots noted in retinas of patients with cerebral malaria (Lewallen et al., 1999) are indeed manifestations of disrupted axonal flow (McLeod et al., 1977; Medana et al., 2002) it would, from the above reasoning, make them secondary changes consequential to the TNF that raised APP levels. Being a DR6 ligand (Nikolaev et al., 2009), this would damage axons, causing local distensions seen as white spots. This implies that cotton wool spots, used as a diagnostic feature in cerebral malaria (Lewallen et al., 1996), are simply another indicator that brain TNF is increased. The same TNF-induced pathway would also explain the increased tau levels in Kenyan children with cerebral malaria (Medana et al., 2007).

#### 5.1.1.1. Coma, hibernation and tau

Finally, what can the cytokine approach offer to explain the mechanism of the coma that is seen, certainly by clinicians, as the centrepiece to understanding malaria encephalopathy? As summarized in Section 3.4.2, H<sub>2</sub>S is a normal part of physiology in brain, and other organs. Like CO, it can both subtly adjust mitochondria and inflammation, and cause gross harm, depending on its concentration. We therefore draw attention, as briefly before (Clark et al., 2008), to the possibility of H<sub>2</sub>S being the primary cause of the reversible coma that can occur in both infectious (malaria, bacteria, viruses) and non-infectious (brain trauma, advanced Alzheimer's disease) encephalopathies. In mice, it can produce a reversible state of hypometabolism and suspended animation that will protect from hypoxia (Blackstone et al., 2005; Blackstone & Roth, 2007). This implies that the coma of cerebral malaria is actually an attempt to protect the patients from the damaging cerebral changes already underway, which could lead to loss of neuronal function, and of neurons, laying the ground for residual cognitive defects (Idro et al., 2005). In addition, the hypometabolism induced by H<sub>2</sub>S is reminiscent of the processes of hibernation. It is therefore noteworthy that the phosphorylated tau protein that indicates axonal pruning or injury – and seen not just in normal fetal and neonatal brains (Lovestone & Reynolds, 1997) but also in brains of adults with malaria (Medana et al., 2002), Alzheimer's and other neurodegenerative states (Goedert, 2004) and brain trauma (Franz et al., 2003) – is also present in brains of hibernating animals (Arendt et al., 2003; Hartig et al., 2007). We also note that, as recently reviewed (Arendt, 2009), hibernation has a literature on altered synaptic connectivity (Popov & Bocharova, 1992; Magarinos et al., 2006) and impaired cognitive function (Millesi et al., 2001) that is analogous to that of Alzheimer's disease. Tau has a literature of its own on inhibiting brain function (Goedert, 2004), recent research referred to on the effects of rapamycin in Section 6.1.3 (Caccamo et al., 2010) being an example of concepts we have not expanded upon for space considerations.

## 5.2. Alzheimer's disease and TNF

Ideas on the mechanism of Alzheimer's disease appear to be undergoing a transition. A contributing factor may be that two controlled trials against A $\beta$  have not produced a positive outcome. One, a six year follow-up of 80 patients in an A $\beta$ <sub>42</sub> immunization trial

(Holmes et al., 2008), had no shorter time to severe dementia or improved survival, even though removal of amyloid plaque was virtually complete. Another, the first bapineuzumab passive immunisation trial against A $\beta$  (Salloway et al., 2009), had a non-significant result. These outcomes, plus a long time awareness that amyloid plaques can be present in normal aged brains – for instance, in a 2001 study (CFAS, 2001) 33% of the 109 non-demented people had equivalent densities of neocortical neuritic plaques to equal-aged affected individuals – have contributed to the sense that the field needs to reconsider its direction (Lee et al., 2007; Castellani et al., 2009).

Clearly, the newer concepts and literature summarized below in Sections 5.2.1 question whether A $\beta$  is as useful a therapeutic target as was widely assumed when the various trials directed at neutralizing its presence or actions were conceived. Nowadays a considerable basic literature from a number of autonomous groups argues for increased brain TNF (Tarkowski et al., 2003b) having a primary role in pathogenesis of this disease (Tweedie et al., 2007; McCoy & Tansey, 2008; Janelins et al., 2008; Gorlovy et al., 2009; McAlpine et al., 2009; McNaull et al., 2010; Medeiros et al., 2010; van Gool et al., 2010). These data, plus the recent important and provocative identification of A $\beta$ <sub>42</sub> as an arm of innate immunity, a TNF-associated event (see Section 5.2.3), indicate a need for controlled trials intended to inhibit cerebral levels of the relevant cytokines, as discussed in Section 6. The study by McAlpine and co-workers, which used novel engineered dominant negative TNF inhibitors, long term, in a mouse model of Alzheimer's disease (McAlpine et al., 2009), is particularly compelling. Further interest in the inflammatory cytokine approach to understanding and treating Alzheimer's arises from a recent study (Holmes et al., 2009) on 300 patients (see Section 4.2). These competing approaches, amyloid and the cytokines that mediate innate immunity and inflammation, can be understood only by realizing how closely their functions are intertwined. The rest of Section 5.2 summarizes this.

### 5.2.1. TNF and APP in fetal development and adult disease

Here we can give no more than a brief overview of this important area of research. Alzheimer's disease is intimately associated with the APP gene (Hardy, 1994), the product of which, APP, is metabolized to A $\beta$ . As has recently been reviewed (Thinakaran & Koo, 2008), more than 25 APP mutations have been identified as being associated with the hereditary form of familial Alzheimer's disease, and APP gene duplication alone, as seen in trisomy 21, causes a high incidence of early onset Alzheimer's. An ample literature (Meda et al., 1995; Wang et al., 2005; Ralay Ranaivo et al., 2006; He et al., 2007) leaves little room to doubt that the consequences of A $\beta$ , downstream from APP, inducing TNF are very real for Alzheimer's disease (see Section 5.2.3). Nevertheless, an in-depth consideration of how closely TNF (and doubtlessly IL-1) are intertwined with this amyloid pathway requires us to begin by considering, in some detail, its relationship with APP, the molecule from which the hallmark A $\beta$  plaques arise. A human gene promoter region study has indicated that APP is a risk factor for Alzheimer's disease (Guyant-Marechal et al., 2007), and increased APP message is present in Alzheimer's disease brains (Moir et al., 1998). Although not along the lines described below, APP dysfunction was argued ten years ago to be central to Alzheimer's disease (Neve et al., 2000).

APP is one of a family of highly conserved transmembrane proteins, with homologues in *Caenorhabditis elegans* and *Drosophila melanogaster*. APP is expressed very early in fetal development (Ott & Bullock, 2001) and becomes ubiquitous. Indeed, the chick embryo is a model for basic research into its roles, and processing (Carrodeguas et al., 2005), and it is expressed in human embryonic stem cells (Porayette et al., 2007). Much more is present in the normal 18–22 week human fetal cerebral cortex than in the same part of the normal adult brain (Selkoe et al., 1988). Likewise, it is abundant in the 18 day fetal and early post-natal mouse brain, but has largely disappeared at 30 days (Trapp & Hauer, 1994). It appears to increase

in a controlled fashion, peaking in the mouse brain at about two weeks after parturition (Loffler & Huber, 1992). Within the normal brain it is an essential component of synaptic membranes, and, as recently reviewed (Arendt, 2009) over-expression and knockout studies in mice, and work with the *Drosophila* APP homologue, APPL, demonstrate essential physiological roles, such as synaptogenesis, axonal arborization, axonal transport, and synaptic plasticity, for these proteins during development and maintenance of the normal mature nervous system.

Some background on TNF receptors is warranted here. The original two were termed TNFR1 and TNFR2, the former becoming known as the death receptor for its link to apoptosis. The death receptors have since blossomed into a superfamily, with a terminology that renames the original TNFR1 as TNFR superfamily 1A, or TNFRSF1A. One of this series, TNFRSF21, also referred to as death receptor 6 (DR6), has been an orphan receptor (Bossen et al., 2006) that is widely expressed on differentiating neurons, but until recently (Nikolaev et al., 2009) had no known function on these cells. Importantly, these authors have identified N-APP, a cleaved amino acid fragment of APP, as a ligand for TNFRSF1A (DR6). From their evidence, they propose that APP uses this pathway to undertake essential axonal pruning and neuronal removal in normal late fetal and early neonatal development. TNF and related cytokines have been shown to be present early in the developing nervous system (Yamasu et al., 1989; Gendron et al., 1991; Merrill, 1992), and proposed to have undefined roles in normal brain development. One study (Yamasu et al., 1989) suggested the term ontogenic inflammation to describe the concept. They found that spontaneous TNF peaked in mouse brain cells at seven days post-partum. Intriguingly, this is not long before the peak brain concentration of APP (Loffler & Huber, 1992). Induction of APP by inflammatory cytokines is a widespread phenomenon, being reported in endothelial cells (Goldgaber et al., 1989), skeletal muscle (Schmidt et al., 2008), and 3 T3 L1 adipocytes (Sommer et al., 2009), as well as brain (Brugg et al., 1995; Buxbaum et al., 1998). Only the adipocyte study employed TNF, the others using IL-1, which functionally overlaps with TNF, or TACE, an esterase that acts on the transmembrane form of TNF to release the extracellular domain as soluble TNF. In addition, upregulation of promoter activity of the *APP* gene by several cytokines, including TNF, has been documented (Ge & Lahiri, 2002). All of this entirely consistent with the capacity of LPS, the prototype TNF inducer, to accelerate the production of APP and acceleration of its pathway in two different APP transgenic mouse models (Qiao et al., 2001; Sheng et al., 2003).

These actions of inflammatory cytokines are reminiscent of the capacity of TNF and IL-1 to induce a related molecule, serum amyloid A (McAdam et al., 1982), the precursor of AA amyloid, which can accumulate as fibrils in parenchymatous organs during certain chronic human systemic inflammatory states (Marhaug & Dowton, 1994). A promising human open trial with anti-TNF therapy has been reported (Kuroda et al., 2009).

From the data summarized above it can be proposed that during healthy fetal and neonatal development physiologically normal increases in cerebral TNF (and other pro-inflammatory cytokines) increase APP, which acts through the TNFRSF21 (DR6) receptor to reduce the excessive density of neurons, axons, and therefore neuronal connections down to the levels required for a normal post-natal existence (Nikolaev et al., 2009). The high TNF and thence APP (which these authors clearly appreciate, but in isolation from its TNF association) of Alzheimer's disease could thus, through an aberration of normal physiology, cause the brain to mistakenly read the signals that saw it so well through its fetal and neonatal development. Thus, for as long as TNF and associated cytokines persist in the brain, they would, via the APP they induce, slowly but eventually, pare down synaptic function to fatal levels. In a sense, therefore, the onset and progression of Alzheimer's disease relives the normal and at the time necessary changes that occurred, decades

earlier, in the patient's late fetal and early neonatal brain in response to physiologically increased TNF. It will be very informative to know what switches TNF production on and off in the normal fetal and neonatal mouse brain (Yamasu et al., 1989). These principles also provide a lead into how excessive fetal brain TNF levels might cause the white matter damage that leads to cerebral palsy (Dammann & Leviton, 1997; Bell & Hallenbeck, 2002).

These arguments are reinforced by a recent report that mice overexpressing non-mutant human APP had functional changes consistent with Alzheimer's disease (Simon et al., 2009). These mice also lost a series of synapse-associated proteins, including AMPA and NMDA subunits, and phosphorylated tau, an indicator of axonal damage, was increased in the hippocampus. Remarkably, A $\beta_{42}$  was barely detectable. This questions the centrality of A $\beta_{42}$ , the historical focus of much research and major patient trials. In addition, the loss of synapse proteins and accumulation of tau in these mice are entirely consistent with the neuronal loss and axonal pruning reported to be caused by APP/DR6 activity (Nikolaev et al., 2009). Mice expressing a mutant human APP demonstrated an entirely different picture, with no signs of neurodegeneration and much A $\beta_{42}$ .

It seems clear that awareness that cerebral TNF upregulates cerebral APP, and the consequences of APP being a ligand for TNFRSF21 (DR6), will help us understand the pathogenesis of all of the conditions in which excess cerebral TNF can be linked with brain dysfunction. This includes (see Section 4.2) the encephalopathies that can accompany infectious diseases caused by bacterial, viral and protozoal organisms (including malaria), burns, and trauma, particularly brain trauma. Clearly, the degree and continuity of TNF and therefore APP upregulation would determine acuteness and reversibility of the pathology. The principles outlined in Sections 4.2, 4.4 and 4.5 seem, at least at present, to be the most likely contender to explain the high cerebral TNF in Alzheimer's disease, although further insight is required to rationalize its persistence and enhancement. Our proposal, a positive feedback of APP enhancement from A $\beta_{42}$ -induced TNF, is summarised in Section 5.2.3.

### 5.2.2. TNF is also a driver of APP cleavage to generate A $\beta_{42}$

Normal metabolism of APP generates A $\beta$ , the major focus of much of the present research into the pathogenesis of Alzheimer's disease, and recent evidence (Section 5.2.3.) indicates that physiological levels are not harmful, but a useful part of innate immunity. Logically, toxicity arising from its rate of production, rather than the total amount represented in plaques, is a plausible explanation for large aggregations, generated slowly, being common in healthy brains (Schmitt et al., 2000). Generation of A $\beta_{42}$ , a potentially harmful form, occurs through the proteolysis of APP by the sequential actions of  $\beta$ - and  $\gamma$ -secretases. In 2004 it was shown that IFN- $\gamma$ , IL-1 $\beta$ , and TNF specifically stimulate  $\gamma$ -secretase activity, concomitant with increased production of A $\beta$  (Liao et al., 2004). Since TNF inhibits mitochondrial oxidative respiration (Busquets et al., 2003), these data were consistent with inhibition of energy metabolism having earlier been shown to enhance APP cleavage (Gabuzda et al., 1994). Subsequently (Yamamoto et al., 2007) IFN- $\gamma$  and TNF were shown to enhance A $\beta$  production from APP-expressing astrocytes and cortical neurons, and the numbers of astrocytes expressing IFN- $\gamma$  was increased. In addition, TNF directly stimulated  $\beta$ -site APP-cleaving enzyme (BACE-1, or  $\beta$ -secretase) expression and thus enhanced  $\beta$ -site processing of APP in astrocytes. Again, this is consistent with the energy inhibition that increases the rate of APP cleavage operating through enhancing  $\beta$ -secretase levels (Velliquette et al., 2005). In addition, TNFR1 depletion reduced BACE-1 activity (He et al., 2007). Logically, therefore, anti-TNF agents should, among their other actions, be effective APP cleavage inhibitors. Results obtained in mice with long-term inhibition of TNF are functionally consistent with this (McAlpine et al., 2009), although these authors conclude that

their data are in part consistent with APP induction, as discussed in Section 5.2.1.

From these studies it is conceivable that one role for physiological levels of TNF in the brain is to maintain APP and A $\beta$  homeostasis, with excessive TNF generation, from whatever origin, upsetting this to the detriment of synaptic function because it generates A $\beta$  too rapidly. Once generated excessively, A $\beta_{42}$  induces more TNF (Section 5.2.3.), which, as described in various sections of this review, opens the way for much subtle pathology associated with Alzheimer's disease to be generated directly from this and functionally related cytokines.

### 5.2.3. A $\beta_{42}$ shown to be an anti-microbial mediator of innate immunity

It is warranted to pause here and consider what might have been the evolutionary advantage of conserving the capacity of TNF to induce APP and enhance its cleavage to form amyloid forms including A $\beta_{42}$ , a protein that, until recently, had a reputation only for harmful effects. In keeping with everything else induced by TNF, a normal physiological role for A $\beta_{42}$  would make these pathways much more likely to be functionally significant *in vivo*. Evidence consistent with this has recently been extensively documented in a report that A $\beta_{42}$  is a potent antimicrobial peptide (AMP), with activity equal or greater than the prototype AMP against seven common human pathogens (Soscia et al., 2010). AMPs are part of the innate immune response (Scott & Hancock, 2000), and a number of them, including defensins (Seo et al., 2001; Hao et al., 2001), as well as the new recruit, A $\beta_{42}$  (Section 5.2.2.), are induced by TNF, one of the pillars of innate immunity (Ortaldo et al., 1986; van der Meer, 1988). Therefore these new data (Soscia et al., 2010) greatly strengthen the logic and plausibility of Section 5.2 of this review as a whole. Credit should also be given to the Chilean group who two years ago had reasoned the logic of a hyperactive cerebral innate immunity being central to Alzheimer's disease (Fernandez et al., 2008). The strengthening of this notion by the above observations (Soscia et al., 2010) has particular interest for our group, in that we argued the case in a malarial context, decades ago, that overexpression of mediators of innate immunity causes pathology as well as host protection (Clark et al., 1981; Clark, 1987).

The reported antimicrobial activity of A $\beta_{42}$  implies, should it be readily reproduced by others, that the goal of much of the last decade's mainstream Alzheimer's research – to permanently remove, by vaccination, as much of this material as possible – could have had long-term unfortunate consequences. Regularly suppressing TNF could also (through inhibiting APP induction (Section 5.2.1.) and cleavage (Section 5.2.2.)) reduce A $\beta_{42}$  levels below what is needed for innate immunity to function normally, but dosage of anti-TNF can be adjusted should infection occur, whereas vaccination aspires to be permanent.

As noted above, A $\beta$  appears, from this detailed new publication, to have a unique role in innate immunity, for as well as being a mediator of its effects it also perpetuates the response. Indeed, the NALP3 inflammasome (Section 2) has recently been shown to be involved as a sensor of A $\beta$ , including in TNF production (Halle et al., 2008). The researchers who first made this connection, and the consequences of their data, are summarized in the next Section.

### 5.2.4. TNF is also generated by A $\beta_{42}$

About five years ago it was demonstrated that the ability of A $\beta$  to induce TNF, a cytokine shown earlier to alter transmission in hippocampal slices (Tancredi et al., 1992), was a likely reason why these soluble forms of A $\beta$  altered synaptic function (Wang et al., 2005; Rowan et al., 2007). By 2005, several laboratories began reporting influences of TNF on aspects of synaptic transmission (Pickering et al., 2005; Stellwagen et al., 2005). Several years later it was reported that the impaired memory induced in mice by A $\beta$  could be prevented by i.c.v. injection of an anti-TNF antibody (Alkam et al., 2008). The capacity of A $\beta$  to act as a ligand for CD14 and TLR2 (Fassbender et al., 2004; Jana et al., 2008; Tukul et al., 2009) and TLR4 (Reed-Geaghan et al., 2009) gives these findings (Cleary et al., 2005; Wang et al., 2005;

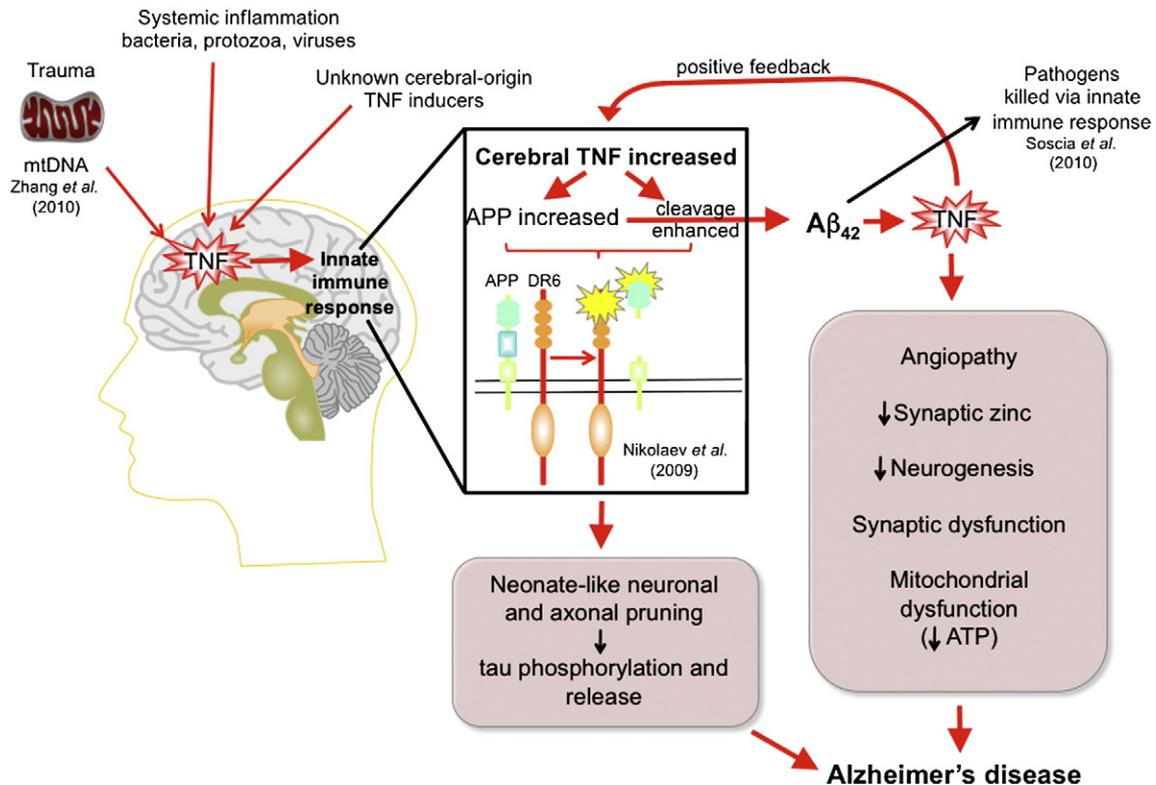
Rowan et al., 2007) authority from basic immunology, since for about a decade it has been accepted that occupying these receptors is how the standard bacterial and protozoal-origin inducers of TNF operate (Beutler et al., 2001). Unfortunately, some groups studying the role of soluble A $\beta$  on synaptic function still fail to incorporate this dependence of TNF in their study designs, even though it is predictably (Fassbender et al., 2004; Jana et al., 2008) present in the *in vitro* model they explore (Smith et al., 2009b). It will be instructive to learn whether PBT2, the metal-protein attenuating compound that targets pathology caused by A $\beta$  (Lannfelt et al., 2008) reduces the capacity of A $\beta$  to induce TNF.

Thus TNF appears to have three interlocking pathogenic roles in encephalopathies in which its brain levels are raised. The first upregulates APP levels to what they were during early neonatal life, fooling physiological reflexes to cause pathology, the second participates in driving APP to A $\beta_{42}$ , and the third, induced by A $\beta_{42}$ , apparently mediates many of its harmful effects. Clearly, all of these actions of TNF are attractive therapeutic targets, as discussed in Section 6. Logically, this A $\beta_{42}$ -induced TNF can be expected to add to the pool inducing APP (Section 5.2.1) as well as cleaving it (Section 5.2.2.). This would provide a positive feedback loop to combine with TNF from over-expression of innate immunity, providing a mechanism for the usual inevitable worsening of untreated Alzheimer's disease (Fig. 2).

### 5.2.5. What apolipoprotein E4 tells us about Alzheimer's

Apolipoprotein E protein (apoE) is expressed in humans as three common isoforms, E2, E3, and E4, and *apoE4* is a susceptibility gene with a highly significant linkage with late-onset and familial Alzheimer disease (Strittmatter et al., 1993). Some Alzheimer's researchers have regarded apolipoprotein E4 itself as the causative factor (Mahley et al., 2006), and a few saw its implications for understanding the pathogenesis of the disease in terms of inflammatory cytokines (Lynch et al., 2001; Ophir et al., 2005), but most rationalized the association in ways allied to traditional thinking about this disease. The reduced levels of total brain apoE seen in carriers of *apoE4* (explained in terms of astrocytes preferentially degrading apoE4 (Riddell et al., 2008)) is argued by these authors and others to imply that useful functional roles argued for apoE, including transporting the cholesterol needed for synaptic repair would not be operating effectively in carriers of this gene. The same is still said for apoE binding to A $\beta$  in order to promote its clearance and degradation (Riddell et al., 2008), although the recent revolution in the role of A $\beta$  (Section 5.2.3.) now questions whether clearing A $\beta$  is the worthy goal it was once thought. Nevertheless, it is largely through its association with A $\beta$  that the *apoE4* allele literature has become part of the argument in favor of the amyloid theory of Alzheimer's pathogenesis.

It is instructive to read of *apoE4* outside the Alzheimer's literature. Head trauma, a condition acknowledged to be mediated through cytokines such as TNF that have been most probably induced by triggers other than LPS (Section 4.3) has been part of the *apoE4* – Alzheimer's equation for some time. While A $\beta$  increases in this condition, it has never been taken to be the prime mover of it, as it has been in Alzheimer's. In 1995 it was reported that *apoE4* positive patients who had experienced head trauma were 10 times more likely to develop Alzheimer's disease, whereas *apoE4* positives with no head trauma history have double the risk (Mayeux et al., 1995). In their study head trauma alone carried no perceptible increased risk. Given the link between head trauma and TNF, this synergy is reminiscent of the report, in the same year, of a significant association of the rs1799724 SNP within the promoter of the TNF gene exhibiting synergy with *apoE4* in conferring risk of Alzheimer's disease (Laws et al., 2005). Experimentally, apoE (–/–) mice exhibit more severe neurological and cognitive deficits after closed head injury (Chen et al., 1997), and transgenic animals bearing human E3 fared better in function recovery than did those bearing E4 (Sabo et al., 2000). It has



**Fig. 2.** Interplay between TNF and the APP pathway it induces physiologically in late gestational and early neonatal brains, and also in adult brains in which TNF is pathologically increased, with Alzheimer's disease as an example of an outcome.

also been reported that apoE4 positivity causes excess dementia in HIV infection (Corder et al., 1998), which is not the A $\beta$  and tau disease that Alzheimer's is (Gisslen et al., 2009), instead being closely associated with the main inflammatory cytokines (Xing et al., 2009). Likewise, the apoE4 genotype doubles the duration of delirium in severely ill patients in intensive care (Ely et al., 2007), a field associated, for some time now, with TNF and IL-1.

In 1999, researchers attempting to explain their observation that apoE (–/–) mice were very readily killed by *Klebsiella pneumoniae* reported that these animals, on i.v. injection with LPS, produced 4–5 times higher plasma levels of TNF than did normal mice (de Bont et al., 1999). They rationalized this in terms of apoE, lacking in these mice, being a potent detoxifier of LPS. These principles have recently been investigated more closely, by another group, using transgenic mice expressing human apoE3 or apoE4 instead of the mouse equivalents. Those expressing human apoE4 proved to be much more susceptible to gut bacterial sepsis (Wang et al., 2009b). The study includes a protective effect, with associated reductions of TNF and functionally similar cytokines, of an apolipoprotein E mimetic that plausibly will prove useful to treat many of the diseases discussed in this review. This same group, previously focussed on apoE isoforms and the regulation of NO production by microglia (Brown et al., 2002), have recently reported on neuroinflammatory responses, including TNF levels, in cells from these same mouse strains (Vitek et al., 2009). In brief, cultured microglia from the hu E4 mice that had received LPS and IFN- $\gamma$  had an appearance classically associated with activated microglia, and produced much more NOS2 message, nitrite (a marker for NO) and more TNF, IL-6 and IL-12 than did microglia from hu E3 mice. Admixing with E2 demonstrated that the E4 effect was dose dependant for this allele, not total apoE.

These authors therefore argue the case for apoE4 being causally linked to enhanced neuroinflammation in a range of neurological diseases, including Alzheimer's disease. Given the additional evidence summarized in this review incriminating TNF, and the capacity of TNF

to induce APP (Section 5.2.1.) as well as push its cleavage to A $\beta$  (Section 5.2.2.), apoE4 is inevitably also a marker for A $\beta$ . But the direct linkage that rationalizes its association with Alzheimer's disease is evidently with TNF. It should be noted that the data from brain trauma, above, tell us that other TNF triggers, as well as LPS, can function in this disease.

### 5.3. Involvement of TNF in any contributions from ischemic hypoxia in 5.1 and 5.2

Some malaria (Maier et al., 2008) and Alzheimer's disease (Napoli & Palinski, 2005; Thal et al., 2009) researchers place emphasis on ischemic hypoxia making an important contribution to the disease they study. Indeed, in falciparum malaria the parasite sequestration that concentrates in the cerebral vasculature in severe cases (see Section 5.1) could well do this, through the ability of TNF to upregulate endothelial molecules (Wenisch et al., 1994), as well as, once sequestration has occurred, provide a local hot spot of further TNF (Ho & White, 1999) and NOS2 (Clark et al., 2003). production. As noted earlier, iNOS has important implications for long-term cognitive loss (Weberpals et al., 2009) after systemic infection.

In Alzheimer's disease a cerebral angiopathy characterized by A $\beta$  deposition within the walls of cerebral arteries and capillaries, is of major concern, and a target of experimental therapies based on reducing Ab deposition (Weller et al., 2009). Unfortunately, this publication makes no reference to the ability of A $\beta$  to induce TNF (Meda et al., 1995; Wang et al., 2005; Ralay Ranaivo et al., 2006; He et al., 2007), nor to the vasculature changes seen in the rat hippocampus after injecting A $\beta$  into this site being significantly reduced by administering thalidomide, an anti-TNF agent, i. p. for 7 days (Ryu & McLarnon, 2008). This is consistent with a recent human study in which the inflammatory arthropathy that tends to accompany rheumatoid arthritis was assessed (by measuring aortic pulse wave velocity) with and without the usual commercial TNF

antagonists with which such patients are treated. Vascular pathology was significantly reduced by this treatment (Angel et al., 2010).

As well as these arguments for the involvement of TNF in causing hypoxia in Alzheimer's disease and rheumatoid arthritis, this cytokine has been demonstrated to be central to primary hypoxia-induced brain pathology (Barone et al., 1997). Others have confirmed this experimentally by administering anti-TNF antibody i.c.v (Liesz et al., 2009) or etanercept i.p. (Aden et al., 2010). The latter study, in rats, is of particular interest. It shows how prior systemic inflammation predisposes neonatal brains to a subsequent, hypoxic/excitotoxic, insult, in a model for the sequence of events that leads to the brain damage known to cause early learning difficulties and cerebral palsy. If etanercept was administered after (but not before) the excitotoxic insult, when the blood-brain barrier was damaged, it greatly decreased brain TNF and reduced cerebral damage by 50%. As the authors note, this is consistent with the concept that systemically administered etanercept cannot normally reach the brain.

#### 5.4. Major depression and TNF

Nearly 20 years ago clinical depression was suggested to arise from cytokines released from activated macrophages (Smith, 1991). This state is sometimes observed in Alzheimer's disease, and argued to be a constituent of the syndrome itself, rather than an understandable secondary consequence of realizing that one has a diagnosis of Alzheimer's disease (Gohier et al., 2009; Korczyn & Halperin, 2009; Sierksma et al., 2010). Likewise, cognitive impairment is recognized to be associated with major depressive disorder, with poor cognitive performance sometimes outlasting the period of depression (Baune et al., 2010). Consistent with a functional link between the two states, there is, as reviewed (Raison et al., 2006; Dantzer et al., 2008; McAfoose & Baune, 2009; Khairova et al., 2009), a considerable literature connecting cytokines such as TNF with the pathogenesis of major depression. Increased serum levels of a range of inflammatory cytokines, including TNF, have been documented in patients (Hestad et al., 2003; Kim et al., 2007; Himmerich et al., 2008). So too have serum levels of soluble TNF receptors, which are less labile, and therefore more reliable, markers of enhanced TNF activity (Himmerich et al., 2008; Grassi-Oliveira et al., 2009; Diniz et al., *in press*). Work on documenting TNF increase in this disorder has since been extended, with the same conceptual outcome, to samples of various brain regions collected at autopsy (Dean et al., 2010). Indeed, a recent meta-analysis covering 24 studies has reported significantly higher concentrations of TNF and IL-6 in depressed subjects compared with control subjects (Dowlati et al., 2010).

Although depression can reasonably be viewed as a manifestation of brain function, it is, as noted (Smith, 1991), significantly associated with coronary heart disease, rheumatoid arthritis, stroke and other diseases, systemic or cerebral, where macrophage activation occurs. This harks back to aspects of Section 4 of this review, in which we note the ways in which systemic and cerebral consequences of macrophage activation are linked. Serum versus CSF levels of TNF appear not to have been studied systematically in major depression, although a recent publication reported CSF levels of IL-6, a cytokine downstream of TNF and IL-1 $\beta$  (i.e. induced by them), was higher in depressed suicide attempters than in normal controls (Lindqvist et al., 2009). Among their data was an intriguing parallel with the much more acute condition of cerebral malaria (John et al., 2008b). While, in each case, plasma cytokine levels were higher than those in the CSF, the two did not correlate, implying independent origins.

It is therefore instructive to consider what is known about the inhibitory effects of some established anti-depressant drugs on TNF and related cytokines. Moclobemide, in clinical use in many countries for this purpose, and widely known as a reversible inhibitor of monoamine oxidase A, significantly suppresses the unstimulated *in vitro* production of TNF and IL-8 in monocytes from human

volunteers (Lin et al., 2000). Another example is bupropion, an aminoketone used as an anti-depressant, largely because it inhibits re-uptake of norepinephrine and dopamine. But in addition bupropion demonstrates, arguably through increasing cyclic AMP, an ability to suppress TNF and IL-1 $\beta$  production so strongly that it protects mice against an otherwise lethal dose of LPS (Brustolim et al., 2006). Furthermore, imipramine, the first tricyclic antidepressant to be developed, and known to influence much cerebral biochemistry, strongly inhibits TNF and IL-1 $\beta$  production from human monocytes (Xia et al., 1996). In the light of present knowledge linking these cytokines and cognitive performance, this inhibition could explain a report that imipramine enhanced learning and memory while being used to treat patients suffering from major depression (Peselow et al., 1991). A recent animal study consistent with this outcome describes imipramine, concomitant with its capacity to prevent A $\beta$ -induced increases in cortex and hippocampal TNF levels, influenced APP processing (see Section 5.2.2.) and long-term memory impairment caused by A $\beta$  treatment (Chavant et al., 2010). For these reasons the authors propose that imipramine should be trialed in patients as an anti-Alzheimer's agent. Thus this agent joins tacrolimus and sirolimus (Section 6.1.4) and rosiglitazone (Section 6.1.5) as pharmaceuticals at present in widespread use for purposes other than their documented capacity to inhibit TNF production, that have been advocated to warrant human Alzheimer's trials.

In keeping with this anti-TNF interpretation of the actions of these drugs usually regarded as anti-depressive agents, a recent publication has reviewed evidence from clinical trials evaluating the efficacy of TNF antagonists in routine clinical use (i.e. adalimumab, etanercept, and infliximab) on depressive symptoms and mental health-associated quality of life measures (Soczynska et al., 2009). While concerned with bipolar disorder (bipolar depression), more than major depression (sometimes termed unipolar depression), the theme of this extensive review is the importance of increased levels of TNF in causing these disease states, and of restoring homeostasis as a basis for their treatment – in other words, returning this cytokine and therefore its downstream mediators to physiological levels.

#### 5.5. Aggression and TNF

Anger and aggression are well-documented aspects of Alzheimer's disease (Ballard & Walker, 1999), brain trauma (Rao et al., 2009), major depression (Keilp et al., 2006) and the post-cerebral malaria syndrome (Bangirana et al., 2009). Indeed, a recent malaria paper describes a proportion of such patients attacking their peers and throwing stones at people and cars with little or no provocation (Idro et al., 2010). TNF has been experimentally associated with aggression in human subjects (Suarez et al., 2002, 2004) and mice selected for non-aggression express lower levels of inflammatory cytokines (Petitto et al., 1994). In addition, aggression in Alzheimer's is more pronounced in patients expressing apoE4 (Craig et al., 2004; van der Flier et al., 2007), an apolipoprotein J variant associated with more TNF generation (see Section 5.2.5). Moreover, the well-known outcome, in a proportion of patients, of anger and aggression when interferon- $\alpha$  is used to treat hepatitis C (Kraus et al., 2003; Preau et al., 2008) has recently been associated with possession of the A allele in the –308 promoter region of the TNF gene (Lotrich et al., 2010), which causes higher TNF production (Sallakci et al., 2005).

Although, as referred to throughout this review, the above disease states in which anger and aggression can occur are circumstances in which TNF is increased, and sometimes have shown positive associations between the two (Suarez et al., 2002, 2004), until now no firmer evidence has been available. However, a recent report has documented the absence of standard measures of aggressive behavior in double TNF receptor knock-out mice – i.e., animals quite unable to use TNF – with aggressive posturing (sideways posture and tail rattling) and bouts of fighting totally lacking when confronted by

previously unseen mice (Patel et al., *in press*). Clearly, this has implications for understanding and treating all of the diseases noted in this section.

## 6. Treating tumor necrosis factor encephalopathies

### 6.1. Indirectly inhibiting TNF production

A rationale for neutralizing TNF indirectly, through inhibiting the pathway that generates it, depends on potential advantages based on either cost, administration by a more convenient route, or both. Disadvantages include having other actions as well, and not being as potent as agents that specifically neutralize TNF itself. Nevertheless the approaches summarized in Section 6.1 are extremely important, since the direct-acting biological therapies discussed in Section 6.2, though likely to be more efficacious, are beyond the means of millions who suffer from cerebral malaria and Alzheimer's disease. Moreover, these biologicals probably require administration by more highly skilled staff than are available. Therefore it is hoped that Section 6.1 might encourage more developmental work with the agents discussed below.

#### 6.1.1. Thalidomide

While thalidomide is notorious for its teratogenic effects, its potential usefulness as an anti-inflammatory agent in patients other than child-bearing age women has been widely explored. Its major property of interest is its capacity to reduce TNF levels (Moreira et al., 1993; Bauditz et al., 2002), and it protects mice against LPS shock (Moreira et al., 1997), largely a TNF-mediated phenomenon. There is considerable research on generating thalidomide derivatives to focus and refine its anti-TNF properties (Hashimoto, 1998; Greig et al., 2004; Alexandre-Moreira et al., 2005), chiefly for use in conditions such as graft-versus-host disease (Holler et al., 1995), rheumatoid arthritis (Ossandon et al., 2002) and Crohn's disease (Facchini et al., 2001), in which anti-TNF biologicals are successful human treatments. Its application in neurodegenerative diseases is also being explored (Tweedie et al., 2007). In particular, thalidomide has been shown to inhibit development of a range of harmful changes caused within 7 days in mice receiving a single injection of A $\beta$ <sub>42</sub> directly into the hippocampus. This short-term model, also used by others (Dinamarca et al., 2006), is acknowledged to be less than ideal for the realities of Alzheimer's disease, but it is widely accepted to be central to the process (but see Simon et al. in Section 5.2.1). The pathology inhibited by thalidomide included vascular abnormalities, microgliosis and astrogliosis, a greatly increased message signal for TNF in microglia, and neuronal death (Ryu & McLarnon, 2008). As well as making a useful argument for this agent and its variants continuing to be developed in this context, these data add further weight to the rationale of treating Alzheimer's diseases by directly inhibiting TNF. We are aware of only one instance of thalidomide being tested in malaria encephalopathy, in a mouse model (Muniz-Junqueira et al., 2005). It was reported to improve survival.

#### 6.1.2. CO and H<sub>2</sub>S donors, resveratrol, and curcumin

As discussed (Sections 3.4.1 and 3.4.2) CO and H<sub>2</sub>S donors act on the same mitochondrial enzyme, and through it are toxic in large doses, while having a host of useful modulatory functions, yet to be appreciably explored, in lower doses. It is part of the nature of inflammatory mediators, among which these two gases must be counted, that they are essentially homeostatic, but decidedly harmful in excess. Nevertheless, the concept of these agents as treatments has its genesis in a highly exciting and relatively untouched area of basic biomedical research, capable of producing concepts often able to surprise us. For example these gases are an excellent window on the function of mitochondria, to which much of the pathogenesis of cytokine-induced brain dysfunction is, in our view, ultimately bound.

Resveratrol is an intriguing polyphenol of plant origin reported to inhibit TNF production in microglia (Bi et al., 2005) and monocytes (Shen et al., 2009), and its association with SIRT1 in these studies (see Section 3.5) suggests that it mirrors how caloric restriction reduces TNF and enhances brain function. Since the literature on caloric restriction influencing brain is so impressive, resveratrol warrants close examination in an Alzheimer's context. Moreover, there are two recent reports of its efficacy in brain trauma (Ates et al., 2007; Sonmez et al., 2007), a condition linked to TNF excess, as is Alzheimer's (see Section 4.3).

Curcumin, an extract of the Indian spice, tumeric, has been mooted as a general anti-inflammatory agent for some time. In terms of Alzheimer's disease, proposed mechanisms of action have been as an anti-oxidant (Calabrese et al., 2003), protecting against A $\beta$  because it is a metal chelator (Baum & Ng, 2004), preventing A $\beta$  fibrils from forming (Ono et al., 2004), and suppressing NF $\kappa$ B (Aggarwal & Shishodia, 2004). We have included it here because of this last-listed property, which ensures reduced generation of certain cytokines, including TNF. This is consistent with, for instance, its capacity to inhibit A $\beta$ -induced neurotoxicity (Park et al., 2008), which the literature points to being TNF mediated (Wang et al., 2005; Ralay Ranaivo et al., 2006). Nevertheless, a 12 month, randomized, placebo-controlled study of oral curcumin for mild-to-moderate Alzheimer's disease was reported to have failed to show benefit (<http://www.clinicaltrials.gov/ct2/show/NCT00099710>). In passing, it might be an interesting exercise to see if populations reknown for tumeric consumption have a lower incidence of Alzheimer's disease.

#### 6.1.3. Intravenous immunoglobulin

Much plasma collected by blood banks is under-utilized, so immunoglobulin preparations have a long history of being administered intravenously to see if they ameliorate disease states. These include not just the hypo- and agammaglobulinemia disorders, but conditions such as disseminated intracellular coagulopathy and severe inhibition bacterial and viral diseases have been treated "off label". As reviewed (Rosen, 1993), it was soon realized that as well as providing more total immunoglobulin, this therapy caused changes to the mediators common to innate immunity and inflammation through containing natural autoantibodies against IL-1 (Gallay et al., 1991), IFN- $\gamma$  (Toungouz et al., 1996) and TNF (Simon & Spath, 2003). Of particular relevance to this review, TNF was found to be reduced (Achiron et al., 1994; Andersson et al., 1996; Darville et al., 1997; Sharief et al., 1999; Gupta et al., 2001; Kishimoto et al., 2003; Asakura et al., 2006). Another group reported an increase of IL-10 and soluble TNF receptor, which would have the same outcome as a fall in TNF (Gullestad et al., 2001).

Unfortunately, these cytokines, the mediators of both innate immunity or inflammation, were still under the radar for most Alzheimer's researchers when the above studies were published. Therefore, when intravenous immunoglobulin began to be used to treat this disease, the accepted paradigm for this disease (Weksler et al., 2005) simply required A $\beta$  levels to be reduced to produce a positive outcome (Dodel et al., 2004; Relkin et al., 2009). Some clinical efficacy may have been apparent in preliminary trials, and a larger trial is currently recruiting patients (<http://www.clinicaltrials.gov/ct2/show/NCT00812565>), but the anti-TNF effects (above) of these agents do not yet appear have been taken into account. Nevertheless, the goal of reducing A $\beta$  levels has been thrown into question by the two concepts discussed earlier: that TNF-induced APP can be expected to cause harm before A $\beta$  forms (Simon et al., Section 5.2.1.), and that A $\beta$ , once formed from APP cleavage, may actually be a useful mediator of cerebral innate immunity (Soscia et al., Section 5.2.3.). If reappraised in cytokine terms intravenous immunoglobulin might, in theory, be a useful way to indirectly reduce TNF, but it has renal toxicity (Graumann & Zawada, 2010), and its cost structure to contend with.

#### 6.1.4. Tacrolimus (FK506) and sirolimus (rapamycin)

Tacrolimus is widely used clinically as an immunosuppressive agent, chiefly to prevent rejection of organ transplants. It is thought to function through inhibiting the activity of calcineurin (protein phosphatase 2B), a widespread enzyme that, apart from involvement in T cell activation, is the most abundant phosphatase in the central nervous system (Mansuy, 2003). The activity of calcineurin most relevant to the purpose of this review is its capacity to activate a transcription factor termed nuclear factor of activated T cells (NFAT) (Clipstone & Crabtree, 1992). Thus tacrolimus inhibits the accumulation of IL-2 mRNA, as well as the mRNAs of other early phase T cell activation genes such as IL-3, IL-4, GM-CSF, TNF alpha, IFN-gamma, and c-myc in activated human peripheral blood T cells (Tocci et al., 1989). NFAT activation also occurs in the brain, with inflammatory mediators such as TNF recruiting astrocyte calcineurin to stimulate an inflammatory pathway involving this transcription factor (Fernandez et al., 2007). In addition expression of TNF in neurons has been reported to involve the calcineurin-NFAT pathway (Canellada et al., 2006). Thus, by suppressing calcineurin, tacrolimus predictably inhibits TNF production. Indeed, this inhibition is a widespread phenomenon (Kawano et al., 1992; Goldfeld et al., 1992; Sakr et al., 1993; Kawano et al., 1994; Squadrito et al., 1999; Sakuma et al., 2000). In addition, inflammatory cytokines, including TNF, induce various calcineurin-dependent activities (Grunnet et al., 2009), so inhibiting calcineurin can be expected to inhibit these down-stream functions of TNF.

Hence the use of tacrolimus and anti-TNF agents in order to produce the same clinical outcomes is logical, with graft-versus-host disease (Jacobsohn & Vogelsang, 2002), inflammatory bowel disease (Legnani & Kornbluth, 2004), organ transplant (Pascher et al., 2005), and rheumatoid arthritis (Miyata et al., 2005) being examples. In a limited comparative literature, tacrolimus appears to be slightly less efficacious than anti-TNF agents, in that infliximab proved to rescue only about a quarter of ulcerative colitis patients refractory to tacrolimus (Herrlinger et al., 2010). Tacrolimus shares with the anti-TNF agents a long history of patient use, but as noted above (Tocci et al., 1989) it inhibits a number of important cytokines in addition to TNF, the implications of which would require exploring before considering long-term use. It could be counterproductive, for instance, to downregulate the anti-inflammatory cytokine IL-4 (Tocci et al., 1989) during a chronic inflammatory disease, but acceptable in a more acute event such as brain trauma or septic encephalopathy.

Nevertheless, of all the possibilities for inhibiting TNF indirectly, tacrolimus appears to be the most promising on current data. Of particular relevance to this review, it has been reported to restore associative learning and memory (Dineley et al., 2007) and reverse novel object recognition memory deficits (Tagliatalata et al., 2009) in a murine Alzheimer's disease model. Certainly, from a basic science perspective the tacrolimus data are important because they add further weight to TNF being a prime therapeutic target. However, as recently reviewed (Naesens et al., 2009), it is well documented that the use of tacrolimus carries a risk of nephrotoxicity, presumably limiting its long-term use.

Siromilus, a macrolide antibiotic generated by *Streptomyces hygroscopicus*, was first identified as an immunosuppressant about a decade ago (Kahan et al., 1991), and, as reviewed (Weir et al., 2010), has been used alone or in conjunction with tacrolimus in recent years in order to lower overall toxicity. Sirolimus has been shown to possess a range of effects that interfere with activation of the immune system, and crosses paths with tacrolimus in the complexity of the immunoregulatory functions of mTOR (mammalian target of rapamycin) inhibition (Thomson et al., 2009). In outline, both tacrolimus and sirolimus are ligands for FKBP12, (FK506 binding protein 12), an isomerase involved in a range of basic cellular processes as well as immunosuppression. In addition, it is involved in LTP, memory, and

perseverative behaviors (Hoeffler et al., 2008). One of the many consequences of these shared actions is that sirolimus, like tacrolimus, inhibits TNF production (Gabryel et al., 2004; Adkins et al., 2004; Lorne et al., 2009; Wang et al., 2009a; Krakauer et al., 2010; Weir et al., 2010). Again like tacrolimus, sirolimus has recently been reported to rescue cognitive deficits in mouse models of Alzheimer's disease (Caccamo et al., 2010; Spilman et al., 2010). In the light of the literature we have summarized, it seems a logical extension of these data to propose that the reduction in TNF caused by these two agents could be sufficient to explain their *in vivo* activities in these Alzheimer models. Thus tacrolimus and sirolimus, two agents presently in use for their immunosuppressive activity, join imipramine, an anti-depressant (Section 5.4), as drugs already in clinical use that, though their TNF-suppressing qualities, possess plausible cases for testing against the encephalopathies discussed in this review.

As discussed (Krakauer et al., 2010), toxicity might well preclude the use of siromilus (rapamycin) in states requiring long-term treatment, but short-term treatment in brain trauma and certain encephalopathies induced by infectious agents show brighter prospects. However, these indirect approaches to inhibiting TNF production approaches are unlikely to approach the efficacy of specific TNF inhibitors. Thus they are likely to be hard pressed, in life-threatening encephalopathies, to equal the outcome possible by targeting pre-formed TNF directly, as discussed below.

#### 6.1.5. Rosiglitazone, a PPAR- $\gamma$ agonist

The peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ) is a member of the nuclear receptor superfamily of ligand-dependant transcription factors. They have naturally occurring, as well as synthetic ligands, the latter having been developed for therapeutic use. Rosiglitazone, a thiazolidinedione, is one such agent, and is commonly used to counter the insulin resistance seen in type 2 diabetes. This property was recognized (Oakes et al., 1994) a few years after TNF had been reported to cause insulin resistance (Lang et al., 1992), but the notion of these PPAR- $\gamma$  ligands acting through inhibiting TNF release had to wait another six years, when they were shown to down-regulate macrophage activation (Ricote et al., 1998) and impair the production of inflammatory cytokines, including TNF, from monocytes (Jiang et al., 1998). This activity of these agents was soon reported to extend to the brain (Heneka et al., 2000). In that same year this class of drugs, the glitazones, were demonstrated to reduce the ability of both LPS and A $\beta$  to activate monocytes and microglia, and inhibit TNF production from cells of this lineage (Combs et al., 2000).

While these researchers subsequently argued the case for glitazones as possible Alzheimer's therapies through their anti-inflammatory properties (Sundararajan et al., 2006), others began mouse experiments (Pedersen et al., 2006) and human trials (Watson et al., 2005; Risner et al., 2006) simply on the rationale that these drugs reverse insulin resistance, their originally described property, without any role attributed to TNF. In the same year as Risner's publication appeared, insulin resistance in rheumatoid arthritis patients proved reversible by treatment with a standard dose of specific anti-TNF drug (Gonzalez Gay et al., 2006). This confirmed the earlier *in vitro* observation (Lang et al., 1992), and implies that rosiglitazone can safely be regarded as a TNF inhibitor in an insulin context, as well as more generally (Sundararajan et al., 2006). Indeed, the observation that apoE4 positive individuals did not respond (Risner et al., 2006) is easier to rationalize through activities of TNF (see Section 5.2.5) than its influence on insulin metabolism. Another group (Escribano et al., 2009) have recently reported that rosiglitazone rescued memory impairment in a transgenic mouse model of Alzheimer's disease, and, like Landreth and co-workers (Combs et al., 2000; Sundararajan et al., 2006) have argued the outcome in terms of inhibiting expression of pro-inflammatory cytokines, such as TNF, for which they assayed. The changes they observed in A $\beta$  and tau can be

accounted for, through TNF reduction, in observations summarized in Sections 5.1.1 and 5.2.2. Likewise, recent data on rosiglitazone improving survival in malaria (Serghides et al., 2009) are based entirely on the rationale of the observed reduction in TNF levels. We have similar data from experiments, with the same rationale, with highly virulent mouse influenza models with gemfibrozil, a functionally similar PPAR- $\alpha$  agonist (Budd et al., 2007) and rosiglitazone (Alleva, L., unpublished). The pathogenesis of neither malaria nor influenza has, or is likely to have in future, a history of being explained in terms of A $\beta$ -origin pathology or insulin tolerance.

A larger phase III trial of rosiglitazone is presently in progress in Alzheimer's patients (see <http://www.alzforum.org/drg/drc/detail.asp?id=116>), whereas imipramine (Section 5.4), tacrolimus or sirolimus (Section 6.1.4), have not yet reached this stage of testing. In summary, the property these four agents share of inhibiting TNF production means that, inadvertently or otherwise, those conducting such trials are testing the consequence of reducing levels of this cytokine. From precedences from other disease states, such as rheumatoid arthritis, outcomes can be expected to be less satisfactory, with more side effects, than when specific purpose-designed anti-TNF agents are employed, as described in the next Section.

## 6.2. Specifically neutralizing TNF once produced

### 6.2.1. Acute systemic infections

The broad evidence summarized in the preceding pages clearly points to treating encephalopathies by neutralizing excess TNF. Conceptually, at least in other organs, this is now an old idea. As has been reviewed (Clark, 2007), once TNF was sequenced (Aggarwal et al., 1985), and cachectin appreciated to be the same molecule as TNF (Beutler et al., 1985), there was soon no doubt that TNF is what causes death from injected LPS, since neutralizing anti-TNF antibody, given before LPS, prevents death in mice (Beutler et al., 1985; Waage et al., 1987). Many early studies in mice demonstrated that serum TNF peaks at 90 min after intravenous LPS, when the animals are not yet particularly sick. In other words, TNF rises before onset of appreciable illness (Waage et al., 1987), so the best time to administer an anti-TNF agent has passed once severe illness is present. Thus it soon became evident that in acute illness the therapeutic potential of these TNF-neutralizing agents is largely limited to circumstances in which the acute cytokine-induced illness was an unavoidable consequence of necessary treatment. Accordingly, positive results in patients were reported in prophylactic OKT3 therapy, which minimizes graft rejection (Chatenoud, 1993; Eason et al., 1996), and graft-versus-host disease (Holler et al., 1995; Jacobsen et al., 1992). Likewise, anti-TNF prevents the human Jarisch-Herxheimer reaction caused by penicillin administered to kill *Borrelia recurrentis*, an organism high in LPS, and the cause of louse-borne relapsing fever, but only provided it is given before the penicillin, i.e. before TNF is induced (Fekade et al., 1996). Given this background, it is not surprising that administering anti-TNF antibody disappointed those using it against falciparum malaria (Kwiatkowski et al., 1993; Van Hensbroek et al., 1996; Looreesuwan et al., 1999) or human sepsis (Abraham et al., 1998; Dinarello, 2001).

### 6.2.2. Chronic non-cerebral and cerebral inflammatory states

During the period that the disappointing outcome of anti-TNF antibody in treating acute infections was emerging, a group in London working on rheumatoid arthritis found that TNF might be a more important mediator than IL-1 in this context, since *in vitro* anti-TNF antibody reduced IL-1 as well as TNF (Brennan et al., 1989). The implications of this for treating rheumatoid arthritis (Feldmann et al., 1990) led to a successful open trial being published a few years later (Elliott et al., 1993). This was opportune timing, in the sense that the pharmaceutical company with which they were involved was one of many looking for new uses for a product that had become an orphan

due to its failure as a treatment for sepsis. Not surprisingly they readily funded a blinded trial, the successful outcome of which appeared the next year (Elliott et al., 1994). This paved the way for the interest of others in treating other non-systemic chronic states with anti-TNF agents, and nowadays patients suffering from Crohn's disease, psoriasis, and ankylosing spondylitis as well as rheumatoid arthritis are treated with infliximab (Remicade), a monoclonal murine-human chimeric antibody to TNF, etanercept (Enbrel), a fusion protein of two p75 chains of the TNF receptor II and the Fc portion of IgG1 and adalimumab (Humira), a fully human anti-TNF monoclonal antibody, on a success and scale, and with financial gain on the part of the patent owners and manufacturers, unique in the biotechnology industry. It has not all been plain sailing, with exacerbation of some, but by no means all, pre-existing infections (Section 6.3). Details of this, and differences between the molecules and their mode of action, have recently been reviewed extensively (Tracey et al., 2008; Esposito & Cuzzocrea, 2009; Furst et al., 2010). Other anti-cytokine agents under development for this purpose are also discussed in these reviews.

It seems that the distinctively long duration of TNF increase in the brain during systemic inflammatory responses (Section 4.5) is a reflection of the dependence of the health of the central nervous system on the innate immune response. Thus diseases of this organ are a prime target for anti-TNF therapeutic agents should its normal surveillance by cells of bone marrow origin (Butovsky et al., 2007a, 2007b; Ron-Harel et al., 2008), with anti-inflammatory activity that is mediated chiefly through IL-10 (Koronyo-Hamaoui et al., 2009), fail. This surveillance has recently been reviewed (Ron-Harel & Schwartz, 2009; Schwartz, *in press*).

### 6.2.3. Getting anti-TNF to where it is needed in encephalopathies

Since anti-TNF agents are administered parenterally in the non-cerebral diseases discussed above, they unsurprisingly encounter no particular anatomical barrier preventing access to the site of pathology. Clearly, a drawback with testing these agents against putative TNF-driven encephalopathies is that they are about as large as albumin and globulin, so cannot be expected to pass the blood-brain barrier efficiently, where, as discussed in various Sections of this review, much evidence is consistent with excess TNF being harmful. One group has addressed the challenge of brain access through adapting a lesson from the basic aviation medicine literature, in which the effects of 5 min of head-down positioning allows appreciable albumin and globulin to enter the CSF, probably through the choroid plexus (Wen et al., 1994). Indeed, these authors suggested this manoeuvre as a way to get large molecules in to the brain for therapeutic purposes. Combined with an injection route for etanercept that drains into Batson's plexus, this has been used with the intention of accessing the CSF via the choroid plexus. In 2006 an open trial with etanercept injected by this route, in which 15 Alzheimer's patients were treated for 6 months, was published (Tobinick et al., 2006), as have case reports since (Tobinick & Gross, 2008). Etanercept administered in this way has recently been shown to reach the cerebral ventricles in the rat (Tobinick et al., 2009). This argues that this route is much less invasive but functionally equivalent to i.c.v., which is routinely used for anti-TNF biological agents of this size to access the CSF in basic animal studies on roles of TNF in brain function (Medeiros et al., 2007; Galic et al., 2008; Riazzi et al., 2008; Liesz et al., 2009). We note that a similar challenge is addressed when treating brain-sited lymphoma with rituximab, a monoclonal antibody effective against non-Hodgkin lymphoma, which expresses CD20, the target of the antibody. Unfortunately its normal intravenous route for non-cerebral lymphomas is ineffective (Rubenstein et al., 2003), so intraventricular administration was seen as the logical alternative. It was successful in a Phase I trial (Rubenstein et al., 2007), but it would be of practical interest to establish whether the less invasive Batson's plexus route, above, will prove to be equally efficacious. Arguments in

favour of this route in neutralizing brain TNF have recently been reviewed (Tobinick, 2010).

Despite the direction of the literature, calls for a double-blind human Alzheimer's trial using etanercept by this route (Tobinick et al., 2006) have not yet attracted industry or government funding. This is in marked contrast to the readiness, in 1993 (see Section 6.2.2), of the manufacturers of another of the above anti-TNF agents to expand a very similar open trial on rheumatoid arthritis (Elliott et al., 1993) into a double-blind study (Elliott et al., 1994). We note, however, that an essentially identical trial except it is to employ etanercept subcutaneously, is soon to begin (see <http://www.clinicaltrials.gov/ct2/show/NCT01068353>). In view of the albumin-like size of this biomolecular construct, and a previous blinded trial (fewer subjects, but over 24 weeks) employing this route having a negative outcome (Bohac et al., 2002), giving funding precedence to a trial using this route is surprising. The challenge in getting etanercept to where it matters in Alzheimer's disease is, in addition to the rituximab example (previous paragraph), also addressed by studies in which intravenous infliximab, a similar sized molecule, can be presumed not to have entered the CSF because TNF levels in this compartment remained unchanged (Andersson et al., 2006; Kikuchi et al., 2008).

In 2003 a promising high-affinity inhibitor of human TNF was discovered to be secreted by Tanapox virus (Brunetti et al., 2003). From its molecular weight (Rahman et al., 2006), its inability to pass the blood brain barrier will still be a concern for using this protein to treat cerebral conditions, but its extraordinarily high activity compared to neutralizing specific antibodies against TNF has encouraged commercial development as VT-346, with Alzheimer's disease as well as rheumatoid arthritis and Crohn's disease the stated targets (March 16 2010 press release at [www.vironinc.com](http://www.vironinc.com)). Others (see <http://www.bionity.com/news/e/107293>) are developing TNF neutralizing antibodies, of a type (Harmsen & De Haard, 2007) small enough to pass the blood brain barrier, for systemic use in human disease. These nanobodies are an attractive second generation drug, but like VT-346 are some years away from clinical trials. In contrast, infliximab, etanercept and adalimumab, already administered to millions of people annually in relative safety, have an immediate attraction for dealing with the large-scale clinical challenge already present, particularly in Alzheimer's disease, provided it actually gets into the CSF.

### 6.3. Avoiding exacerbating infections

Over 25 years ago, when working with purified TNF in the days before recombinant proteins, our laboratory argued that TNF was central to both disease and cell-mediated immunity (Clark et al., 1981), as recently reviewed (Clark, 2007). For example, TNF inhibits malaria parasites (Taverne et al., 1981; Clark et al., 1987), *Mycobacterium* spp. (Bermudez & Young, 1988), *Salmonella typhimurium* (Degre & Bukholm, 1990), *Leishmania* spp. (Titus et al., 1989), *Toxoplasma gondii* (Chang et al., 1990), *Coxiella brunetii* (Tokarevich et al., 1992), and *Listeria monocytogenes* (Rothe et al., 1993). It was always inevitable, therefore, that dormant infections with these organisms had the potential to flare when anti-TNF agents were chronically administered to, for example, rheumatoid arthritis patients (Keane et al., 2001; Slifman et al., 2003; Rijkeboer et al., 2007). Hence patients are nowadays tested, monitored, and if necessary treated for these conditions before and while undergoing anti-TNF therapy. While this is a real phenomenon, it should nevertheless be kept in perspective, in that a recent consensus document puts the risk rate for all bacterial diseases for TNF neutralizing agents as 0.07–0.09 per patient year, compared to 0.06 for all other disease modifying anti-rheumatic drugs (Furst et al., 2010). As reviewed in 2008 (Domm et al., 2008), the influence of anti-TNF therapy on concomitant viral diseases seems to have been surprisingly mild, and there are as yet very few guidelines on whether

or not to screen, and for what conditions. Outcomes of monitoring for hepatitis B, hepatitis C, HIV, herpes, E-B virus, cytomegalovirus, and varicella zoster virus are discussed by these authors. Since this publication single cases of cytomegalovirus retinitis (Haerter et al., 2004) and, of more concern, H1N1 influenza (Kling et al., 2010) have been reported. Since the total number of people receiving regular dosage with these agents probably now approaches 10 million, individual cases are inevitably difficult to interpret.

The degree of this concern in other encephalopathies depends, of course, on the circumstance. Neurodysfunction after brain trauma or myocardial infarct would present the least risk, whereas after surgery or burns opportunist TNF-sensitive organisms are more of a possibility. An encephalopathy associated with a known systemic infection (Sections 4.4 and 4.5) caused by an agent sensitive to an antimicrobial, in the widest sense, need not be as much concern as an unsuspected dormant infection when long-term treatment is envisaged. For example, the point has been made that anti-TNF did not reduce the rate of parasite clearance after antimalarial treatment (Looareesuwan et al., 1999). This implies that anti-TNF agents could be safely administered, along with antimalarials, in trials against long-term malaria-induced cognitive defects (Kihara et al., 2006). Alzheimer's disease would be expected to have the same need for testing and monitoring for dormant infections such as tuberculosis and salmonellosis, as does rheumatoid arthritis.

### 7. The perceived advantages of normalizing exacerbated cerebral levels of tumor necrosis factor

The evidence summarized in this review indicates that the advantages of reducing pathologically increased brain levels of TNF in the brain dysfunctions and diseases we have discussed are manifold. Most typically, cerebral APP can be expected to be reduced, so it does not reach levels expected to be harmful in adults because they activate TNFRSF21 (DR6) and thus thin out neurons and axons (Nikolaev et al., 2009), as is normal in early neonates. The rate of cleavage of APP to A $\beta$ <sub>42</sub> can also be expected to be reduced (Liao et al., 2004; Yamamoto et al., 2007), and, through reducing the effects of TNF induced by A $\beta$ <sub>42</sub> (Wang et al., 2005; Ralay Ranaivo et al., 2006), so too would be many of the effects of A $\beta$ <sub>42</sub> on synapses and small cerebral blood vessels. In addition, inhibitory functional effects of excessive TNF on the interaction between glial cells and neurons can be expected to be inhibited, neurogenesis enhanced (Wu et al., 2000), synaptic zinc availability increased (Section 3.2), and effects of cerebral hypoxia minimized (Aden et al., 2010).

In contrast, 17 years ago, when rheumatoid arthritis treatment with infliximab was tested in a blinded control trial, very little, inevitably seen as simplistic by today's literature, was appreciated about how it might work, and key mechanisms actually came to light later (Feldmann & Maini, 2003). The same will inevitably be true for the encephalopathies, but we can safely say we know a lot more to begin with.

Finally, we again draw attention to the views expressed by two clear thinkers that their respective fields, cerebral malaria (Maegraith, 1948) and Alzheimer's disease (Castellani et al., 2008) were being led astray by what was historically obvious down a microscope, and that, in their words, invisible molecules and subcellular damage are probably what matter more. Eureka moments can often be quietly unwelcomed by many researchers because of the death knell they sound for popular paradigms, but in the long run they are very necessary. Just as work from New Guinea served that purpose in cerebral malaria (Genton et al., 2008; Tjitra et al., 2008), the combined implications of the APP/DR6 (Nikolaev et al., 2009) and innate immunity/A $\beta$  (Soscia et al., 2010) studies promise to do so in Alzheimer's disease.

We trust that our background in the roles of TNF and similar mediators in physiology, pathology and innate immunity has allowed

us to strengthen these cases, and thus help focus minds on rational treatment for this group of functionally-related diseases. We stress that to feel at home thinking about mediators such as TNF one must appreciate, in some depth, their primary, essential, role in basic physiology, examples relevant here being synapse homeostasis and setting the diurnal clock. From time to time, when a primitive signal is recognized, they also set innate immunity in motion. Less frequently, these signals are so strong that they provoke the host to overdo it, causing what we recognize as either organ-specific or systemic pathology, and have discussed here. Finally, treatment is about ameliorating this overblown response, not eliminating the cytokine as if it were some exogenous poison.

## References

- Aarden, L. A., & others. (1979). Revised nomenclature for antigen-nonspecific T cell proliferation and helper factors. *J Immunol* 123, 2928–2929.
- Abdipranoto, A., Wu, S., Stayte, S., & Vissel, B. (2008). The role of neurogenesis in neurodegenerative diseases and its implications for therapeutic development. *CNS Neurol Disord Drug Targets* 7, 187–210.
- Abdipranoto-Cowley, A., Park, J. S., Croucher, D., Daniel, J., Henshall, S., Galbraith, S., et al. (2009). Activin A is essential for neurogenesis following neurodegeneration. *Stem Cells* 27, 1330–1346.
- Abraham, J., & Johnson, R. W. (2009). Consuming a diet supplemented with resveratrol reduced infection-related neuroinflammation and deficits in working memory in aged mice. *Rejuvenation Res* 12, 445–453.
- Abraham, E., Anzueto, A., Gutierrez, G., Tessler, S., Sanpedro, G., Wunderink, R., et al. (1998). Double-blind randomised controlled trial of monoclonal antibody to human tumour necrosis factor in treatment of septic shock. *Lancet* 351, 929–933.
- Abramov, E., Dolev, I., Fogel, H., Ciccotosto, G. D., Ruff, E., & Slutsky, I. (2009). Amyloid-beta as a positive endogenous regulator of release probability at hippocampal synapses. *Nat Neurosci* 12, 1567–1576.
- Achiron, A., Margalit, R., Hershkovitz, R., Markovits, D., Reshef, T., Melamed, E., et al. (1994). Intravenous immunoglobulin treatment of experimental T cell-mediated autoimmune disease. Upregulation of T cell proliferation and downregulation of tumor necrosis factor alpha secretion. *J Clin Invest* 93, 600–605.
- Aden, U., Favrais, G., Plaisant, F., Winerdal, M., Felderhoff-Mueser, U., Lampa, J., et al. (2010). Systemic inflammation sensitizes the neonatal brain to excitotoxicity through a pro-/anti-inflammatory imbalance: Key role of TNF pathway and protection by etanercept. *Brain Behav Immun* 24, 747–758.
- Adkins, J. R., Castresana, M. R., Wang, Z., & Newman, W. H. (2004). Rapamycin inhibits release of tumor necrosis factor-alpha from human vascular smooth muscle cells. *Am Surg* 70, 384–387.
- Aggarwal, B. B., Henzel, B. J., Moffat, B., Kohr, W. J., & Harkins, R. N. (1985). Primary structure of human lymphotoxin derived from 1788 lymphoblastoid cell line. *J Biol Chem* 260, 2334–2344.
- Aggarwal, B. B., & Shishodia, S. (2004). Suppression of the nuclear factor-kappaB activation pathway by spice-derived phytochemicals: reasoning for seasoning. *Ann N Y Acad Sci*, 434–441.
- Aimone, J. B., Wiles, J., & Gage, F. H. (2006). Potential role for adult neurogenesis in the encoding of time in new memories. *Nat Neurosci* 9, 723–727.
- Albani, D., Polito, L., & Forloni, G. (in press). Sirtuins as novel targets for Alzheimer's disease and other neurodegenerative disorders: experimental and genetic evidence. *J Alzheimers Dis*. doi:10.3233/JAD-2010-1215.
- Albensi, B. C., & Janigro, D. (2003). Traumatic brain injury and its effects on synaptic plasticity. *Brain Inj* 17, 653–663.
- Aleardi, A. M., Benard, G., Augereau, O., Malgat, M., Talbot, J. C., Mazat, J. P., et al. (2005). Gradual alteration of mitochondrial structure and function by beta-amyloids: importance of membrane viscosity changes, energy deprivation, reactive oxygen species production, and cytochrome c release. *J Bioenerg Biomembr* 37, 207–225.
- Alexander, J. J., Jacob, A., Cunningham, P., Hensley, L., & Quigg, R. J. (2008). TNF is a key mediator of septic encephalopathy acting through its receptor, TNF receptor-1. *Neurochem Int* 52, 447–456.
- Alexandre-Moreira, M. S., Takiya, C. M., de Arruda, L. B., Pascarelli, B., Gomes, R. N., Castro Faria Neto, H. C., et al. (2005). LASSBio-468: a new achiral thalidomide analogue which modulates TNF-alpha and NO production and inhibits endotoxic shock and arthritis in an animal model. *Int Immunopharmacol* 5, 485–494.
- Alkam, T., Nitta, A., Mizoguchi, H., Saito, K., Seshima, M., Itoh, A., et al. (2008). Restraining tumor necrosis factor-alpha by thalidomide prevents the amyloid beta-induced impairment of recognition memory in mice. *Behav Brain Res* 189, 100–106.
- Andersson, J., Skansen Saphir, U., Sparrelid, E., & Andersson, U. (1996). Intravenous immune globulin affects cytokine production in T lymphocytes and monocytes/macrophages. *Clin Exp Immunol* 1, 10–20.
- Andersson, M., Svenungsson, E., Khademi, M., Lampa, J., Brundin, L., & Wallstrom, E. (2006). No signs of immunoactivation in the cerebrospinal fluid during treatment with infliximab. *Ann Rheum Dis* 65, 1237–1240.
- Anekonda, T. S. (2006). Resveratrol—a boon for treating Alzheimer's disease? *Brain Res Rev* 52, 316–326.
- Angel, K., Provan, S. A., Gulseth, H. L., Mowinckel, P., Kvien, T. K., & Atar, D. (2010). Tumor necrosis factor-alpha antagonists improve aortic stiffness in patients with inflammatory arthropathies: a controlled study. *Hypertension* 55, 333–338.
- Arendt, T. (2009). Synaptic degeneration in Alzheimer's disease. *Acta Neuropathol* 118, 167–179.
- Arendt, T., Stieler, J., Strijkstra, A. M., Hut, R. A., Rudiger, J., Van der Zee, E. A., et al. (2003). Reversible paired helical filament-like phosphorylation of tau is an adaptive process associated with neuronal plasticity in hibernating animals. *J Neurosci* 23, 6972–6981.
- Asakura, H., Takahashi, Y., Kubo, A., Ontachi, Y., Hayashi, T., Omote, M., et al. (2006). Immunoglobulin preparations attenuate organ dysfunction and hemostatic abnormality by suppressing the production of cytokines in lipopolysaccharide-induced disseminated intravascular coagulation in rats. *Crit Care Med* 34, 2421–2425.
- Ates, O., Cayli, S., Altinoz, E., Gurses, I., Yucel, N., Sener, M., et al. (2007). Neuroprotection by resveratrol against traumatic brain injury in rats. *Mol Cell Biochem* 294, 137–144.
- Bachman, D., & Rabins, P. (2006). "Sundowning" and other temporally associated agitation states in dementia. *Annu Rev Med* 57, 499–511.
- Ballard, C., & Walker, M. (1999). Neuropsychiatric aspects of Alzheimer's disease. *Curr Psychiatry Rep* 1, 49–60.
- Bangirana, P., Nakasujja, N., Giordani, B., Opoka, R. O., John, C. C., & Boivin, M. J. (2009). Reliability of the Luganda version of the Child Behaviour Checklist in measuring behavioural problems after cerebral malaria. *Child Adolesc Psychiatry Ment Health* 3, 38.
- Barichello, T., Dos Santos, I., Savi, G. D., Florentino, A. F., Silvestre, C., Comim, C. M., et al. (2009). Tumor necrosis factor alpha (TNF-alpha) levels in the brain and cerebrospinal fluid after meningitis induced by *Streptococcus pneumoniae*. *Neurosci Lett* 467, 217–219.
- Barone, F. C., Arvin, B., White, R. F., Miller, A., Webb, C. L., Willette, R. N., et al. (1997). Tumor necrosis factor-alpha – A mediator of focal ischemic brain injury. *Stroke* 28, 1233–1244.
- Bauditz, J., Wedel, S., & Lochs, H. (2002). Thalidomide reduces tumour necrosis factor alpha and interleukin-12 production in patients with chronic active Crohn's disease. *Gut* 50, 196–200.
- Baum, L., & Ng, A. (2004). Curcumin interaction with copper and iron suggests one possible mechanism of action in Alzheimer's disease animal models. *J Alzheimers Dis* 6, 367–377.
- Baune, B. T., Miller, R., McAfoose, J., Johnson, M., Quirk, F., & Mitchell, D. (2010). The role of cognitive impairment in general functioning in major depression. *Psychiatry Res* 176, 183–189.
- Beare, N. A., Harding, S. P., Taylor, T. E., Lewallen, S., & Molyneux, M. E. (2009). Perfusion abnormalities in children with cerebral malaria and malarial retinopathy. *J Infect Dis* 199, 263–271.
- Becker, M., Lavie, V., & Solomon, B. (2007). Stimulation of endogenous neurogenesis by anti-EFRH immunization in a transgenic mouse model of Alzheimer's disease. *Proc Natl Acad Sci USA* 104, 1691–1696.
- Behr, D., Wu, J., Cumine, S., Kim, K. W., Lu, S. C., Atangan, L., et al. (2009). Resveratrol is not a direct activator of SIRT1 enzyme activity. *Chem Biol Drug Des* 74, 619–624.
- Bell, M. J., & Hallenbeck, J. M. (2002). Effects of intrauterine inflammation on developing rat brain. *J Neurosci Res* 70, 570–579.
- Bemeur, C., Qu, H., Desjardins, P., & Butterworth, R. F. (2010). IL-1 or TNF receptor gene deletion delays onset of encephalopathy and attenuates brain edema in experimental acute liver failure. *Neurochem Int* 56, 213–215.
- Bemis, D. L., Katz, A. E., & Buttyan, R. (2006). Clinical trials of natural products as chemopreventive agents for prostate cancer. *Expert Opin Investig Drugs* 15, 1191–1200.
- Bercik, P., Verdu, E. F., Foster, J. A., Macri, J., Potter, M., Huang, X. et al. (in press). Chronic gastrointestinal inflammation induces anxiety-like behavior and alters central nervous system biochemistry in mice. *Gastroenterology*. doi:10.1053/j.gastro.2010.06.063.
- Bermudez, L. E., & Young, L. S. (1988). TNF, alone or in combination with IL-2, but not IFN-gamma, is associated with macrophage killing of *Mycobacterium avium* complex. *J Immunol* 140, 3006–3013.
- Bernardino, L., Agasse, F., Silva, B., Ferreira, R., Grade, S., & Malva, J. O. (2008). Tumor necrosis factor-alpha modulates survival, proliferation, and neuronal differentiation in neonatal subventricular zone cell cultures. *Stem Cells* 26, 2361–2371.
- Beutler, B., & Poltorak, A. (2001). Sepsis and evolution of the innate immune response. *Crit Care Med* 29, S2–S6.
- Beutler, B., Greenwald, D., Hulmes, J. D., Chang, M., Pan, Y. -C., Mathison, J., et al. (1985). Identity of tumour necrosis factor and the macrophage-secreted factor cachectin. *Nature* 316, 552–554.
- Beutler, B., Gelbart, T., & West, C. (2001). Synergy between TLR2 and TLR4: a safety mechanism. *Blood Cells Mol Dis* 27, 728–730.
- Bhatia, M., Brady, M., Shokui, S., Christmas, S., Neoptolemos, J. P., & Slavin, J. (2000). Inflammatory mediators in acute pancreatitis. *J Pathol* 190, 117–125.
- Bi, X. L., Yang, J. Y., Dong, Y. X., Wang, J. M., Cui, Y. H., Ikeshima, T., et al. (2005). Resveratrol inhibits nitric oxide and TNF-alpha production by lipopolysaccharide-activated microglia. *Int Immunopharmacol* 5, 185–193.
- Bilban, M., Haschemi, A., Wegiel, B., Chin, B. Y., Wagner, O., & Otterbein, L. E. (2008). Heme oxygenase and carbon monoxide initiate homeostatic signaling. *J Mol Med* 86, 267–279.
- Bitanhirwe, B. K., & Cunningham, M. G. (2009). Zinc: The brain's dark horse. *Synapse* 63, 1029–1049.
- Blackstone, E., & Roth, M. B. (2007). Suspended animation-like state protects mice from lethal hypoxia. *Shock* 27, 370–372.
- Blackstone, E., Morrison, M., & Roth, M. B. (2005). H2S induces a suspended animation-like state in mice. *Science* 308, 518.
- Bliss, T. V., & Collingridge, G. L. (1993). A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature* 361, 31–39.

- Boehning, D., & Snyder, S. H. (2003). Novel neural modulators. *Annu Rev Neurosci* 26, 105–131.
- Bohac, D., Burke, W., Cotter, R., Zheng, J., & Potter, J. (2002). A 24-week randomized, double-blind, placebo-controlled study of the efficacy and tolerability of TNFR: Fc (etanercept) in the treatment of dementia of the Alzheimer type. *Neurobiol Aging* 23 (Suppl. 1) S1–S606 abstract 315.
- Boivin, M. J., Bangirana, P., Byarugaba, J., Opoka, R. O., Idro, R., Jurek, A. M., et al. (2007). Cognitive impairment after cerebral malaria in children: A prospective study. *Pediatr* 119, E360–E366.
- Boobis, S., & Hartley, R. E. (1981). Measurement of hypozincemia in mice: A sensitive test for detection of pyrogens. *Clin Sci* 61, 445–449.
- Bossen, C., Ingold, K., Tardivel, A., Bodmer, J. L., Gaide, O., Hertig, S., et al. (2006). Interactions of tumor necrosis factor (TNF) and TNF receptor family members in the mouse and human. *J Biol Chem* 281, 13964–13971.
- Bour, E. S., Ward, L. K., Cornman, G. A., & Isom, H. C. (1996). Tumor necrosis factor- $\alpha$ -induced apoptosis in hepatocytes in long-term culture. *Am J Pathol* 148, 485–495.
- Brennan, F. M., Chantry, D., Jackson, A., Maini, R., & Feldmann, M. (1989). Inhibitory effect of TNF  $\alpha$  antibodies on synovial cell interleukin-1 production in rheumatoid arthritis. *Lancet* 2, 244–247.
- Brewster, D. R., Kwiatkowski, D., & White, N. J. (1990). Neurological sequelae of cerebral malaria in children. *Lancet* 336, 1039–1043.
- Brinton, R. D., & Wang, J. M. (2006). Therapeutic potential of neurogenesis for prevention and recovery from Alzheimer's disease: Allopregnanolone as a proof of concept neurogenic agent. *Curr Alzheimer Res* 3, 185–190.
- Brown, C. M., Wright, E., Colton, C. A., Sullivan, P. M., Laskowitz, D. T., & Vitek, M. P. (2002). Apolipoprotein E isoform mediated regulation of nitric oxide release. *Free Radic Biol Med* 32, 1071–1075.
- Brugg, B., Dubreuil, Y. L., Huber, G., Wollman, E. E., Delhaye Bouchaud, N., & Mariani, J. (1995). Inflammatory processes induce beta-amyloid precursor protein changes in mouse brain. *Proc Natl Acad Sci USA* 92, 3032–3035.
- Brunetti, C. R., Paulose Murphy, M., Singh, R., Qin, J., Barrett, J. W., Tardivel, A., et al. (2003). A secreted high-affinity inhibitor of human TNF from Tanapox virus. *Proc Natl Acad Sci USA* 100, 4831–4836.
- Brustolim, D., Ribeiro dos Santos, R., Kast, R. E., Altschuler, E. L., & Soares, M. B. (2006). A new chapter opens in anti-inflammatory treatments: the antidepressant bupropion lowers production of tumor necrosis factor- $\alpha$  and interferon- $\gamma$  in mice. *Int Immunopharmacol* 6, 903–907.
- Buchhave, P., Zetterberg, H., Blennow, K., Minthon, L., Janciauskiene, S., & Hansson, O. (2009). Soluble TNF receptors are associated with A $\beta$  metabolism and conversion to dementia in subjects with mild cognitive impairment. *Neurobiol Aging* 450, 56–59.
- Budd, A., Alleva, L., Alsharif, M., Koskinen, A., Smythe, V., Mullbacher, A., et al. (2007). Increased survival after gemfibrozil treatment of severe mouse influenza. *Antimicrob Agents Chemother* 51, 2965–2968.
- Bulger, E. M., Garcia, I., & Maier, R. V. (2003). Induction of heme-oxygenase 1 inhibits endothelial cell activation by endotoxin and oxidant stress. *Surgery* 134, 146–152.
- Burgner, D., Usen, S., Rockett, K., Jallow, M., Ackerman, H., Cervino, A., et al. (2003). Nucleotide and haplotypic diversity of the NOS2A promoter region and its relationship to cerebral malaria. *Hum Genet* 112, 379–386.
- Busquets, S., Aranda, X., Ribas-Carbo, M., Azcon-Bieto, J., Lopez-Soriano, F. J., & Argiles, J. M. (2003). Tumour necrosis factor- $\alpha$  uncouples respiration in isolated rat mitochondria. *Cytokine* 22, 1–4.
- Butovsky, O., Bukshpan, S., Kunis, G., Jung, S., & Schwartz, M. (2007). Microglia can be induced by IFN- $\gamma$  or IL-4 to express neural or dendritic-like markers. *Mol Cell Neurosci* 35, 490–500.
- Butovsky, O., Kunis, G., Koronyo Hamauui, M., & Schwartz, M. (2007). Selective ablation of bone marrow-derived dendritic cells increases amyloid plaques in a mouse Alzheimer's disease model. *Eur J Neurosci* 26, 413–416.
- Buxbaum, J. D., Liu, K. N., Luo, Y., Slack, J. L., Stocking, K. L., Peschon, J. J., et al. (1998). Evidence that tumor necrosis factor  $\alpha$  converting enzyme is involved in regulated  $\alpha$ -secretase cleavage of the Alzheimer amyloid protein precursor. *J Biol Chem* 273, 27765–27767.
- Caccamo, A., Majumder, S., Richardson, A., Strong, R., & Oddo, S. (2010). Molecular interplay between mTOR, A $\beta$  and tau: Effects on cognitive impairments. *J Biol Chem* 285, 13107–13120.
- Cacci, E., Ajmone Cat, M. A., Anelli, T., Biagioni, S., & Minghetti, L. (2008). In vitro neuronal and glial differentiation from embryonic or adult neural precursor cells are differentially affected by chronic or acute activation of microglia. *Glia* 56, 4124–4125.
- Calabrese, V., Butterfield, D. A., & Stella, A. M. (2003). Nutritional antioxidants and the heme oxygenase pathway of stress tolerance: novel targets for neuroprotection in Alzheimer's disease. *Ital J Biochem* 52, 177–181.
- Camara, N. O. S., & Soares, M. P. (2005). Heme oxygenase-1 (HO-1), a protective gene that prevents chronic graft dysfunction. *Free Radic Biol Med* 38, 426–435.
- Cameron, B., & Landreth, G. E. (2010). Inflammation, microglia, and Alzheimer's disease. *Neurobiol Dis* 37, 503–509.
- Canellada, A., Cano, E., Sanchez Ruiloba, L., Zafra, F., & Redondo, J. M. (2006). Calcium-dependent expression of TNF- $\alpha$  in neural cells is mediated by the calcineurin/NFAT pathway. *Mol Cell Neurosci* 31, 692–701.
- Cantoni, L., Rossi, C., Rizzardini, M., Gadina, M., & Gheezi, P. (1991). Interleukin-1 and tumor necrosis factor induce hepatic haem oxygenase. *Biochem J*, 891–894.
- Carmen, J., Rothstein, J. D., & Kerr, D. A. (2009). Tumor necrosis factor- $\alpha$  modulates glutamate transport in the CNS and is a critical determinant of outcome from viral encephalomyelitis. *Brain Res* 31, 143–154.
- Carrodegas, J. A., Rodolose, A., Garza, M. V., Sanz Clemente, A., Perez Pe, R., Lacosta, A. M., et al. (2005). The chick embryo appears as a natural model for research in beta-amyloid precursor protein processing. *Neuroscience* 134, 1285–1300.
- Carswell, E. A., Old, L. J., Kassel, R. L., Green, S., Fiore, N., & Williamson, B. (1975). An endotoxin-induced serum factor that causes necrosis of tumors. *Proc Natl Acad Sci USA* 72, 3666–3670.
- Carter, J. A., Mung'ala-Odera, V., Neville, B. G. R., Murira, G., Mturi, N., Musumba, C., et al. (2005). Persistent neurocognitive impairments associated with severe falciparum malaria in Kenyan children. *J Neurol Neurosurg Psychiatry* 76, 476–481.
- Carvalho, C., Correia, S. C., Santos, R. X., Cardoso, S., Moreira, P. I., Clark, T. A., et al. (2009). Role of mitochondrial-mediated signaling pathways in Alzheimer disease and hypoxia. *J Bioenerg Biomembr* 41, 433–440.
- Castellani, R. J., Lee, H. G., Zhu, X., Perry, G., & Smith, M. A. (2008). Alzheimer disease pathology as a host response. *J Neuropathol Exp Neurol* 67, 523–531.
- Castellani, R. J., Zhu, X., Lee, H. G., Smith, M. A., & Perry, G. (2009). Molecular pathogenesis of Alzheimer's disease: reductionist versus expansionist approaches. *Int J Mol Sci* 10, 1386–1406.
- CFAS (2001). Pathological correlates of late-onset dementia in a multicentre, community-based population in England and Wales. Neuro-pathology Group of the Medical Research Council Cognitive Function and Ageing Study (MRC CFAS). *Lancet* 357, 169–175.
- Chang, H. R., Grau, G. E., & Perchere, J. C. (1990). Role of TNF and IL-1 in infections with *Toxoplasma gondii*. *Immunology* 69, 33–37.
- Charles, P., Elliott, M. J., Davis, D., Potter, A., Kalden, J. R., Antoni, C., et al. (1999). Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF- $\alpha$  therapy in rheumatoid arthritis. *J Immunol* 163, 1521–1528.
- Chatenoud, L. (1993). OKT3-induced cytokine-release syndrome – Preventive effect of anti-tumor necrosis factor monoclonal antibody. *Transplant Proc* 25(Suppl. 1), 47–51.
- Chavant, F., Deguil, J., Pain, S., Ingrand, I., Milin, S., Fauconneau, B., et al. (2010). Imipramine, in part through TNF- $\alpha$  inhibition, prevents cognitive decline and A $\beta$  accumulation in a mouse model of Alzheimer's disease. *J Pharmacol Exp Ther* 332, 505–514.
- Chen, Y., Lomnitski, L., Michaelson, D. M., & Shohami, E. (1997). Motor and cognitive deficits in apolipoprotein E-deficient mice after closed head injury. *Neuroscience* 80, 1255–1262.
- Chen, X., Stern, D., & Yan, S. D. (2006). Mitochondrial dysfunction and Alzheimer's disease. *Curr Alzheimer Res* 3, 515–520.
- Clark, I. A. (1987). Cell-mediated immunity in protection and pathology of malaria. *Parasitol Today* 3, 300–305.
- Clark, I. A. (2007). How TNF was recognized to be a key mechanism of disease. *Cytokine Growth Factor Rev* 18, 335–343.
- Clark, I. A., & Alleva, L. M. (2009). Is human malarial coma caused, or merely deepened, by sequestration? *Trends Parasitol* 25, 314–318.
- Clark, I. A., & Chaudhri, G. (1988). Tumour necrosis factor may contribute to the anaemia of malaria by causing dyserythropoiesis and erythrophagocytosis. *Br J Haematol* 70, 99–103.
- Clark, I. A., & Cowden, W. B. (2003). The pathophysiology of falciparum malaria. *Pharmacol Ther* 99, 221–260.
- Clark, I. A., Viellezier, J.-L., Carswell, E. A., & Wood, P. R. (1981). Possible importance of macrophage-derived mediators in acute malaria. *Infect Immun* 32, 1058–1066.
- Clark, I. A., Hunt, N. H., Butcher, G. A., & Cowden, W. B. (1987). Inhibition of murine malaria (*Plasmodium chabaudi*) in vivo by recombinant interferon- $\gamma$  or tumor necrosis factor, and its enhancement by butylated hydroxyanisole. *J Immunol* 139, 3493–3496.
- Clark, I. A., Awburn, M. M., Whitten, R. O., Harper, C. G., Liomba, N. G., Molyneux, M. E., et al. (2003). Tissue distribution of migration inhibitory factor and inducible nitric oxide synthase in falciparum malaria and sepsis in African children. *Malar J* 2, 6.
- Clark, I. A., Alleva, L. E., Mills, A. C., & Cowden, W. B. (2004). Pathogenesis in malaria and clinically similar conditions. *Clin Microbiol Rev* 17, 509–539.
- Clark, I. A., Budd, A. C., Alleva, L. M., & Cowden, W. B. (2006). Human malarial disease: A consequence of inflammatory cytokine release. *Malar J* 5, 85.
- Clark, I. A., Budd, A. C., & Alleva, L. M. (2008). Sickness behaviour pushed too far – The basis of the syndrome seen in severe protozoal, bacterial and viral diseases and post-trauma. *Malar J* 7, 208.
- Clavier, N., Rahimy, C., Falanga, P., Ayivi, B., & Payen, D. (1999). No evidence for cerebral hypoperfusion during cerebral malaria. *Crit Care Med* 27, 628–632.
- Cleary, J. P., Walsh, D. M., Hofmeister, J. J., Shankar, G. M., Kuskowski, M. A., Selkoe, D. J., et al. (2005). Natural oligomers of the amyloid- $\beta$  protein specifically disrupt cognitive function. *Nat Neurosci* 8, 79–84.
- Clipstone, N. A., & Crabtree, G. R. (1992). Identification of calcineurin as a key signalling enzyme in T-lymphocyte activation. *Nature* 357, 695–697.
- Collman, J. P., Ghosh, S., Dey, A., & Decreau, R. A. (2009). Using a functional enzyme model to understand the chemistry behind hydrogen sulfide induced hibernation. *Proc Natl Acad Sci USA* 106, 22090–22095.
- Combs, C. K., Johnson, D. E., Karlo, J. C., Cannady, S. B., & Landreth, G. E. (2000). Inflammatory mechanisms in Alzheimer's disease: Inhibition of beta-amyloid-stimulated proinflammatory responses and neurotoxicity by PPAR $\gamma$  agonists. *J Neurosci* 20, 558–567.
- Corder, E. H., Robertson, K., Lannfelt, L., Bogdanovic, N., Eggertsen, G., Wilkins, J., et al. (1998). HIV-infected subjects with the E4 allele for APOE have excess dementia and peripheral neuropathy. *Nat Med* 4, 1182–1184.
- Craig, D., Hart, D. J., McCool, K., McLroy, S. P., & Passmore, A. P. (2004). Apolipoprotein E4 allele influences aggressive behaviour in Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 75, 1327–1330.
- Cunningham, C., Campion, S., Lunnon, K., Murray, C. L., Woods, J. F., Deacon, R. M., et al. (2009). Systemic inflammation induces acute behavioral and cognitive changes and accelerates neurodegenerative disease. *Biol Psychiatry* 65, 304–332.
- Dammann, O., & Leviton, A. (1997). Maternal intrauterine infection, cytokines, and brain damage in the preterm newborn. *Pediatr Res* 42, 1–8.

- Dantzer, R., O'Connor, J. C., Freund, G. G., Johnson, R. W., & Kelley, K. W. (2008). From inflammation to sickness and depression: When the immune system subjugates the brain. *Nat Rev Neurosci* 9, 46–56.
- Darville, T., Milligan, L. B., & Laffoon, K. K. (1997). Intravenous immunoglobulin inhibits staphylococcal toxin-induced human mononuclear phagocyte tumor necrosis factor alpha production. *Infect Immun* 65, 366–372.
- d'Avila, J. C., Santiago, A. P., Amancio, R. T., Galina, A., Oliveira, M. F., & Bozza, F. A. (2008). Sepsis induces brain mitochondrial dysfunction. *Crit Care Med* 36, 1925–1932.
- de Bont, N., Netea, M. G., Demacker, P. N., Verschueren, I., Kullberg, B. J., van Dijk, K. W., et al. (1999). Apolipoprotein E knock-out mice are highly susceptible to endotoxemia and *Klebsiella pneumoniae* infection. *J Lipid Res* 40, 680–685.
- Dean, B., Tawadros, N., Scarr, E., & Gibbons, A. S. (2010). Regionally-specific changes in levels of tumour necrosis factor in the dorsolateral prefrontal cortex obtained postmortem from subjects with major depressive disorder. *J Affect Disord* 120, 245–248.
- Degre, M., & Bukholm, G. (1990). Effect of tumor necrosis factor-alpha on infection with *Salmonella typhimurium* in a mouse model. *J Biol Regul Homeost Agents* 4, 157–161.
- DeKosky, S. T., & Scheff, S. W. (1990). Synapse loss in frontal cortex biopsies in Alzheimer's disease: Correlation with cognitive severity. *Ann Neurol* 27, 457–464.
- Desruisseaux, M. S., Gulinello, M., Smith, D. N., Lee, S. C., Tsuji, M., Weiss, L. M., et al. (2008). Cognitive dysfunction in mice infected with *Plasmodium berghei* strain ANKA. *J Infect Dis* 197, 1621–1627.
- Diehl, A. M., & Rai, R. (1996). Review – Regulation of liver regeneration by pro-inflammatory cytokines. *J Gastroenterol Hepatol* 11, 466–470.
- Dinamarca, M. C., Cerpa, W., Garrido, J., Hancke, J. L., & Inestrosa, N. C. (2006). Hyperforin prevents beta-amyloid neurotoxicity and spatial memory impairments by disaggregation of Alzheimer's amyloid-beta-deposits. *Mol Psychiatry* 11, 1032–1048.
- Dinarello, C. A. (1999). IL-18: A TH1-inducing, proinflammatory cytokine and new member of the IL-1 family. *J Allergy Clin Immunol* 103, 11–24.
- Dinarello, C. A. (2001). Anti-cytokine therapies in response to systemic infection. *J Invest Dermatol* 6, 244–250.
- Dineley, K. T., Hogan, D., Zhang, W. R., & Tagliatalata, G. (2007). Acute inhibition of calcineurin restores associative learning and memory in Tg2576 APP transgenic mice. *Neurobiol Learn Mem* 88, 217–224.
- Diniz, B. S., Teixeira, A. L., Talib, L. L., Mendonca, V. A., Gattaz, W. F., & Forlenza, O. V. (in press). Increased soluble TNF receptor 2 in antidepressant-free patients with late-life depression. *J Psychiatr Res*. doi:10.1016/j.psychires.2010.02.008.
- D'Mello, C., Le, T., & Swain, M. G. (2009). Cerebral microglia recruit monocytes into the brain in response to tumor necrosis factor-alpha signaling during peripheral organ inflammation. *J Neurosci* 29, 2089–2102.
- Dodel, R. C., Du, Y., Depboylu, C., Hampel, H., Frolich, L., Haag, A., et al. (2004). Intravenous immunoglobulins containing antibodies against beta-amyloid for the treatment of Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 75, 1472–1474.
- Domercq, M., Brambilla, L., Pilati, E., Marchaland, J., Volterra, A., & Bezzi, P. (2006). P2Y1 receptor-evoked glutamate exocytosis from astrocytes – Control by tumor necrosis factor-alpha and prostaglandins. *J Biol Chem* 281, 30684–30696.
- Dom, S., Cinatl, J., & Mrowietz, U. (2008). The impact of treatment with tumour necrosis factor-alpha antagonists on the course of chronic viral infections: A review of the literature. *Br J Dermatol* 159, 1217–1228.
- Dowlati, Y., Herrmann, N., Swardfager, W., Liu, H., Sham, L., Reim, E. K., et al. (2010). A meta-analysis of cytokines in major depression. *Biol Psychiatry* 67, 446–457.
- Dunn, N., Mullen, M., Perry, V. H., & Holmes, C. (2005). Association between dementia and infectious disease: Evidence from a case-control study. *Alzheimer Dis Assoc Disord* 19, 91–94.
- Eason, J. D., Pascual, M., Wee, S., Farrell, M., Phelan, J., Boskovic, S., et al. (1996). Evaluation of recombinant human soluble dimeric tumor necrosis factor receptor for prevention of OKT3-associated acute clinical syndrome. *Transplantation* 61, 224–228.
- Edwards, M. M., & Robinson, S. R. (2006). TNF alpha affects the expression of GFAP and S100B: implications for Alzheimer's disease. *J Neural Transm* 113, 1709–1715.
- Elliott, M. J., Maini, R. N., Feldmann, M., Long Fox, A., Charles, P., Katsikis, P., et al. (1993). Treatment of rheumatoid arthritis with chimeric monoclonal antibodies to tumor necrosis factor alpha. *Arthritis Rheum* 36, 1681–1690.
- Elliott, M. J., Maini, R. N., Feldmann, M., Kalden, J. R., Antoni, C., Smolen, J. S., et al. (1994). Randomised double-blind comparison of chimeric monoclonal antibody to tumour necrosis factor alpha (cA2) versus placebo in rheumatoid arthritis. *Lancet* 344, 1105–1110.
- Ely, E. W., Girard, T. D., Shintani, A. K., Jackson, J. C., Gordon, S. M., Thomason, J. W., et al. (2007). Apolipoprotein E4 polymorphism as a genetic predisposition to delirium in critically ill patients. *Crit Care Med* 35, 112–117.
- Emelyanov, V. V. (2001). Evolutionary relationship of Rickettsiae and mitochondria. *FEBS Lett* 501, 11–18.
- Engelhart, M. J., Geerlings, M. I., Meijer, J., Kiliaan, A., Ruitenber, A., van Swieten, J. C., et al. (2004). Inflammatory proteins in plasma and the risk of dementia: The Rotterdam study. *Arch Neurol* 61, 668–672.
- Escribano, L., Simon, A. M., Perez Mediavilla, A., Salazar Colacho, P., Del Rio, J., & Frechilla, D. (2009). Rosiglitazone reverses memory decline and hippocampal glucocorticoid receptor down-regulation in an Alzheimer's disease mouse model. *Biochem Biophys Res Commun* 379, 406–410.
- Esposito, E., & Cuzzocrea, S. (2009). TNF-alpha as a therapeutic target in inflammatory diseases, ischemia-reperfusion injury and trauma. *Curr Med Chem* 16, 3152–3167.
- Facchini, S., Candusso, M., Martellosi, S., Liubich, E., Manfilii, E., & Ventura, A. (2001). Efficacy of long-term treatment with thalidomide in children and young adults with Crohn disease: Preliminary results. *J Pediatr Gastroenterol Nutr* 32, 178–181.
- Fassbender, K., Walter, S., Kuhl, S., Landmann, R., Ishii, K., Bertsch, T., et al. (2004). The LPS receptor (CD14) links innate immunity with Alzheimer's disease. *FASEB J* 18, 203–205.
- Fekade, D., Knox, K., Hussein, K., Melka, A., Laloo, D. G., Coxon, R. E., et al. (1996). Prevention of Jarisch–Herxheimer reactions by treatment with antibodies against tumor necrosis factor alpha. *N Engl J Med* 335, 311–315.
- Feldmann, M., Brennan, F. M., Chantry, D., Haworth, C., Turner, M., Abney, E., et al. (1990). Cytokine production in the rheumatoid joint: Implications for treatment. *Ann Rheum Dis* 49, 480–486.
- Feldmann, M., & Maini, R. N. (2003). TNF defined as a therapeutic target for rheumatoid arthritis and other autoimmune diseases. *Nat Med* 9, 1245–1250.
- Fernandez, A. M., Fernandez, S., Carrero, P., Garcia Garcia, M., & Torres Aleman, I. (2007). Calcineurin in reactive astrocytes plays a key role in the interplay between proinflammatory and anti-inflammatory signals. *J Neurosci* 27, 8745–8756.
- Fernandez, J. A., Rojo, L., Kuljis, R. O., & Maccioni, R. B. (2008). The damage signals hypothesis of Alzheimer's disease pathogenesis. *J Alzheimers Dis* 14, 329–333.
- Ferrer, I. (2009). Altered mitochondria, energy metabolism, voltage-dependent anion channel, and lipid rafts converge to exhaust neurons in Alzheimer's disease. *J Bioenerg Biomembr* 2, 2.
- Fink, M. (1997). Cytopathic hypoxia in sepsis. *Acta Anaesthesiol Scand* 110, 87–95.
- Fink, M. P. (2000). Cytopathic hypoxia. A concept to explain organ dysfunction in sepsis. *Minerva Anestesiol* 66, 337–342.
- Fink, M. P. (2001). Cytopathic hypoxia. Mitochondrial dysfunction as mechanism contributing to organ dysfunction in sepsis. *Crit Care Clin* 17, 219–237.
- Fink, M. P. (2004). Research: advances in cell biology relevant to critical illness. *Curr Opin Crit Care* 10, 279–291.
- Floyd, R. A., & Krueger, J. M. (1997). Diurnal variation of TNF-alpha in the rat brain. *NeuroReport* 8, 915–918.
- Fontan-Lozano, A., Lopez-Lluch, G., Delgado-Garcia, J. M., Navas, P., & Carrion, A. M. (2008). Molecular bases of caloric restriction regulation of neuronal synaptic plasticity. *Mol Neurobiol* 38, 167–177.
- Foresti, R., Bani Hani, M. G., & Motterlini, R. (2008). Use of carbon monoxide as a therapeutic agent: Promises and challenges. *Intensive Care Med* 34, 649–658.
- Franchi, L., Eigenbrod, T., & Nunez, G. (2009). Cutting edge: TNF-alpha mediates sensitization to ATP and silica via the NLRP3 inflammasome in the absence of microbial stimulation. *J Immunol* 183, 792–796.
- Francis, J., Chu, Y., Johnson, A. K., Weiss, R. M., & Felder, R. B. (2004). Acute myocardial infarction induces hypothalamic cytokine synthesis. *Am J Physiol* 286, H2264–H2271.
- Franz, G., Beer, R., Kampfl, A., Engelhardt, K., Schmutzhard, E., Ulmer, H., et al. (2003). Amyloid beta 1–42 and tau in cerebrospinal fluid after severe traumatic brain injury. *Neurology* 60, 1457–1461.
- Frederickson, R. E., Frederickson, C. J., & Danscher, G. (1990). In situ binding of bouton zinc reversibly disrupts performance on a spatial memory task. *Behav Brain Res* 38, 25–33.
- Fuller, C. (1907). A study of neurofibrils in dementia paralytica, dementia senilis, chronic alcoholism, cerebral lues and microcephalic idiocy. *Am J Insanity* 63, 415–468.
- Furst, D. E., Keystone, E. C., Fleischmann, R., Mease, P., Breedveld, F. C., Smolen, J. S., et al. (2010). Updated consensus statement on biological agents for the treatment of rheumatic diseases, 2009. *Ann Rheum Dis* 69(Suppl 1), i2–i29.
- Gabryel, B., Labuzek, K., Malecki, A., & Herman, Z. S. (2004). Immunophilin ligands decrease release of pro-inflammatory cytokines (IL-1beta, TNF-alpha and IL-2) in rat astrocyte cultures exposed to simulated ischemia in vitro. *Pol J Pharmacol* 56, 129–136.
- Gabuzda, D., Busciglio, J., Chen, L. B., Matsudaira, P., & Yankner, B. A. (1994). Inhibition of energy metabolism alters the processing of amyloid precursor protein and induces a potentially amyloidogenic derivative. *J Biol Chem* 269, 13623–13628.
- Galic, M. A., Riaz, K., Heida, J. G., Mouhate, A., Fournier, N. M., Spencer, S. J., et al. (2008). Postnatal inflammation increases seizure susceptibility in adult rats. *J Neurosci* 28, 6904–6913.
- Gallay, P., Mach, J. P., & Carrel, S. (1991). Characterization and detection of naturally occurring antibodies against IL-1 alpha and IL-1 beta in normal human plasma. *Eur Cytokine Netw* 2, 329–338.
- Ge, Y. W., & Lahiri, D. K. (2002). Regulation of promoter activity of the APP gene by cytokines and growth factors: Implications in Alzheimer's disease. *Ann NY Acad Sci* 973, 463–467.
- Gendron, R. L., Nestel, F. P., Lapp, W. S., & Baines, M. G. (1991). Expression of tumor necrosis factor alpha in the developing nervous system. *Int J Neurosci* 60, 129–136.
- Genton, B., D'Acremont, V., Rare, L., Baea, K., Reeder, J. C., Alpers, M. P., et al. (2008). *Plasmodium vivax* and mixed infections are associated with severe malaria in children: A prospective cohort study from Papua New Guinea. *PLoS Med* 5, e127.
- Ghezzi, P., Di Santo, E., Sacco, S., Foddi, C., Barbaccia, M. L., & Mennini, T. (2000). Neurosteroid levels are increased in vivo after LPS treatment and negatively regulate LPS-induced TNF production. *Eur Cytokine Netw* 11, 464–469.
- Giannini, L., Vannacci, A., Fabrizi, F., Uliva, C., Bani, D., Masini, E., et al. (2005). Protection from cardiac injury by induction of heme oxygenase-1 and nitric oxide synthase in a focal ischaemia-reperfusion model. *Cell Mol Biol* 51, 393–401.
- Gilroy, D. W., Colville Nash, P. R., Willis, J., Paul Clark, M. J., & Willoughby, D. A. (1999). Inducible cyclooxygenase may have anti-inflammatory properties. *Nat Med* 5, 698–701.
- Gisslen, M., Krut, J., Andreasson, U., Blennow, K., Cinque, P., Brew, B. J., et al. (2009). Amyloid and tau cerebrospinal fluid biomarkers in HIV infection. *BMC Neurol* 9, 63.
- Goedert, M. (2004). Tau protein and neurodegeneration. *Semin Cell Dev Biol* 15, 45–49.
- Gohier, B., Ferracci, L., Surguladze, S. A., Lawrence, E., El Hage, W., Kefi, M. Z., et al. (2009). Cognitive inhibition and working memory in unipolar depression. *J Affect Disord* 116, 100–105.

- Goldbaum, L. R., Orellano, T., & Dergal, E. (1976). Mechanism of the toxic action of carbon monoxide. *Ann Clin Lab Sci* 6, 372–376.
- Goldfeld, A. E., Fleming, E. K., Boussiotis, V. A., Theodos, C. M., Titus, R. G., Strominger, J. L., et al. (1992). Transcription of the tumor necrosis factor alpha gene is rapidly induced by anti-immunoglobulin and blocked by cyclosporin A and FK506 in human B cells. *Proc Natl Acad Sci USA* 89, 12198–12201.
- Goldgaber, D., Harris, H. W., Hla, T., Maciag, T., Donnelly, R. J., Jacobsen, J. S., et al. (1989). Interleukin 1 regulates synthesis of amyloid beta-protein precursor mRNA in human endothelial cells. *Proc Natl Acad Sci USA* 86, 7606–76010.
- Gong, Q. H., Wang, Q., Pan, L. L., Liu, X. H., Huang, H., & Zhu, Y. Z. (2010). Hydrogen sulfide attenuates lipopolysaccharide-induced cognitive impairment: A pro-inflammatory pathway in rats. *Pharmacol Biochem Behav* 96, 52–58.
- Gonzalez Gay, M. A., De Matias, J. M., Gonzalez Juanatey, C., Garcia Porrua, C., Sanchez Andrade, A., Martin, J., et al. (2006). Anti-tumor necrosis factor- $\alpha$  blockade improves insulin resistance in patients with rheumatoid arthritis. *Clin Exp Rheumatol* 24, 83–86.
- Goodman, J. C., Robertson, C. S., Grossman, R. G., & Narayan, R. K. (1990). Elevation of tumor necrosis factor in head injury. *J Neuroimmunol* 30, 217.
- Gordon, G. R., Baimoukhametova, D. V., Hewitt, S. A., Rajapaksha, W. R., Fisher, T. E., & Bains, J. S. (2005). Norepinephrine triggers release of glial ATP to increase postsynaptic efficacy. *Nat Neurosci* 8, 1078–1086.
- Gordon, G. R., Iremonger, K. J., Kantevari, S., Ellis Davies, G. C., MacVicar, B. A., & Bains, J. S. (2009). Astrocyte-mediated distributed plasticity at hypothalamic glutamate synapses. *Neuron* 64, 391–403.
- Gorlovoy, P., Larionov, S., Pham, T. T., & Neumann, H. (2009). Accumulation of tau induced in neurites by microglial proinflammatory mediators. *FASEB J* 23, 2502–2513.
- Granger, D. L., Taintor, R. R., Cook, J. L., & Hibbs, J. B. (1980). Injury of neoplastic cells by murine macrophages leads to inhibition of mitochondrial respiration. *J Clin Invest* 65, 357–370.
- Grassi-Oliveira, R., Brietzke, E., Pezzi, J. C., Lopes, R. P., Teixeira, A. L., & Bauer, M. E. (2009). Increased soluble tumor necrosis factor- $\alpha$  receptors in patients with major depressive disorder. *Psychiatry Clin Neurosci* 63, 202–208.
- Grau, G. E., Taylor, T. E., Molyneux, M. E., Wirima, J. J., Vassalli, P., Hommel, M., et al. (1989). Tumor necrosis factor and disease severity in children with falciparum malaria. *N Engl J Med* 320, 1586–1591.
- Graumann, A., & Zawada, E. T. (2010). Case report: acute renal failure after administering intravenous immunoglobulin. *Postgrad Med* 122, 142–147.
- Green, A. J., Harvey, R. J., Thompson, E. J., & Rossor, M. N. (1997). Increased S100beta in the cerebrospinal fluid of patients with frontotemporal dementia. *Neurosci Lett* 235, 5–8.
- Greig, N. H., Giordano, T., Zhu, X., Yu, Q. S., Perry, T. A., Holloway, H. W., et al. (2004). Thalidomide-based TNF- $\alpha$  inhibitors for neurodegenerative diseases. *Acta Neurobiol Exp* 64, 1–9.
- Grimm, M., Zimpfer, D., Czerny, M., Kilo, J., Kasimir, M. T., Kramer, L., et al. (2003). Neurocognitive deficit following mitral valve surgery. *Eur J Cardiothorac Surg* 23, 265–271.
- Grunnet, L. G., Aikin, R., Tonnesen, M. F., Paraskevas, S., Blaabjerg, L., Storing, J., et al. (2009). Pro-inflammatory cytokines activate the intrinsic apoptotic pathway in  $\beta$ -cells. *Diabetes* 58, 1807–1815.
- Gullestad, L., Aass, H., Fjeld, J. G., Wikeby, L., Andreassen, A. K., Ihlen, H., et al. (2001). Immunomodulating therapy with intravenous immunoglobulin in patients with chronic heart failure. *Circulation* 103, 220–225.
- Gupta, M., Noel, G. J., Schaefer, M., Friedman, D., Bussell, J., & Johann Liang, R. (2001). Cytokine modulation with immune gamma-globulin in peripheral blood of normal children and its implications in Kawasaki disease treatment. *J Clin Immunol* 21, 193–199.
- Guyant-Marechal, L., Rovelet-Lecrux, A., Goumidi, L., Cousin, E., Hannequin, D., Raux, G., et al. (2007). Variations in the APP gene promoter region and risk of Alzheimer disease. *Neurology* 68, 684–687.
- Haden, D. W., Suliman, H. B., Carraway, M. S., Welty Wolf, K. E., Ali, A. S., Shitara, H., et al. (2007). Mitochondrial biogenesis restores oxidative metabolism during *Staphylococcus aureus* sepsis. *Am J Respir Crit Care Med* 176, 768–777.
- Haerter, G., Manfras, B. J., de Jong-Hesse, Y., Wilts, H., Mertens, T., Kern, P., et al. (2004). Cytomegalovirus reinitiation in a patient treated with anti-tumor necrosis factor alpha antibody therapy for rheumatoid arthritis. *Clin Infect Dis* 39, E88–E94.
- Halle, A., Hornung, V., Petzold, G. C., Stewart, C. R., Monks, B. G., Reinheckel, T., et al. (2008). The NALP3 inflammasome is involved in the innate immune response to amyloid- $\beta$ . *Nat Immunol* 9, 857–865.
- Halperin, J. J. (2010). A tale of two spirochetes: Lyme disease and syphilis. *Neurol Clin* 28, 277–291.
- Hao, H. N., Zhao, J., Lotoczky, G., Grever, W. E., & Lyman, W. D. (2001). Induction of human beta-defensin-2 expression in human astrocytes by lipopolysaccharide and cytokines. *J Neurochem* 77, 1027–1035.
- Hardardottir, I., Kunitake, S. T., Moser, A. H., Doerfler, W. T., Rapp, J. H., Grunfeld, C., et al. (1994). Endotoxin and cytokines increase hepatic messenger RNA levels and serum concentrations of apolipoprotein J (clusterin) in Syrian hamsters. *J Clin Invest* 94, 1304–1309.
- Hardy, J. (1994). Alzheimer's disease. Clinical molecular genetics. *Clin Geriatr Med* 10, 239–247.
- Harmsen, M. M., & De Haard, H. J. (2007). Properties, production, and applications of camelid single-domain antibody fragments. *Appl Microbiol Biotechnol* 77, 13–22.
- Harrison, N. L., & Gibbons, S. J. (1994). Zn<sup>2+</sup> as an endogenous modulator of ligand- and voltage-gated ion channels. *Neuropharmacology* 33, 935–952.
- Hartig, W., Stieler, J., Boerema, A. S., Wolf, J., Schmidt, U., Weissfuss, J., et al. (2007). Hibernation model of tau phosphorylation in hamsters: selective vulnerability of cholinergic basal forebrain neurons – Implications for Alzheimer's disease. *Eur J Neurosci* 25, 69–80.
- Hashimoto, Y. (1998). Novel biological response modifiers derived from thalidomide. *Curr Med Chem* 5, 163–178.
- Haughey, N. J., Nath, A., Chan, S. L., Borchard, A. C., Rao, M. S., & Mattson, M. P. (2002). Disruption of neurogenesis by amyloid beta-peptide, and perturbed neural progenitor cell homeostasis, in models of Alzheimer's disease. *J Neurochem* 83, 1509–1524.
- Hauptmann, S., Scherping, I., Drose, S., Brandt, U., Schulz, K. L., Jendrach, M., et al. (2009). Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol Aging* 30, 1574–1586.
- Hayakata, T., Shiozaki, T., Tasaki, O., Ikegawa, H., Inoue, Y., Toshiyuki, F., et al. (2004). Changes in CSF S100B and cytokine concentrations in early-phase severe traumatic brain injury. *Shock* 22, 102–107.
- Haydon, P. G. (2001). GLIA: Listening and talking to the synapse. *Nat Rev Neurosci* 2, 185–193.
- Haydon, P. G., & Carmignoto, G. (2006). Astrocyte control of synaptic transmission and neurovascular coupling. *Physiol Rev* 86, 1009–2031.
- He, J., Evans, C. O., Hoffman, S. W., Oyesiku, N. M., & Stein, D. G. (2004). Progesterone and allopregnanolone reduce inflammatory cytokines after traumatic brain injury. *Exp Neurol* 189, 404–412.
- He, P., Zhong, Z., Lindholm, K., Berning, L., Lee, W., Lemere, C., et al. (2007). Deletion of tumor necrosis factor death receptor inhibits amyloid beta generation and prevents learning and memory deficits in Alzheimer's mice. *J Cell Biol* 178, 829–841.
- Heldmann, U., Thored, P., Claassen, J. H., Arvidsson, A., Kokaia, Z., & Lindvall, O. (2005). TNF- $\alpha$  antibody infusion impairs survival of stroke-generated neuroblasts in adult rat brain. *Exp Neurol* 196, 204–208.
- Heneka, M. T., Klockgether, T., & Feinstein, D. L. (2000). Peroxisome proliferator-activated receptor- $\gamma$  ligands reduce neuronal inducible nitric oxide synthase expression and cell death in vivo. *J Neurosci* 20, 6862–6867.
- Hernandez Caselles, T., & Stutman, O. (1993). Immune functions of tumor necrosis factor. I. Tumor necrosis factor induces apoptosis of mouse thymocytes and can also stimulate or inhibit IL-6-induced proliferation depending on the concentration of mitogenic costimulation. *J Immunol* 151, 3999–4012.
- Herrlinger, K. R., Barthel, D. N., Schmidt, K. J., Buning, J., Barthel, C. S., Wehkamp, J., et al. (2010). Infliximab as rescue medication for patients with severe ulcerative/indeterminate colitis refractory to tacrolimus. *Aliment Pharmacol Ther* 31, 1036–1041.
- Hesse, G. W. (1979). Chronic zinc deficiency alters neuronal function of hippocampal mossy fibers. *Science* 205, 1005–1007.
- Hestad, K. A., Tonseth, S., Stoen, C. D., Ueland, T., & Aukrust, P. (2003). Raised plasma levels of tumor necrosis factor alpha in patients with depression: Normalization during electroconvulsive therapy. *J ECT* 19, 183–188.
- Hibbs, J. B., Jr., Vavrin, Z., & Taintor, R. R. (1987). L-arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. *J Immunol* 138, 550–565.
- Hibbs, J. B., Taintor, R. R., Vavrin, Z., & Rachlin, E. M. (1988). Nitric oxide: A cytotoxic activated macrophage effector molecule. *Biochem Biophys Res Commun* 157, 87–94.
- Hildebrand, F., van Griensven, M., Giannoudis, P., Luerig, A., Harwood, P., Harms, O., et al. (2005). Effects of hypothermia and re-warming on the inflammatory response in a murine multiple hit model of trauma. *Cytokine* 31, 382–393.
- Himmerich, H., Fulda, S., Linseisen, J., Seiler, H., Wolfram, G., Himmerich, S., et al. (2008). Depression, comorbidities and the TNF- $\alpha$  system. *Eur Psychiatry* 23, 421–429.
- Ho, M., & White, N. J. (1999). Molecular mechanisms of cytoadherence in malaria. *Am J Physiol* 45, C1231–C1242.
- Hobbs, M. R., Udhayakumar, V., Levesque, M. C., Booth, J., Roberts, J. M., Tkachuk, A. N., et al. (2002). A new NOS2 promoter polymorphism associated with increased nitric oxide production and protection from severe malaria in Tanzanian and Kenyan children. *Lancet* 360, 1468–1475.
- Hoeffler, C. A., Tang, W., Wong, H., Santillan, A., Patterson, R. J., Martinez, L. A., et al. (2008). Removal of FKBP12 enhances mTOR-Raptor interactions, LTP, memory, and perseverative/repetitive behavior. *Neuron* 60, 832–845.
- Hoffmann, O., Mahrhofer, C., Rueter, N., Freyer, D., Bert, B., Fink, H., et al. (2007). Pneumococcal cell wall-induced meningitis impairs adult hippocampal neurogenesis. *Infect Immun* 75, 4289–4297.
- Holler, E., Kolb, H. J., Mittermuller, J., Kaul, M., Ledderose, G., Duell, T., et al. (1995). Modulation of acute graft-versus-host-disease after allogeneic bone marrow transplantation by tumor necrosis factor alpha (TNF alpha) release in the course of pretransplant conditioning: role of conditioning regimens and prophylactic application of a monoclonal antibody neutralizing human TNF alpha (MAK 195F). *Blood* 86, 890–899.
- Holmes, C., Boche, D., Wilkinson, D., Yadegarfar, G., Hopkins, V., Bayer, A., et al. (2008). Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet* 372, 216–223.
- Holmes, C., Cunningham, C., Zotova, E., Woolford, J., Dean, C., Kerr, S., et al. (2009). Systemic inflammation and disease progression in Alzheimer disease. *Neurology* 73, 768–774.
- Hopkins, R. O., & Jackson, J. C. (2006). Long-term neurocognitive function after critical illness. *Chest* 130, 869–878.
- Hu, N. W., Klyubin, I., Anwyl, R., & Rowan, M. J. (2009). GluN2B subunit-containing NMDA receptor antagonists prevent A $\beta$ (1–42)-mediated synaptic plasticity disruption in vivo. *Proc Natl Acad Sci USA* 106, 20504–20509.
- Hu, L. F., Lu, M., Tiong, C. X., Dawe, G. S., Hu, G., & Bian, J. S. (2010). Neuroprotective effects of hydrogen sulfide on Parkinson's disease rat models. *Aging Cell* 9, 135–146.
- Hunt, N. H., & Stocker, R. (2007). Heme moves to center stage in cerebral malaria. *Nat Med* 13, 667–669.

- Ichiyama, T., Isumi, H., Ozawa, H., Matsubara, T., Morishima, T., & Furukawa, S. (2003). Cerebrospinal fluid and serum levels of cytokines and soluble tumor necrosis factor receptor in influenza virus-associated encephalopathy. *Scand J Infect Dis* 35, 59–61.
- Idro, R., Jenkins, N. E., & Newton, C. R. (2005). Pathogenesis, clinical features, and neurological outcome of cerebral malaria. *Lancet Neurol* 4, 827–840.
- Idro, R., Kakooza Mwesige, A., Balyejussa, S., Mirembe, G., Mugasha, C., Tugumisirize, J., et al. (2010). Severe neurological sequelae and behaviour problems after cerebral malaria in Ugandan children. *BMC Res Notes* 3, 104.
- Imayoshi, I., Sakamoto, M., Ohtsuka, T., Takao, K., Miyakawa, T., Yamaguchi, M., et al. (2008). Roles of continuous neurogenesis in the structural and functional integrity of the adult forebrain. *Nat Neurosci* 11, 1153–1161.
- Ingram, D. K., Weindruch, R., Spangler, E. L., Freeman, J. R., & Walford, R. L. (1987). Dietary restriction benefits learning and motor performance of aged mice. *J Gerontol* 42, 78–81.
- Isaacs, A., & Lindenmann, J. (1957). Virus interference. I. The interferon. *Proc R Soc Lond B Biol Sci* 147, 258–267.
- Ishigami, M., Hiraki, K., Umemura, K., Ogasawara, Y., Ishii, K., & Kimura, H. (2009). A source of hydrogen sulfide and a mechanism of its release in the brain. *Antioxid Redox Signal* 11, 205–214.
- Jacobsen, S. E. W., Ruscetti, F. W., Dubois, C. M., & Keller, J. R. (1992). Tumor necrosis factor- $\alpha$  directly and indirectly regulates hematopoietic progenitor cell proliferation – Role of colony-stimulating factor receptor modulation. *J Exp Med* 175, 1759–1772.
- Jacobsohn, D. A., & Vogelsang, G. B. (2002). Novel pharmacotherapeutic approaches to prevention and treatment of GVHD. *Drugs* 62, 879–889.
- Jana, M., Palencia, C. A., & Pahan, K. (2008). Fibrillar amyloid-beta peptides activate microglia via TLR2: Implications for Alzheimer's disease. *J Immunol* 181, 7254–7262.
- Janelins, M. C., Mastrangelo, M. A., Park, K. M., Sudol, K. L., Narrow, W. C., Oddo, S., et al. (2008). Chronic neuron-specific tumor necrosis factor expression enhances the local inflammatory environment ultimately leading to neuronal death in 3xTg-AD mice. *Am J Pathol* 173, 1768–1782.
- Janeway, C. A. (1992). The immune system evolved to discriminate infectious nonself from noninfectious self. *Immunol Today* 13, 111–116.
- Jiang, C., Ting, A. T., & Seed, B. (1998). PPAR- $\gamma$  agonists inhibit production of monocyte inflammatory cytokines. *Nature* 391, 82–86.
- Jiang, W., Desjardins, P., & Butterworth, R. F. (2009). Cerebral inflammation contributes to encephalopathy and brain edema in acute liver failure: protective effect of minocycline. *J Neurochem* 109, 485–493.
- John, C. C., Bangirana, P., Byarugaba, J., Opoka, R. O., Idro, R., Jurek, A. M., et al. (2008). Cerebral malaria in children is associated with long-term cognitive impairment. *Pediatr* 122, 92–99.
- John, C. C., Panoskaltis Mortari, A., Opoka, R. O., Park, G. S., Orchard, P. J., Jurek, A. M., et al. (2008). Cerebrospinal fluid cytokine levels and cognitive impairment in cerebral malaria. *Am J Trop Med Hyg* 78, 198–205.
- Kahan, B. D., Chang, J. Y., & Sehgal, N. E. (1991). Preclinical evaluation of a new potent immunosuppressive agent, rapamycin. *Transplantation* 52, 185–191.
- Karunaweera, N. D., Grau, G. E., Gamage, P., Carter, R., & Mendis, K. N. (1992). Dynamics of fever and serum levels of tumor necrosis factor are closely associated during clinical paroxysms in *Plasmodium vivax* malaria. *Proc Natl Acad Sci USA* 89, 3200–3203.
- Katakowski, M., Chen, J., Zhang, Z. G., Santra, M., Wang, Y., & Chopp, M. (2007). Stroke-induced subventricular zone proliferation is promoted by tumor necrosis factor- $\alpha$ -converting enzyme protease activity. *J Cereb Blood Flow Metab* 27, 669–678.
- Kawano, K., Kim, Y. I., Goto, S., Kai, T., Shimada, T., Kamada, N., et al. (1992). FK 506 ameliorates normothermic liver ischemia in rats by suppressing production of tumor necrosis factor. *Transpl Int* 5, S665–S669.
- Kawano, K., Kim, Y. I., Kai, T., Ishii, T., Tatsuma, T., Morimoto, A., et al. (1994). Evidence that FK506 alleviates ischemia/reperfusion injury to the rat liver: in vivo demonstration for suppression of TNF- $\alpha$  production in response to endotoxemia. *Eur Surg Res* 26, 108–115.
- Keane, J., Gershon, S., Wise, R. P., Mirabile Levens, E., Kasznica, J., Schwieterman, W. D., et al. (2001). Tuberculosis associated with infliximab, a tumor necrosis factor alpha-neutralizing agent. *N Engl J Med* 345, 1098–1104.
- Keilp, J. G., Gorlyn, M., Oquendo, M. A., Brodsky, B., Ellis, S. P., Stanley, B., et al. (2006). Aggressiveness, not impulsiveness or hostility, distinguishes suicide attempters with major depression. *Psychol Med* 36, 1779–1788.
- Kerfoot, S. M., D'Mello, C., Nguyen, H., Ajuabor, M. N., Kubes, P., Le, T., et al. (2006). TNF- $\alpha$ -secreting monocytes are recruited into the brain of cholestatic mice. *Hepatology* 43, 154–162.
- Khairova, R. A., Machado Vieira, R., Du, J., & Manji, H. K. (2009). A potential role for pro-inflammatory cytokines in regulating synaptic plasticity in major depressive disorder. *Int J Neuropsychopharmacol* 12, 561–578.
- Khan, A. U., Delude, R. L., Han, Y. Y., Sappington, P. L., Han, X., Carcillo, J. A., et al. (2002). Liposomal NAD(+) prevents diminished O-2 consumption by immunostimulated Caco-2 cells. *Am J Physiol* 282, L1082–L1091.
- Kihara, M., Carter, J. A., & Newton, C. R. (2006). The effect of *Plasmodium falciparum* on cognition: A systematic review. *Trp Med Int Health* 11, 386–397.
- Kihara, M., Carter, J. A., Holding, P. A., Vargha Khadem, F., Scott, R. C., Idro, R., et al. (2009). Impaired everyday memory associated with encephalopathy of severe malaria: The role of seizures and hippocampal damage. *Malar J* 8, 273.
- Kikuchi, H., Aramaki, K., & Hirohata, S. (2008). Effect of infliximab in progressive Neuro-Behcet's syndrome. *J Neurol Sci* 272, 99–105.
- Kilbourn, R. G., Jubran, A., Gross, S. S., Griffith, O. W., Levi, R., Adams, J., et al. (1990). Reversal of endotoxin-mediated shock by N<sup>G</sup>-methyl-L-arginine, an inhibitor of nitric oxide synthesis. *Biochem Biophys Res Commun* 172, 1132–1138.
- Kim, Y. K., Na, K. S., Shin, K. H., Jung, H. Y., Choi, S. H., & Kim, J. B. (2007). Cytokine imbalance in the pathophysiology of major depressive disorder. *Prog Neuropsychopharmacol Biol Psychiatry* 31, 1044–1053.
- Kishimoto, C., Shioji, K., Kinoshita, M., Iwase, T., Tamaki, S., Fujii, M., et al. (2003). Treatment of acute inflammatory cardiomyopathy with intravenous immunoglobulin ameliorates left ventricular function associated with suppression of inflammatory cytokines and decreased oxidative stress. *Int J Cardiol* 91, 173–178.
- Kling, M. C., Larian, A. A., Scordi Bello, I., Emer, J., & Lebwohl, M. G. (2010). Fatal influenza A(H1N1) respiratory tract infection in a patient having psoriasis treated with infliximab. *Arch Dermatol* 146, 651–654.
- Knobloch, S. M., & Faden, A. I. (1998). Interleukin-10 improves outcome and alters proinflammatory cytokine expression after experimental traumatic brain injury. *Exp Neurol* 153, 143–151.
- Kochar, D. K., Das, A., Kochar, S. K., Saxena, V., Sirohi, P., Garg, S., et al. (2009). Severe *Plasmodium vivax* malaria: A report on serial cases from Bikaner in northwestern India. *Am J Trop Med Hyg* 80, 194–198.
- Koistinaho, M., & Koistinaho, J. (2005). Interactions between Alzheimer's disease and cerebral ischemia—Focus on inflammation. *Brain Res Brain Res Rev* 48, 240–250.
- Korczyn, A. D., & Halperin, I. (2009). Depression and dementia. *J Neurol Sci* 283, 139–142.
- Koronyo-Hamaoui, M., Ko, M. K., Koronyo, Y., Azoulay, D., Seksenyan, A., Kunis, G., et al. (2009). Attenuation of AD-like neuropathology by harnessing peripheral immune cells: local elevation of IL-10 and MMP-9. *J Neurochem* 111, 1409–1424.
- Krakauer, T., Buckley, M., Issaq, H. J., & Fox, S. D. (2010). Rapamycin protects mice from staphylococcal enterotoxin B-induced toxic shock and blocks cytokine release in vitro and in vivo. *Antimicrob Agents Chemother* 54, 1125–1131.
- Kraus, M. R., Schafer, A., Faller, H., Csef, H., & Scheurlen, M. (2003). Psychiatric symptoms in patients with chronic hepatitis C receiving interferon alfa-2b therapy. *J Clin Psychiatry* 64, 708–714.
- Kriegstein, A., & Alvarez Buylla, A. (2009). The glial nature of embryonic and adult neural stem cells. *Annu Rev Neurosci* 32, 149–184.
- Kristensson, K., Nygard, M., Bertini, G., & Bentivoglio, M. (in press). African trypanosome infections of the nervous system: Parasite entry and effects on sleep and synaptic functions. *Prog Neurobiol*.
- Kruman, I. I., Nath, A., Maragos, W. F., Chan, S. L., Jones, M., Rangnekar, V. M., et al. (1999). Evidence that Par-4 participates in the pathogenesis of HIV encephalitis. *Am J Pathol* 155, 39–46.
- Kubota, K., Inoue, K., Hashimoto, R., Kumamoto, N., Kosuga, A., Tatsumi, M., et al. (2009). Tumor necrosis factor receptor-associated protein 1 regulates cell adhesion and synaptic morphology via modulation of N-cadherin expression. *J Neurochem* 110, 496–508.
- Kuchibhotla, K. V., Lattarulo, C. R., Hyman, B. T., & Bacskai, B. J. (2009). Synchronous hyperactivity and intercellular calcium waves in astrocytes in Alzheimer mice. *Science* 323, 1211–1215.
- Kunze, K. (2002). Metabolic encephalopathies. *J Neurol* 249, 1150–1159.
- Kuroda, T., Wada, Y., Kobayashi, D., Murakami, S., Sakai, T., Hirose, S., et al. (2009). Effective anti-TNF- $\alpha$  therapy can induce rapid resolution and sustained decrease of gastroduodenal mucosal amyloid deposits in reactive amyloidosis associated with rheumatoid arthritis. *J Rheumatol* 36, 2409–2415.
- Kwiatkowski, D., Hill, A. V. S., Sambou, I., Twumasi, P., Castracane, J., Manogue, K. R., et al. (1990). TNF concentration in fatal cerebral, non-fatal cerebral, and uncomplicated *Plasmodium falciparum* malaria. *Lancet* 336, 1201–1204.
- Kwiatkowski, D., Molyneux, M. E., Stephens, S., Curtis, N., Klein, N., Pointaire, P., et al. (1993). Anti-TNF therapy inhibits fever in cerebral malaria. *Q J Med* 86, 91–98.
- Lam, N. Y., Rainer, T. H., Chiu, R. W., Joynt, G. M., & Lo, Y. M. (2004). Plasma mitochondrial DNA concentrations after trauma. *Clin Chem* 50, 213–216.
- Lancel, S., Hassoun, S. M., Favory, R., Decoster, B., Motterlini, R., & Neviere, R. (2009). Carbon monoxide rescues mice from lethal sepsis by supporting mitochondrial energetic metabolism and activating mitochondrial biogenesis. *J Pharmacol Exp Ther* 329, 641–648.
- Lang, C. H., Dobrescu, C., & Bagby, G. J. (1992). Tumor necrosis factor impairs insulin action on peripheral glucose disposal and hepatic glucose output. *Endocrinology* 130, 43–52.
- Lannfelt, L., Blennow, K., Zetterberg, H., Batsman, S., Ames, D., Harrison, J., et al. (2008). Safety, efficacy, and biomarker findings of PBT2 in targeting Abeta as a modifying therapy for Alzheimer's disease: A phase Ia, double-blind, randomised, placebo-controlled trial. *Lancet Neurol* 7, 779–786.
- Laurin, D., David Curb, J., Masaki, K. H., White, L. R., & Launer, L. J. (2009). Midlife C-reactive protein and risk of cognitive decline: A 31-year follow-up. *Neurobiol Aging* 30, 1724–1727.
- Laws, S. M., Pernecky, R., Wagenpfeil, S., Muller, U., Forstl, H., Martins, R. N., et al. (2005). TNF polymorphisms in Alzheimer disease and functional implications on CSF beta-amyloid levels. *Hum Mutat* 26, 29–35.
- Lee, J., Seroogy, K. B., & Mattson, M. P. (2002). Dietary restriction enhances neurotrophin expression and neurogenesis in the hippocampus of adult mice. *J Neurochem* 80, 539–547.
- Lee, T. S., Tsai, H. L., & Chau, L. Y. (2003). Induction of heme oxygenase-1 expression in murine macrophages is essential for the anti-inflammatory effect of low dose 15-deoxy-Delta(12, 14)-prostaglandin J(2). *J Biol Chem* 278, 19325–19330.
- Lee, H. G., Zhu, X., Castellani, R. J., Nunomura, A., Perry, G., & Smith, M. A. (2007). Amyloid-beta in Alzheimer disease: The null versus the alternate hypotheses. *J Pharmacol Exp Ther* 321, 823–829.
- Lee, M., Schwab, C., Yu, S., McGeer, E., & McGeer, P. L. (2009). Astrocytes produce the antiinflammatory and neuroprotective agent hydrogen sulfide. *Neurobiol Aging* 30, 1523–1534.
- Lee, M., Sparatore, A., Del Soldato, P., McGeer, E., & McGeer, P. L. (2010). Hydrogen sulfide-releasing NSAIDs attenuate neuroinflammation induced by microglial and astrocytic activation. *Glia* 58, 103–113.

- Leeper-Woodford, S. K., Carey, P. D., Byrne, K., Jenkins, J. K., Fisher, B. J., Blocher, C., et al. (1991). Tumor necrosis factor. Alpha and beta subtypes appear in circulation during onset of sepsis-induced lung injury. *Am Rev Respir Dis* 143, 1076–1082.
- Legnani, P., & Kornbluth, A. (2004). Newer therapies for inflammatory bowel disease. *Curr Treat Options Gastroenterol* 7, 161–167.
- Lewallen, S., Bakker, H., Taylor, T. E., Wills, B. A., Courtright, P., & Molyneux, M. E. (1996). Retinal findings predictive of outcome in cerebral malaria. *Trans R Soc Trop Med Hyg* 90, 144–146.
- Lewallen, S., Harding, S. P., Ajewole, J., Schulenburg, W. E., Molyneux, M. E., Marsh, K., et al. (1999). A review of the spectrum of clinical ocular fundus findings in *P. falciparum* malaria in African children with a proposed classification and grading system. *Trans R Soc Trop Med Hyg* 93, 619–622.
- Li, L., Bhatia, M., Zhu, Y. Z., Zhu, Y. C., Rammath, R. D., Wang, Z. J., et al. (2005). Hydrogen sulfide is a novel mediator of lipopolysaccharide-induced inflammation in the mouse. *FASEB J* 19, 1196–1198.
- Li, L., Salto Tellez, M., Tan, C. H., Whiteman, M., & Moore, P. K. (2009). GYY4137, a novel hydrogen sulfide-releasing molecule, protects against endotoxin shock in the rat. *Free Radic Biol Med* 47, 103–113.
- Liao, Y. F., Wang, B. J., Cheng, H. T., Kuo, L. H., & Wolfe, M. S. (2004). Tumor necrosis factor- $\alpha$ , interleukin-1 $\beta$ , and interferon- $\gamma$  stimulate gamma-secretase-mediated cleavage of amyloid precursor protein through a JNK-dependent MAPK pathway. *J Biol Chem* 279, 49523–49532.
- Liesch, A., Suri Payer, E., Veltkamp, C., Doerr, H., Sommer, C., Rivest, S., et al. (2009). Regulatory T cells are key cerebroprotective immunomodulators in acute experimental stroke. *Nat Med* 15, 192–199.
- Lin, A., Song, C., Kenis, G., Bosmans, E., De Jongh, R., Scharpe, S., et al. (2000). The in vitro immunosuppressive effects of moclobemide in healthy volunteers. *J Affect Disord* 58, 69–74.
- Lindqvist, D., Janelidze, S., Hagell, P., Erhardt, S., Samuelsson, M., Minthon, L., et al. (2009). Interleukin-6 is elevated in the cerebrospinal fluid of suicide attempters and related to symptom severity. *Biol Psychiatry* 66, 287–292.
- Liu, Y. R., Zhu, B., Luo, L. Q., Li, P., Paty, D. W., & Cyander, M. S. (2001). Heme oxygenase-1 plays an important protective role in experimental autoimmune encephalomyelitis. *NeuroReport* 12, 1841–1845.
- Löffler, J., & Huber, G. (1992). Beta-amyloid precursor protein isoforms in various rat brain regions and during brain development. *J Neurochem* 59, 1316–1324.
- Looaeresuwana, S., Sjöstrom, L., Krudsood, S., Wilairatana, P., Porter, R. S., Hills, F., et al. (1999). Polyclonal anti-tumor necrosis factor- $\alpha$  Fab used as an ancillary treatment for severe malaria. *Am J Trop Med Hyg* 61, 26–33.
- Lorne, E., Zhao, X., Zmijewski, J. W., Liu, G., Park, Y. J., Tsuruta, Y., et al. (2009). Participation of mammalian target of rapamycin complex 1 in Toll-like receptor 2- and 4-induced neutrophil activation and acute lung injury. *Am J Respir Cell Mol Biol* 41, 237–245.
- Lotrich, F. E., Ferrell, R. E., Rabinovitz, M., & Pollock, B. G. (2010). Labile anger during interferon alpha treatment is associated with a polymorphism in tumor necrosis factor alpha. *Clin Neuropharmacol* 33, 191–197.
- Lovestone, S., & Reynolds, C. H. (1997). The phosphorylation of tau: A critical stage in neurodevelopment and neurodegenerative processes. *Neuroscience* 78, 309–324.
- Lynch, J. R., Morgan, D., Mance, J., Matthew, W. D., & Laskowitz, D. T. (2001). Apolipoprotein E modulates glial activation and the endogenous central nervous system inflammatory response. *J Neuroimmunol* 114, 107–113.
- Maegraith, B. (1948). Pathological Processes in Malaria and Blackwater Fever. In: (Ed.), (pp 367–369) Oxford: Blackwell.
- Maegraith, B. G. (1954). Physiological aspects of protozoan infection. *Ann Rev Microbiol* 8, 273–288.
- Maegraith, B., & Fletcher, A. (1972). The pathogenesis of mammalian malaria. *Adv Parasitol* 10, 49–75.
- Magarinos, A. M., McEwen, B. S., Saboureaux, M., & Pevet, P. (2006). Rapid and reversible changes in intrahippocampal connectivity during the course of hibernation in European hamsters. *Proc Natl Acad Sci USA* 103, 18775–18780.
- Mahley, R. W., Weisgraber, K. H., & Huang, Y. (2006). Apolipoprotein E4: A causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proc Natl Acad Sci USA* 103, 5644–5651.
- Maier, A. G., Rug, M., O'Neill, M. T., Brown, M., Chakravorty, S., Szeszak, T., et al. (2008). Exported proteins required for virulence and rigidity of *Plasmodium falciparum*-infected human erythrocytes. *Cell* 134, 48–61.
- Mansuy, I. M. (2003). Calcineurin in memory and bidirectional plasticity. *Biochem Biophys Res Commun* 311, 1195–1208.
- Marchiafava, E. & Bignami, A. (1894). Sulle Fibrille Estivo-Autumnali. (pp Rome: E. Loescher.
- Marchiori, P. E., Lino, A. M., Hirata, M. T., Carvalho, N. B., Brotto, M. W., & Scaff, M. (2006). Occurrence of nervous system involvement in SIRS. *J Neurol Sci* 250, 147–152.
- Margulis, L., & Chapman, M. J. (1998). Endosymbioses: Cyclical and permanent in evolution. *Trends Microbiol* 6, 342–345.
- Marhaug, G., & Dowton, S. B. (1994). Serum amyloid A: An acute phase apolipoprotein and precursor of AA amyloid. *Baillieres Clin Rheumatol* 8, 553–573.
- Mariappan, N., Elks, C. M., Fink, B., & Francis, J. (2009). TNF-induced mitochondrial damage: a link between mitochondrial complex I activity and left ventricular dysfunction. *Free Radic Biol Med* 46, 462–470.
- Marsh, K., & Snow, R. W. (1997). Host-parasite interaction and morbidity in malaria endemic areas. *Philos Trans R Soc Lond* 352, 1385–1394.
- Marsland, A. L., Gianaros, P. J., Abramowitz, S. M., Manuck, S. B., & Hariri, A. R. (2008). Interleukin-6 covaries inversely with hippocampal grey matter volume in middle-aged adults. *Biol Psychiatry* 64, 484–490.
- Martinon, F., Burns, K., & Tschopp, J. (2002). The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-1 $\beta$ . *Mol Cell* 10, 417–426.
- Martinon, F., Mayor, A., & Tschopp, J. (2009). The inflammasomes: Guardians of the body. *Annu Rev Immunol* 27, 229–265.
- Marx, C. E., Trost, W. T., Shampine, L. J., Stevens, R. D., Hulette, C. M., Steffens, D. C., et al. (2006). The neurosteroid allopregnanolone is reduced in prefrontal cortex in Alzheimer's disease. *Biol Psychiatry* 60, 1287–1294.
- Mastroianni, C. M., Paoletti, F., Massetti, A. P., Falciano, M., & Vullo, V. (1990). Elevated levels of tumor necrosis factor (TNF) in the cerebrospinal fluid from patients with HIV-associated neurological disorders. *Acta Neurol Napoli* 12, 66–67.
- Mattson, M. P., Barger, S. W., Furukawa, K., Bruce, A. J., Wyss Coray, T., Mark, R. J., et al. (1997). Cellular signaling roles of TGF beta, TNF alpha and beta APP in brain injury responses and Alzheimer's disease. *Brain Res Brain Res Rev* 23, 47–61.
- Mattson, M. P., Duan, W., & Guo, Z. (2003). Meal size and frequency affect neuronal plasticity and vulnerability to disease: Cellular and molecular mechanisms. *J Neurochem* 84, 417–431.
- Matzinger, P. (2002). The danger model: A renewed sense of self. *Science* 296, 301–305.
- Mayeux, R., Ottman, R., Maestre, G., Ngai, C., Tang, M. X., Ginsberg, H., et al. (1995). Synergistic effects of traumatic head injury and apolipoprotein-epsilon 4 in patients with Alzheimer's disease. *Neurology* 45, 555–557.
- Mbele, G. O., Deloulme, J. C., Gentil, B. J., Delphin, C., Ferro, M., Garin, J., et al. (2002). The zinc- and calcium-binding S100B interacts and co-localizes with IQGAP1 during dynamic rearrangement of cell membranes. *J Biol Chem* 277, 49998–50007.
- McAdam, K. P., Li, J., Knowles, J., Foss, N. T., Dinarello, C. A., Rosenwasser, L. J., et al. (1982). The biology of SAA: identification of the inducer, in vitro synthesis, and heterogeneity demonstrated with monoclonal antibodies. *Ann NY Acad Sci* 389, 126–136.
- McAfoose, J., & Baune, B. T. (2009). Evidence for a cytokine model of cognitive function. *Neurosci Behav Rev* 34, 615–619.
- McAlpine, F. E., Lee, J. K., Harms, A. S., Ruhn, K. A., Blurton Jones, M., Hong, J., et al. (2009). Inhibition of soluble TNF signaling in a mouse model of Alzheimer's disease prevents pre-plaque amyloid-associated neuropathology. *Neurobiol Dis* 34, 163–177.
- McCoy, M. K., & Tansey, M. G. (2008). TNF signaling inhibition in the CNS: Implications for normal brain function and neurodegenerative disease. *J Neuroinflammation* 5, 45.
- McGeer, P. L., & McGeer, E. G. (1995). The inflammatory response system of brain – Implications for therapy of Alzheimer and other neurodegenerative diseases. *Brain Res Rev* 21, 195–218.
- McLeod, D., Marshall, J., Kohner, E. M., & Bird, A. C. (1977). The role of axoplasmic transport in the pathogenesis of retinal cotton-wool spots. *Br J Ophthalmol* 61, 177–191.
- McNaull, B. B., Todd, S., McGuinness, B., & Passmore, A. P. (2010). Inflammation and anti-inflammatory strategies for Alzheimer's disease – A mini-review. *Gerontology* 56, 3–14.
- Means, L. W., Higgins, J. L., & Fernandez, T. J. (1993). Mid-life onset of dietary restriction extends life and prolongs cognitive functioning. *Physiol Behav* 54, 503–508.
- Meda, L., Cassatella, M. A., Szendrei, G. I., Otvol, L. Jr., Baron, P., Villalba, M., et al. (1995). Activation of microglial cells by beta-amyloid protein and interferon-gamma. *Nature* 374, 647–650.
- Medana, I. M., Hunt, N. H., & Chaudhri, G. (1997). Tumor necrosis factor- $\alpha$  expression in the brain during fatal murine cerebral malaria: Evidence for production by microglia and astrocytes. *Am J Pathol* 150, 1473–1486.
- Medana, I. M., Day, N. P., Hien, T. T., Mai, N. T., Bethell, D., Phu, N. H., et al. (2002). Axonal injury in cerebral malaria. *Am J Pathol* 160, 655–666.
- Medana, I. M., Lindert, R. B., Wurster, U., Hien, T. T., Day, N. P. J., Phu, N. H., et al. (2005). Cerebrospinal fluid levels of markers of brain parenchymal damage in Vietnamese adults with severe malaria. *Trans R Soc Trop Med Hyg* 99, 610–617.
- Medana, I. M., Idro, R., & Newton, C. R. (2007). Axonal and astrocyte injury markers in the cerebrospinal fluid of Kenyan children with severe malaria. *J Neurol Sci* 258, 93–98.
- Medeiros, R., Prediger, R. D. S., Passos, G. F., Pandolfo, P., Duarte, F. S., Franco, J. L., et al. (2007). Connecting TNF- $\alpha$  signaling pathways to iNOS expression in a mouse model of Alzheimer's disease: Relevance for the behavioral and synaptic deficits induced by amyloid beta protein. *J Neurosci* 27, 5394–5404.
- Medeiros, R., Figueiredo, C. P., Pandolfo, P., Duarte, F. S., Prediger, R. D., Passos, G. F., et al. (2010). The role of TNF- $\alpha$  signaling pathway on COX-2 upregulation and cognitive decline induced by beta-amyloid peptide. *Behav Brain Res* 209, 165–173.
- Mela, L. M., Miller, L. D., Bacalzo, L. V., Jr., Olofsson, K., & White, R. R. t. (1972). Alterations of mitochondrial structure and energy-linked functions in hemorrhagic shock and endotoxemia. *Adv Exp Med Biol* 33, 231–242.
- Merrill, J. E. (1992). Tumor necrosis factor alpha, interleukin 1 and related cytokines in brain development: Normal and pathological. *Dev Neurosci* 14, 1–10.
- Michaelson, M., Taitelman, U., Bshouty, Z., Bar Joseph, G., & Bursztin, S. (1984). Crush syndrome: experience from the Lebanon War, 1982. *Isr J Med Sci* 20, 305–307.
- Migaud, M., Charlesworth, P., Dempster, M., Webster, L. C., Watabe, A. M., Makhinson, M., et al. (1998). Enhanced long-term potentiation and impaired learning in mice with mutant postsynaptic density-95 protein. *Nature* 396, 433–439.
- Millesi, E., Prossinger, H., Dittami, J. P., & Fieder, M. (2001). Hibernation effects on memory in European ground squirrels (*Spermophilus citellus*). *J Biol Rhythms* 16, 264–271.
- Miyata, S., Ohkubo, Y., & Mutoh, S. (2005). A review of the action of tacrolimus (FK506) on experimental models of rheumatoid arthritis. *Inflamm Res* 54, 1–9.
- Moe, G. W., Marin-Garcia, J., Konig, A., Goldenthal, M., Lu, X. R., & Feng, Q. P. (2004). In vivo TNF- $\alpha$  inhibition ameliorates cardiac mitochondrial dysfunction, oxidative stress, and apoptosis in experimental heart failure. *Am J Physiol* 287, H1813–H1820.
- Mogi, M., Harada, M., Riederer, P., Narabayashi, H., Fujita, K., & Nagatsu, T. (1994). Tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) increases both in the brain and in the cerebrospinal fluid from Parkinsonian patients. *Neurosci Lett* 165, 208–210.

- Mohnot, D., Snead, O. C., & Benton, J. W. (1982). Burn encephalopathy in children. *Ann Neurol* 12, 42–47.
- Moir, R. D., Lynch, T., Bush, A. I., Whyte, S., Henry, A., Portbury, S., et al. (1998). Relative increase in Alzheimer's disease of soluble forms of cerebral Abeta amyloid protein precursor containing the Kunitz protease inhibitory domain. *J Biol Chem* 273, 5013–5019.
- Moller, B., Mogensen, S. C., Wendelboe, P., Bendtzen, K., & Petersen, C. M. (1991). Bioactive and inactive forms of tumor necrosis factor- $\alpha$  in spinal fluid from patients with meningitis. *J Infect Dis* 163, 886–889.
- Moller, K., Tofteng, F., Qvist, T., Sahl, C., Sonderkaer, S., & Pedersen, B. K. (2005). Cerebral output of cytokines in patients with pneumococcal meningitis. *Crit Care Med* 33, 979–983.
- Molloy, R. G., O'Riordain, M., Holzheimer, R., Nestor, M., Collins, K., Mannick, J. A., et al. (1993). Mechanism of increased tumor necrosis factor production after thermal injury. Altered sensitivity to PGE2 and immunomodulation with indomethacin. *J Immunol* 151, 2142–2149.
- Moncada, S., Palmer, R. M. J., & Higgs, E. A. (1989). Biosynthesis of nitric oxide from L-arginine: A pathway for the regulation of cell function and communication. *Biochem Pharmacol* 38, 1709–1715.
- Monje, M. L., Toda, H., & Palmer, T. D. (2003). Inflammatory blockade restores adult hippocampal neurogenesis. *Science* 302, 1760–1765.
- Moravan, M., & Segal, B. M. (2009). Treatment of CNS sarcoidosis with infliximab and mycophenolate mofetil. *Neurology* 72, 337–340.
- Moreira, A. L., Sampaio, E. P., Zmuidzinas, A., Frindt, P., Smith, K. A., & Kaplan, G. (1993). Thalidomide exerts its inhibitory action on tumor necrosis factor alpha by enhancing mRNA degradation. *J Exp Med* 177, 1675–1678.
- Moreira, A. L., Wang, J., Sarno, E. N., & Kaplan, G. (1997). Thalidomide protects mice against LPS-induced shock. *Braz J Med Biol Res* 30, 1199–1207.
- Mori, T., Koyama, N., Arendash, G. W., Horikoshi Sakuraba, Y., Tan, J., & Town, T. (2010). Overexpression of human S100B exacerbates cerebral amyloidosis and gliosis in the Tg2576 mouse model of Alzheimer's disease. *Glia* 58, 300–314.
- Mortimer, J. A., van Duijn, C. M., Chandra, V., Fratiglioni, L., Graves, A. B., Heyman, A., et al. (1991). Head trauma as a risk factor for Alzheimer's disease: A collaborative re-analysis of case-control studies. EURODEM Risk Factors Research Group. *Int J Epidemiol* 20, S28–S35.
- Moses, H. L., Branum, E. L., Proper, J. A., & Robinson, R. A. (1981). Transforming growth factor production by chemically transformed cells. *Cancer Res* 41, 2842–2848.
- Muniz-Junqueira, M. I., Silva, F. O., de Paula-Junior, M. R., & Tosta, C. E. (2005). Thalidomide influences the function of macrophages and increases the survival of *Plasmodium berghei*-infected CBA mice. *Acta Trop* 94, 128–138.
- Naesens, M., Lerut, E., Sarwal, M., Van Damme, B., Vanrenterghem, Y., & Kuypers, D. R. (2009). Balancing efficacy and toxicity of kidney transplant immunosuppression. *Transplant Proc* 41, 3393–3395.
- Nakao, A., Kaczorowski, D. J., Zuckerbraun, B. S., Lei, J., Faleo, G., Deguchi, K., et al. (2008). Galantamine and carbon monoxide protect brain microvascular endothelial cells by heme oxygenase-1 induction. *Biochem Biophys Res Commun* 367, 674–679.
- Napoli, C., & Palinski, W. (2005). Neurodegenerative diseases: Insights into pathogenic mechanisms from atherosclerosis. *Neurobiol Aging* 26, 293–302.
- Nayagam, V. M., Wang, X., Tan, Y. C., Poulsen, A., Goh, K. C., Ng, T., et al. (2006). SIRT1 modulating compounds from high-throughput screening as anti-inflammatory and insulin-sensitizing agents. *J Biomol Screen* 11, 959–967.
- Neve, R. L., McPhie, D. L., & Chen, Y. (2000). Alzheimer's disease: a dysfunction of the amyloid precursor protein(1). *Brain Res* 886, 54–66.
- Newman, M. F. (2007). Open heart surgery and cognitive decline. *Cleve Clin J Med* 74, S52–S55.
- Newton, C. R. J. C., Marsh, K., Peshu, N., & Kirkham, F. J. (1996). Perturbations of cerebral hemodynamics in Kenyans with cerebral malaria. *Pediatr Neurol* 15, 41–49.
- Nikolaev, A., McLaughlin, T., O'Leary, D. D., & Tessier-Lavigne, M. (2009). APP binds DR6 to trigger axon pruning and neuron death via distinct caspases. *Nature* 457, 981–989.
- Nishioku, T., Dohgu, S., Takata, F., Eto, T., Ishikawa, N., Kodama, K. B., et al. (2009). Detachment of brain pericytes from the basal lamina is involved in disruption of the blood-brain barrier caused by lipopolysaccharide-induced sepsis in mice. *Cell Mol Neurobiol* 29, 309–316.
- Nisoli, E., & Carruba, M. O. (2006). Nitric oxide and mitochondrial biogenesis. *J Cell Sci* 119, 2855–2862.
- Oakes, N. D., Kennedy, C. J., Jenkins, A. B., Laybutt, D. R., Chisholm, D. J., & Kraegen, E. W. (1994). A new antidiabetic agent, BRL 49653, reduces lipid availability and improves insulin action and glucose regulation in the rat. *Diabetes* 43, 1203–1210.
- Ogoshi, F., Yin, H. Z., Kuppumbatti, Y., Song, B., Amindari, S., & Weiss, J. H. (2005). Tumor necrosis-factor-alpha (TNF-alpha) induces rapid insertion of Ca<sup>2+</sup>-permeable alpha-amino-3-hydroxyl-5-methyl-4-isoxazole-propionate (AMPA)/kainate (Ca-A/K) channels in a subset of hippocampal pyramidal neurons. *Exp Neurol* 193, 384–393.
- Ohta, K., Kikuchi, T., Arai, S., Yoshida, N., Sato, A., & Yoshimura, N. (2003). Protective role of heme oxygenase-1 against endotoxin-induced uveitis in rats. *Exp Eye Res* 77, 665–673.
- Ondrejcek, T., Klyubin, I., Hu, N. W., Barry, A. E., Cullen, W. K., & Rowan, M. J. (2009). Alzheimer's disease amyloid beta protein and synaptic function. *Neuromolecular Med* 4, e8497.
- Ono, K., Hasegawa, K., Naiki, H., & Yamada, M. (2004). Curcumin has potent anti-amyloidogenic effects for Alzheimer's beta-amyloid fibrils in vitro. *J Neurosci Res* 75, 742–750.
- Ophir, G., Amariglio, N., Jacob Hirsch, J., Elkon, R., Rechavi, G., & Michaelson, D. M. (2005). Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. *Neurobiol Dis* 20, 709–718.
- Ortaldo, J. R., Mason, L. H., Mathieson, B. J., Liang, S. M., Flick, D. A., & Herberman, R. B. (1986). Mediation of mouse natural cytotoxic activity by tumour necrosis factor. *Nature* 321, 700–702.
- Ossandon, A., Cassara, E. A., Priori, R., & Valesini, G. (2002). Thalidomide: Focus on its employment in rheumatologic diseases. *Clin Exp Rheumatol* 20, 709–718.
- Ott, M. O., & Bullock, S. L. (2001). A gene trap insertion reveals that amyloid precursor protein expression is a very early event in murine embryogenesis. *Dev Genes Evol* 211, 355–357.
- Otterbein, L. E., Bach, F. H., Alam, J., Soares, M., Tao Lu, H., Wysk, M., et al. (2000). Carbon monoxide has anti-inflammatory effects involving the mitogen-activated protein kinase pathway. *Nat Med* 6, 422–428.
- Pamplona, A., Clark, I. A., & Mota, M. M. (2007). Severe malaria increased the list of heme oxygenase-1-protected diseases. *Future Med* 2, 361–363.
- Pamplona, A., Ferreira, A., Balla, J., Jeney, V., Balla, G., Epiphany, S., et al. (2007). Heme oxygenase-1 and carbon monoxide suppress the pathogenesis of experimental cerebral malaria. *Nat Med* 13, 703–710.
- Park, K. M., Yule, D. I., & Bowers, W. J. (2008). Tumor necrosis factor-alpha potentiates intraneuronal CA2+ signaling via regulation of the inositol 1, 4, 5-trisphosphate receptor. *J Biol Chem* 283, 33069–33079.
- Pascher, A., Klupp, J., Langrehr, J. M., & Neuhaus, P. (2005). Anti-TNF-alpha therapy for acute rejection in intestinal transplantation. *Transplant Proc* 37, 1635–1636.
- Patel, A., Siegel, A., & Zalcman, S. S. (in press). Lack of aggression and anxiolytic-like behavior in TNF receptor (TNF-R1 and TNF-R2) deficient mice. *Brain Behav Immun*. doi:10.1016/j.bbi.2010.05.005.
- Pedersen, W. A., McMillan, P. J., Kulstad, J. J., Leverenz, J. B., Craft, S., & Haynatzki, G. R. (2006). Rosiglitazone attenuates learning and memory deficits in Tg2576 Alzheimer mice. *Exp Neurol* 199, 265–273.
- Perea, G., Navarrete, M., & Araque, A. (2009). Tripartite synapses: Astrocytes process and control synaptic information. *Trends Neurosci* 32, 421–431.
- Perry, V. H., Cunningham, C., & Holmes, C. (2007). Systemic infections and inflammation affect chronic neurodegeneration. *Nat Rev Immunol* 7, 161–167.
- Peselow, E. D., Corwin, J., Fieve, R. R., Rotrosen, J., & Cooper, T. B. (1991). Disappearance of memory deficits in outpatient depressives responding to imipramine. *J Affect Disord* 21, 1731–1783.
- Peskind, E. R., Griffin, W. S., Akama, K. T., Raskind, M. A., & Van Eldik, L. J. (2001). Cerebrospinal fluid S100B is elevated in the earlier stages of Alzheimer's disease. *Neurochem Int* 39, 409–413.
- Petitot, J. M., Lysle, D. T., Garipey, J. L., & Lewis, M. H. (1994). Association of genetic differences in social behavior and cellular immune responsiveness: Effects of social experience. *Brain Behav Immun* 8, 111–122.
- Pickering, M., Cumiskey, D., & O'Connor, J. J. (2005). Actions of TNF-alpha on glutamatergic synaptic transmission in the central nervous system. *Exp Physiol* 90, 663–670.
- Pitossi, F., Delrey, A., Kabiersch, A., & Besedovsky, H. (1997). Induction of cytokine transcripts in the central nervous system and pituitary following peripheral administration of endotoxin to mice. *J Neurosci Res* 48, 287–298.
- Pitsikas, N., & Algeri, S. (1992). Deterioration of spatial and nonspatial reference and working memory in aged rats: Protective effect of life-long calorie restriction. *Neurobiol Aging* 13, 369–373.
- Platzer, C., Meisel, C., Vogt, K., Platzer, M., & Volk, H. D. (1995). Up-regulation of monocytic IL-10 by tumor necrosis factor-alpha and cAMP elevating drugs. *Int Immunol* 7, 517–523.
- Popov, V. I., & Bocharova, L. S. (1992). Hibernation-induced structural changes in synaptic contacts between mossy fibres and hippocampal pyramidal neurons. *Neuroscience* 48, 53–62.
- Porayette, P., Gallego, M. J., Kaltcheva, M. M., Meethal, S. V., & Atwood, C. S. (2007). Amyloid-beta precursor protein expression and modulation in human embryonic stem cells: A novel role for human chorionic gonadotropin. *Biochem Biophys Res Commun* 364, 522–527.
- Preau, M., Marcellin, F., Spire, B., Ravau, I., Dellamonica, P., Blanc, D., et al. (2008). Impaired anger control as an underappreciated side effect of treatments for chronic HCV infection in HIV-HCV coinfecting patients. *J Clin Gastroenterol* 42, 92–96.
- Qiao, X., Cummins, D. J., & Paul, S. M. (2001). Neuroinflammation-induced acceleration of amyloid deposition in the APPV717F transgenic mouse. *Eur J Neurosci* 14, 474–482.
- Qin, L. Y., Wu, X. F., Block, M. L., Liu, Y. X., Bresse, G. R., Hong, J. S., et al. (2007). Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia* 55, 453–462.
- Quan, N., Mhlanga, J., Whiteside, M. B., McCop, A. N., Kristensson, K., & Herkenham, M. (1999). Chronic overexpression of proinflammatory cytokines and histopathology in the brains of rats infected with *Trypanosoma brucei*. *J Comp Neurol* 414, 114–130.
- Quan, N., He, L. L., & Lai, W. M. (2003). Intraventricular infusion of antagonists of IL-1 and TNF alpha attenuates neurodegeneration induced by the infection of *Trypanosoma brucei*. *J Neuroimmunol* 138, 92–98.
- Raghavan, A. V., Horowitz, J. M., & Fuller, C. A. (1999). Diurnal modulation of long-term potentiation in the hamster hippocampal slice. *Brain Res* 833, 311–314.
- Rahman, M. M., Barrett, J. W., Brouckaert, P., & McFadden, G. (2006). Variation in ligand binding specificities of a novel class of poxvirus-encoded tumor necrosis factor-binding protein. *J Biol Chem* 281, 22517–22526.
- Raison, C. L., Capuron, L., & Miller, A. H. (2006). Cytokines sing the blues: inflammation and the pathogenesis of depression. *Trends Immunol* 27, 24–31.
- Ralay Ranaivo, H., Craft, J. M., Hu, W., Guo, L., Wing, L. K., Van Eldik, L. J., et al. (2006). Glia as a therapeutic target: selective suppression of human amyloid-beta-induced upregulation of brain proinflammatory cytokine production attenuates neurodegeneration. *J Neurosci* 26, 662–670.
- Ramesh, G., Borda, J. T., Dufour, J., Kaushal, D., Ramamoorthy, R., Lackner, A. A., et al. (2008). Interaction of the Lyme disease spirochete *Borrelia burgdorferi* with brain

- parenchyma elicits inflammatory mediators from glial cells as well as glial and neuronal apoptosis. *Am J Pathol* 173, 1415–1427.
- Ranges, G. E., Zlotnik, A., Espevik, T., Dinarello, C. A., Cerami, A., & Palladino, M. A. (1988). Tumor necrosis factor alpha/cachectin is a growth factor for thymocytes. Synergistic interactions with other cytokines. *J Exp Med* 167, 1472–1478.
- Rao, V., Rosenberg, P., Bertrand, M., Salehnia, S., Spiro, J., Vaishnavi, S., et al. (2009). Aggression after traumatic brain injury: Prevalence and correlates. *J Neuropsychiatry Clin Neurosci* 21, 420–429.
- Rebel, V. I., Hartnett, S., Hill, G. R., Lazo Kallanian, S. B., Ferrara, J. L., & Sieff, C. A. (1999). Essential role for the p55 tumor necrosis factor receptor in regulating hematopoiesis at a stem cell level. *J Exp Med* 190, 1493–1504.
- Reed-Geaghan, E. G., Savage, J. C., Hise, A. G., & Landreth, G. E. (2009). CD14 and toll-like receptors 2 and 4 are required for fibrillar A(beta)-stimulated microglial activation. *J Neurosci* 29, 11982–11992.
- Relkin, N. R., Szabo, P., Adamiak, B., Burgut, T., Monthe, C., Lent, R. W., et al. (2009). 18-month study of intravenous immunoglobulin for treatment of mild Alzheimer disease. *Neurobiol Aging* 30, 1728–1736.
- Rhodes, M. A., Carraway, M. S., Piantadosi, C. A., Reynolds, C. M., Cherry, A. D., Wester, T. E., et al. (2009). Carbon monoxide, skeletal muscle oxidative stress, and mitochondrial biogenesis in humans. *Am J Physiol* 297, H392–H399.
- Riazi, K., Galic, M. A., Kuzmiski, J. B., Ho, W., Sharkey, K. A., & Pittman, Q. J. (2008). Microglial activation and TNF(alpha) production mediate altered CNS excitability following peripheral inflammation. *Proc Natl Acad Sci USA* 105, 17151–17156.
- Ricote, M., Li, A. C., Willson, T. M., Kelly, C. J., & Glass, C. K. (1998). The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. *Nature* 391, 79–82.
- Riddell, D. R., Zhou, H., Atchison, K., Warwick, H. K., Atkinson, P. J., Jefferson, J., et al. (2008). Impact of apolipoprotein E (ApoE) polymorphism on brain ApoE levels. *J Neurosci* 28, 11445–11453.
- Rijkeboer, A., Voskuyl, A., & Van Agtmael, M. (2007). Fatal *Salmonella enteritidis* septicemia in a rheumatoid arthritis patient treated with a TNF-alpha antagonist. *Scand J Infect Dis* 39, 80–83.
- Riley, M. V., & Maegraith, B. G. (1962). A factor in the serum of malaria-infected animals capable of inhibiting the in vitro oxidative metabolism of normal liver mitochondria. *Ann Trop Med Parasitol* 55, 489–497.
- Risner, M. E., Saunders, A. M., Altman, J. F., Ormandy, G. C., Craft, S., Foley, I. M., et al. (2006). Efficacy of rosiglitazone in a genetically defined population with mild-to-moderate Alzheimer's disease. *Pharmacogenomics* 7, 246–254.
- Rodgers, J. T., Lerin, C., Haas, W., Gygi, S. P., Spiegelman, B. M., & Puigserver, P. (2005). Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature* 434, 113–118.
- Rola, R., Raber, J., Rizk, A., Otsuka, S., VandenBerg, S. R., Morhardt, D. R., et al. (2004). Radiation-induced impairment of hippocampal neurogenesis is associated with cognitive deficits in young mice. *Exp Neurol* 188, 316–330.
- Ron-Harel, N., & Schwartz, M. (2009). Immune senescence and brain aging: Can rejuvenation of immunity reverse memory loss? *Trends Neurosci* 32, 367–375.
- Ron-Harel, N., Segev, Y., Lewitus, G. M., Cardon, M., Ziv, Y., Netanel, D., et al. (2008). Age-dependent spatial memory loss can be partially restored by immune activation. *Rejuvenation Res* 11, 903–913.
- Rosen, F. S. (1993). Putative mechanisms of the effect of intravenous gamma-globulin. *Clin Immunol Immunopathol* 67, S41–S43.
- Rossi, D., Brambilla, L., Valori, C. F., Crugnola, A., Giaccone, G., Capobianco, R., et al. (2005). Defective tumor necrosis factor-alpha-dependent control of astrocyte glutamate release in a transgenic mouse model of Alzheimer disease. *J Biol Chem* 280, 42088–42096.
- Rothe, J., Lesslauer, W., Lotscher, H., Lang, Y., Koebel, P., Kontgen, F., et al. (1993). Mice lacking the tumour necrosis factor receptor-1 are resistant to TNF-mediated toxicity but highly susceptible to infection by listeria monocytogenes. *Nature* 364, 798–802.
- Rowan, M. J., Klyubin, I., Wang, Q., Hu, N. W., & Anwyl, R. (2007). Synaptic memory mechanisms: Alzheimer's disease amyloid beta-peptide-induced dysfunction. *Biochem Soc Trans* 35, 1219–1223.
- Rubenstein, J. L., Combs, D., Rosenberg, J., Levy, A., McDermott, M., Damon, L., et al. (2003). Rituximab therapy for CNS lymphomas: Targeting the leptomeningeal compartment. *Blood* 101, 466–468.
- Rubenstein, J. L., Fridlyand, J., Abrey, L., Shen, A., Karch, J., Wang, E., et al. (2007). Phase I study of intraventricular administration of rituximab in patients with recurrent CNS and intraocular lymphoma. *J Clin Oncol* 25, 1350–1356.
- Ruddle, N. H., & Waksman, B. H. (1967). Cytotoxic effect of lymphocyte-antigen interaction in delayed hypersensitivity. *Science* 157, 1060–1062.
- Ryu, J. K., & McLarnon, J. G. (2008). Thalidomide inhibition of perturbed vasculature and glial-derived tumor necrosis factor-alpha in an animal model of inflamed Alzheimer's disease brain. *Neurobiol Dis* 29, 254–266.
- Sabo, T., Lomnitski, L., Nyska, A., Beni, S., Maronpot, R. R., Shohami, E., et al. (2000). Susceptibility of transgenic mice expressing human apolipoprotein E to closed head injury: the allele E3 is neuroprotective whereas E4 increases fatalities. *Neuroscience* 101, 879–884.
- Sakr, M. F., McClain, C. J., Gavalier, J. S., Zetti, G. M., Starzl, T. E., & Van Thiel, D. H. (1993). FK 506 pre-treatment is associated with reduced levels of tumor necrosis factor and interleukin 6 following hepatic ischemia/reperfusion. *J Hepatol* 17, 301–307.
- Sakuma, S., Kato, Y., Nishigaki, F., Sasakawa, T., Magari, K., Miyata, S., et al. (2000). FK506 potentially inhibits T cell activation induced TNF-alpha and IL-1beta production in vitro by human peripheral blood mononuclear cells. *Br J Pharmacol* 130, 1655–1663.
- Sallakci, N., Akcurin, G., Koksoy, S., Kardelen, F., Uguz, A., Coskun, M., et al. (2005). TNF-alpha G-308A polymorphism is associated with rheumatic fever and correlates with increased TNF-alpha production. *J Autoimmun* 25, 150–154.
- Salloway, S., Sperling, R., Gilman, S., Fox, N. C., Blennow, K., Raskind, M., et al. (2009). A phase 2 multiple ascending dose trial of bapineuzumab in mild to moderate Alzheimer disease. *Neurology* 73, 2061–2070.
- Salminen, A., Kauppinen, A., Suuronen, T., & Kaarniranta, K. (2008). SIRT1 longevity factor suppresses NF-kappaB-driven immune responses: Regulation of aging via NF-kappaB acetylation? *Bioessays* 30, 939–942.
- Sanchez-Alcazar, J. A., Schneider, E., Martinez, M. A., Carmona, P., Hernandez-Munoz, I., Siles, E., et al. (2000). Tumor necrosis factor-alpha increases the steady-state reduction of cytochrome b of the mitochondrial respiratory chain in metabolically inhibited L929 cells. *J Biol Chem* 275, 13353–13361.
- Sawle, P., Foresti, R., Mann, B. E., Johnson, T. R., Green, C. J., & Motterlini, R. (2005). Carbon monoxide-releasing molecules (CO-RMs) attenuate the inflammatory response elicited by lipopolysaccharide in RAW264.7 murine macrophages. *Br J Pharmacol* 145, 800–810.
- Scaffidi, P., Misteli, T., & Bianchi, M. E. (2002). Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. *Nature* 418, 191–195.
- Schipper, H. M. (2004). Heme oxygenase-1: Transducer of pathological brain iron sequestration under oxidative stress. *Ann NY Acad Sci*, 84–93.
- Schipper, H. M., Cisse, S., & Stopa, E. G. (1995). Expression of heme oxygenase-1 in the senescent and Alzheimer-diseased brain. *Ann Neurol* 37, 758–768.
- Schmidt, J., Barthel, K., Wrede, A., Salajegheh, M., Bahr, M., & Dalakas, M. C. (2008). Interrelation of inflammation and APP in sIBM: IL-1 beta induces accumulation of beta-amyloid in skeletal muscle. *Brain* 131, 1228–1240.
- Schmitt, F. A., Davis, D. G., Wekstein, D. R., Smith, C. D., Ashford, J. W., & Markesbery, W. R. (2000). "Preclinical" AD revisited: neuropathology of cognitively normal older adults. *Neurology* 55, 370–376.
- Schuitmaker, A., Dik, M. G., Veerhuis, R., Scheltens, P., Schoonenboom, N. S., Hack, C. E., et al. (2009). Inflammatory markers in AD and MCI patients with different biomarker profiles. *Neurobiol Aging* 30, 1885–1889.
- Schwartz, M. (in press). "Tissue-repairing" blood-derived macrophages are essential for healing of the injured spinal cord: From skin-activated macrophages to infiltrating blood-derived cells? *Brain Behav Immun*. doi:10.1016/j.bbi.2010.01.010.
- Scott, M. G., & Hancock, R. E. (2000). Cationic antimicrobial peptides and their multifunctional role in the immune system. *Crit Rev Immunol* 20, 407–431.
- Selkoe, D. J. (2002). Alzheimer's disease is a synaptic failure. *Science* 298, 789–791.
- Selkoe, D. J., Podlisny, M. B., Joachim, C. L., Vickers, E. A., Lee, G., Fritz, L. C., et al. (1988). Beta-amyloid precursor protein of Alzheimer disease occurs as 110- to 135-kilodalton membrane-associated proteins in neural and nonneural tissues. *Proc Natl Acad Sci USA* 85, 7341–7345.
- Seo, S. J., Ahn, S. W., Hong, C. K., & Ro, B. I. (2001). Expressions of beta-defensins in human keratinocyte cell lines. *J Dermatol Sci* 27, 183–191.
- Serghides, L., Patel, S. N., Ayi, K., Lu, Z., Gowda, D. C., Liles, W. C., et al. (2009). Rosiglitazone modulates the innate immune response to *Plasmodium falciparum* infection and improves outcome in experimental cerebral malaria. *J Infect Dis* 199, 1536–1545.
- Shaffer, N., Grau, G. E., Hedberg, K., Davachi, F., Lyamba, B., Hightower, A. W., et al. (1991). Tumor necrosis factor and severe malaria. *J Infect Dis* 163, 96–101.
- Shankar, G. M., Bloodgood, B. L., Townsend, M., Walsh, D. M., Selkoe, D. J., & Sabatini, B. L. (2007). Natural oligomers of the Alzheimer amyloid-beta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. *J Neurosci* 27, 2866–2875.
- Sharief, M. K., Ingram, D. A., Swash, M., & Thompson, E. J. (1999). I.v. immunoglobulin reduces circulating proinflammatory cytokines in Guillain-Barre syndrome. *Neurology* 52, 1833–1838.
- Shen, Z., Ajmo, J. M., Rogers, C. Q., Liang, X., Le, L., Murr, M. M., et al. (2009). Role of SIRT1 in regulation of LPS- or two ethanol metabolites-induced TNF-alpha production in cultured macrophage cell lines. *Am J Physiol* 296, G1047–G1053.
- Sheng, J. G., Bora, S. H., Xu, G., Borchelt, D. R., Price, D. L., & Koliatsos, V. E. (2003). Lipopolysaccharide-induced-neuroinflammation increases intracellular accumulation of amyloid precursor protein and amyloid beta peptide in APPsw transgenic mice. *Neurobiol Dis* 14, 133–145.
- Shohami, E., Bass, R., Wallach, D., Yamin, A., & Gallily, R. (1996). Inhibition of tumor necrosis factor alpha (TNF alpha) activity in rat brain is associated with cerebroprotection after closed head injury. *J Cereb Blood Flow Metab* 16, 378–384.
- Shohami, E., Gallily, R., Mechoulam, R., Bass, R., & Ben Hur, T. (1997). Cytokine production in the brain following closed head injury: dexamethasone (HU-211) is a novel TNF-alpha inhibitor and an effective neuroprotectant. *J Neuroimmunol* 72, 169–177.
- Sierksma, A. S., van den Hove, D. L., Steinbusch, H. W., & Prickaerts, J. (2010). Major depression, cognitive dysfunction and Alzheimer's disease: is there a link? *Eur J Pharmacol* 626, 72–82.
- Simon, H. U., & Spath, P. J. (2003). IVIG-mechanisms of action. *Allergy* 58, 5435–5452.
- Simon, A. M., Schiapparelli, L., Salazar-Colocho, P., Cuadrado-Tejedor, M., Escrbano, L., Lopez de Maturana, R., et al. (2009). Overexpression of wild-type human APP in mice causes cognitive deficits and pathological features unrelated to A-beta levels. *Neurobiol Dis* 33, 369–378.
- Singer, M., De Santis, V., Vitale, D., & Jeffcoate, W. (2004). Multiorgan failure is an adaptive, endocrine-mediated, metabolic response to overwhelming systemic inflammation. *Lancet* 364, 545–548.
- Sjogren, M., Folkesson, S., Blennow, K., & Tarkowski, E. (2004). Increased intrathecal inflammatory activity in frontotemporal dementia: Pathophysiological implications. *J Neurol Neurosurg Psychiatry* 75, 1107–1111.
- Slifman, N. R., Gershon, S. K., Lee, J. H., Edwards, E. T., & Braun, M. M. (2003). *Listeria monocytogenes* infection as a complication of treatment with tumor necrosis factor alpha-neutralizing agents. *Arthritis Rheum* 48, 319–324.

- Smart, T. G., Xie, X., & Krishek, B. J. (1994). Modulation of inhibitory and excitatory amino acid receptor ion channels by zinc. *Prog Neurobiol* 42, 393–441.
- Smith, R. S. (1991). The macrophage theory of depression. *Med Hypotheses* 35, 298–306.
- Smith, J. J., Kenney, R. D., Gagne, D. J., Frushour, B. P., Ladd, W., Galonek, H. L., et al. (2009). Small molecule activators of SIRT1 replicate signaling pathways triggered by calorie restriction in vivo. *BMC Syst Biol* 3, 31.
- Smith, J. P., Lal, V., Bowser, D., Cappai, R., Masters, C. L., & Ciccotosto, G. D. (2009). Stimulus pattern dependence of the Alzheimer's disease amyloid-beta 42 peptide's inhibition of long term potentiation in mouse hippocampal slices. *Brain Res* 7, 176–184.
- Snyder, J. S., Hong, N. S., McDonald, R. J., & Wojtowicz, J. M. (2005). A role for adult neurogenesis in spatial long-term memory. *Neuroscience* 130, 843–852.
- Soczynska, J. K., Kennedy, S. H., Goldstein, B. I., Lachowski, A., Woldeyohannes, H. O., & McIntyre, R. S. (2009). The effect of tumor necrosis factor antagonists on mood and mental health-associated quality of life: Novel hypothesis-driven treatments for bipolar depression? *Neurotoxicology* 30, 497–521.
- Sommer, G., Kralisch, S., Lipfert, J., Weise, S., Krause, K., Jessnitzer, B., et al. (2009). Amyloid precursor protein expression is induced by tumor necrosis factor alpha in 3T3-L1 adipocytes. *J Cell Biochem* 108, 1418–1422.
- Song, W., Patel, A., Qureshi, H. Y., Han, D., Schipper, H. M., & Paudel, H. K. (2009). The Parkinson disease-associated A30P mutation stabilizes alpha-synuclein against proteasomal degradation triggered by heme oxygenase-1 over-expression in human neuroblastoma cells. *J Neurochem* 110, 719–733.
- Sonmez, U., Sonmez, A., Erbil, G., Tekmen, I., & Baykara, B. (2007). Neuroprotective effects of resveratrol against traumatic brain injury in immature rats. *Neurosci Lett* 420, 133–137.
- Socia, S. J., Kirby, J. E., Washicosky, K. J., Tucker, S. M., Ingelsson, M., Hyman, B., et al. (2010). The Alzheimer's disease-associated amyloid beta protein is an antimicrobial peptide. *PLoS ONE* 5, e9505.
- Spaulding, C. C., Walford, R. L., & Effros, R. B. (1997). Calorie restriction inhibits the age-related dysregulation of the cytokines TNF-alpha and IL-6 in C3B10RF1 mice. *Mech Ageing Dev* 93, 87–94.
- Spilman, P., Poduitskay, N., Hart, M. J., Debnath, J., Gorostiza, O., Bredesen, D., et al. (2010). Inhibition of mTOR by rapamycin abolishes cognitive deficits and reduces amyloid- $\beta$  levels in a mouse model of Alzheimer's disease. *PLoS ONE* 5, e9979.
- Squadrito, F., Altavilla, D., Squadrito, G., Ferlito, M., Campo, G. M., Arlotta, M., et al. (1999). Tacrolimus suppresses tumour necrosis factor-alpha and protects against splanchic artery occlusion shock. *Br J Pharmacol* 127, 498–504.
- Steinshamm, S., & Waage, A. (2000). Lack of endotoxin tolerance with respect to TNF alpha production in the subarachnoid space. *APMS* 108, 107–112.
- Stellwagen, D., & Malenka, R. C. (2006). Synaptic scaling mediated by glial TNF-alpha. *Nature* 440, 1054–1059.
- Stellwagen, D., Beattie, E. C., Seo, J. Y., & Malenka, R. C. (2005). Differential regulation of AMPA receptor and GABA receptor trafficking by tumor necrosis factor-alpha. *J Neurosci* 25, 3219–3228.
- Stevens, R. D., & Pronovost, P. J. (2006). The spectrum of encephalopathy in critical illness. *Semin Neurol* 26, 440–451.
- Stewart, J., Mitchell, J., & Kalant, N. (1989). The effects of life-long food restriction on spatial memory in young and aged Fischer 344 rats measured in the eight-arm radial and the Morris water mazes. *Neurobiol Aging* 10, 669–675.
- Strittmatter, W. J., Saunders, A. M., Schmechel, D., Pericak Vance, M., Englund, J., Salvesen, G. S., et al. (1993). Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci USA* 90, 1977–1981.
- Suarez, E. C., Lewis, J. G., & Kuhn, C. (2002). The relation of aggression, hostility, and anger to lipopolysaccharide-stimulated tumor necrosis factor (TNF)-alpha by blood monocytes from normal men. *Brain Behav Immun* 16, 675–684.
- Suarez, E. C., Lewis, J. G., Krishnan, R. R., & Young, K. H. (2004). Enhanced expression of cytokines and chemokines by blood monocytes in vitro lipopolysaccharide stimulation are associated with hostility and severity of depressive symptoms in healthy women. *Psychoneuroendocrinology* 29, 1119–1128.
- Suliman, H. B., Welty Wolf, K. E., Carraway, M. S., Schwartz, D. A., Hollingsworth, J. W., & Piantadosi, C. A. (2005). Toll-like receptor 4 mediates mitochondrial DNA damage and biogenic responses after heat-inactivated *E. coli*. *FASEB J* 19, 1531–1533.
- Suliman, H. B., Carraway, M. S., Ali, A. S., Reynolds, C. M., Welty Wolf, K. E., & Piantadosi, C. A. (2007). The CO/HO system reverses inhibition of mitochondrial biogenesis and prevents murine doxorubicin cardiomyopathy. *J Clin Invest* 117, 3730–3741.
- Sundararajan, S., Jiang, Q., Heneka, M., & Landreth, G. (2006). PPARgamma as a therapeutic target in central nervous system diseases. *Neurochem Int* 49, 1361–???
- Tagliatalata, G., Hogan, D., Zhang, W. R., & Dineley, K. T. (2009). Intermediate- and long-term recognition memory deficits in Tg2576 mice are reversed with acute calcineurin inhibition. *Behav Brain Res* 200, 959–???
- Taille, C., Foresti, R., Lanone, S., Zedda, C., Green, C., Aubier, M., et al. (2001). Protective role of heme oxygenases against endotoxin-induced diaphragmatic dysfunction in rats. *Am J Respir Crit Care Med* 163, 753–761.
- Takeda, A., Takefuta, S., Okada, S., & Oku, N. (2000). Relationship between brain zinc and transient learning impairment of adult rats fed zinc-deficient diet. *Brain Res* 859, 352–357.
- Tancredi, V., D'Arcangelo, G., Grassi, F., Tarroni, P., Palmieri, G., Santoni, A., et al. (1992). Tumor necrosis factor alters synaptic transmission in rat hippocampal slices. *Neurosci Lett* 146, 17617–17618.
- Tang, B. L. (2009). Resveratrol is neuroprotective because it is not a direct activator of Sirt1-A hypothesis. *Brain Res Bull* 21, 21.
- Tangpong, J., Sompol, P., Vore, M., St Clair, W., Butterfield, D. A., & St Clair, D. K. (2008). Tumor necrosis factor alpha-mediated nitric oxide production enhances manganese superoxide dismutase nitration and mitochondrial dysfunction in primary neurons: an insight into the role of glial cells. *Neuroscience* 151, 622–629.
- Tarkowski, E., Andreasen, N., Tarkowski, A., & Blennow, K. (2003). Intrathecal inflammation precedes development of Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 74, 1200–1205.
- Tarkowski, E., Liljeroth, A. M., Minthon, L., Tarkowski, A., Wallin, A., & Blennow, K. (2003). Cerebral pattern of pro- and anti-inflammatory cytokines in dementias. *Brain Res Bull* 61, 255–260.
- Taverne, J., Dockrell, H. M., & Playfair, J. H. L. (1981). Endotoxin-induced serum factor kills malarial parasites in vitro. *Infect Immun* 33, 83–89.
- Taylor, C. J., Ireland, D. R., Ballagh, I., Bourne, K., Marechal, N. M., Turner, P. R., et al. (2008). Endogenous secreted amyloid precursor protein-alpha regulates hippocampal NMDA receptor function, long-term potentiation and spatial memory. *Neurobiol Dis* 31, 250–260.
- Thal, D. R., Capetillo Zarate, E., Larionov, S., Staufenbiel, M., Zurbrugg, S., & Beckmann, N. (2009). Capillary cerebral amyloid angiopathy is associated with vessel occlusion and cerebral blood flow disturbances. *Neurobiol Aging* 30, 1936–1948.
- Thambisetty, M., Simmons, A., Velayudhan, L., Hye, A., Campbell, J., Zhang, Y., et al. (2010). Association of plasma clusterin concentration with severity, pathology, and progression in Alzheimer's disease. *Arch Gen Psychiatry* 67, 739–748.
- Thinakaran, G., & Koo, E. H. (2008). Amyloid precursor protein trafficking, processing, and function. *J Biol Chem* 283, 29615–29619.
- Thomson, A. W., Turnquist, H. R., & Raimondi, G. (2009). Immunoregulatory functions of mTOR inhibition. *Nat Rev Immunol* 9, 324–337.
- Titus, R. G., Sherry, B., & Cerami, A. (1989). Tumor necrosis factor plays a protective role in experimental murine cutaneous leishmaniasis. *J Exp Med* 170, 2097–2104.
- Tjitra, E., Anstey, N. M., Sugiarto, P., Warikar, N., Kenangalem, E., Karyana, M., et al. (2008). Multidrug-resistant *Plasmodium vivax* associated with severe and fatal malaria: a prospective study in Papua, Indonesia. *PLoS Med* 5, e128.
- Tobinick, E. (2009). Tumour necrosis factor modulation for treatment of Alzheimer's disease: rationale and current evidence. *CNS Drugs* 23, 713–725.
- Tobinick, E. (2010). Perispinal etanercept: A new therapeutic paradigm in neurology. *Expert Rev Neurother* 10, 985–1002.
- Tobinick, E. L., & Gross, H. (2008). Rapid cognitive improvement in Alzheimer's disease following perispinal etanercept administration. *J Neuroinflammation* 5, 2.
- Tobinick, E. L., Gross, H., Weinberger, A., & Cohen, H. (2006). TNF-alpha modulation for treatment of Alzheimer's disease: A 6-month pilot study. *MedGenMed Neurol Neurosurg* 8, 25.
- Tobinick, E. L., Chen, K., & Chen, X. (2009). Rapid intracerebroventricular delivery of Cu-DOTA-etanercept after peripheral administration demonstrated by PET imaging. *BMC Res Notes* 2, 28.
- Tocci, M. J., Matkovich, D. A., Collier, K. A., Kwok, P., Dumont, F., Lin, S., et al. (1989). The immunosuppressant FK506 selectively inhibits expression of early T cell activation genes. *J Immunol* 143, 718–726.
- Tokarevich, N. K., Prokopyev, A. A., Prokopyeva, E. D., Simbirtsev, A. S., Toropova, B. G., Daiter, A. B., et al. (1992). Role of tumor necrosis factor and interleukin-1 in the formation of resistance in experimental Q fever. *Zh Mikrobiol Epidemiol Immunobiol* 5, 46–47.
- Toungouz, M., Denys, C., & Dupont, E. (1996). Blockade of proliferation and tumor necrosis factor-alpha production occurring during mixed lymphocyte reaction by interferon-gamma-specific natural antibodies contained in intravenous immunoglobulins. *Transplantation* 62, 1292–1296.
- Tracey, D., Klareskog, L., Sasso, E. H., Salfeld, J. G., & Tak, P. P. (2008). Tumor necrosis factor antagonist mechanisms of action: A comprehensive review. *Pharmacol Ther* 117, 244–279.
- Trapp, B. D., & Hauer, P. E. (1994). Amyloid precursor protein is enriched in radial glia: Implications for neuronal development. *J Neurosci Res* 37, 538–550.
- Tsutsui, H., Matsui, K., Kawada, N., Hyodo, Y., Hayashi, N., Okamura, H., et al. (1997). IL-18 accounts for both TNF-alpha- and Fas ligand-mediated hepatotoxic pathways in endotoxin-induced liver injury in mice. *J Immunol* 159, 3961–3967.
- Tufo, H. M., Ostfeld, A. M., & Shekelle, R. (1970). Central nervous system dysfunction following open-heart surgery. *J Am Med Assoc* 212, 1333–1340.
- Tukel, C., Wilson, R. P., Nishimori, J. H., Pezeshki, M., Chromy, B. A., & Baumler, A. J. (2009). Responses to amyloids of microbial and host origin are mediated through toll-like receptor 2. *Cell Host Microbe* 6, 45–53.
- Turco, J., & Winkler, H. H. (1993). Role of the nitric oxide synthase pathway in inhibition of growth of interferon-sensitive and interferon-resistant *Rickettsia prowazekii* strains in L929 cells treated with tumor necrosis factor alpha and gamma interferon. *Infect Immun* 61, 4317–4325.
- Turnbull, A. V., Pitossi, F. J., Lebrun, J. J., Lee, S., Meltzer, J. C., Nance, D. M., et al. (1997). Inhibition of tumor necrosis factor-alpha action within the CNS markedly reduces the plasma adrenocorticotropin response to peripheral local inflammation in rats. *J Neurosci* 17, 3262–3273.
- Turrigiano, G. G. (2008). The self-tuning neuron: synaptic scaling of excitatory synapses. *Cell* 135, 422–435.
- Tweedie, D., Sambamurti, K., & Greig, N. H. (2007). TNF-alpha inhibition as a treatment strategy for neurodegenerative disorders: New drug candidates and targets. *Curr Alzheimer Res* 4, 378–385.
- Tyring, S. K., Cauda, R., Tumbarello, M., Ortona, L., Kennedy, R. C., Chanh, T. C., et al. (1991). Synthetic peptides corresponding to sequences in HIV envelope gp41 and gp120 enhance in vitro production of interleukin-1 and tumor necrosis factor but depress production of interferon-alpha, interferon-gamma and interleukin-2. *Viral Immunol* 4, 33–42.

- Ugochukwu, N. H., & Figgers, C. L. (2007). Caloric restriction inhibits up-regulation of inflammatory cytokines and TNF- $\alpha$ , and activates IL-10 and haptoglobin in the plasma of streptozotocin-induced diabetic rats. *J Nutr Biochem* 18, 120–126.
- Uguz, F., Akman, C., Kucuksarac, S., & Tufekci, O. (2009). Anti-tumor necrosis factor- $\alpha$  therapy is associated with less frequent mood and anxiety disorders in patients with rheumatoid arthritis. *Psychiatry Clin Neurosci* 63, 50–55.
- van der Flier, W. M., Staekenborg, S., Pijnenburg, Y. A., Gillissen, F., Romkes, R., Kok, A., et al. (2007). Apolipoprotein E genotype influences presence and severity of delusions and aggressive behavior in Alzheimer disease. *Dement Geriatr Cogn Disord* 23, 42–46.
- van der Meer, J. W. (1988). The effects of recombinant interleukin-1 and recombinant tumor necrosis factor on non-specific resistance to infection. *Biotherapy* 1, 19–25.
- Van Eldik, L. J., & Griffin, W. S. (1994). S100 beta expression in Alzheimer's disease: Relation to neuropathology in brain regions. *Biochim Biophys Acta* 1223, 398–403.
- van Gool, W. A., van de Beek, D., & Eikelenboom, P. (2010). Systemic infection and delirium: When cytokines and acetylcholine collide. *Lancet* 375, 773–775.
- Van Hensbroek, M. B., Palmer, A., Onyiorah, E., Schneider, G., Jaffar, S., Dolan, G., et al. (1996). The effect of a monoclonal antibody to tumor necrosis factor on survival from childhood cerebral malaria. *J Infect Dis* 174, 1091–1097.
- Van Reeth, K. (2000). Cytokines in the pathogenesis of influenza. *Vet Microbiol* 74, 109–116.
- Velliquette, R. A., O'Connor, T., & Vassar, R. (2005). Energy inhibition elevates beta-secretase levels and activity and is potentially amyloidogenic in APP transgenic mice: Possible early events in Alzheimer's disease pathogenesis. *J Neurosci* 25, 10874–10883.
- Vitek, M. P., Brown, C. M., & Colton, C. A. (2009). APOE genotype-specific differences in the innate immune response. *Neurobiol Aging* 30, 1350–1360.
- Volterra, A., & Meldolesi, J. (2005). Astrocytes, from brain glue to communication elements: The revolution continues. *Nat Rev Neurosci* 6, 626–640.
- Waage, A., Halstensen, A., & Espevik, T. (1987). Association between tumour necrosis factor in serum and fatal outcome in patients with meningococcal disease. *Lancet*, 355–357.
- Waelput, W., Broekaert, D., Vandekerckhove, J., Brouckaert, P., Tavernier, J., & Libert, C. (2001). A mediator role for metallothionein in tumor necrosis factor-induced lethal shock. *J Exp Med* 194, 1617–1624.
- Wallwork, J. C. (1987). Zinc and the central nervous system. *Prog Food Nutr Sci* 11, 203–247.
- Walsh, D. M., Klyubin, I., Fadeeva, J. V., Cullen, W. K., Anwyl, R., Wolfe, M. S., et al. (2002). Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416, 535–539.
- Wang, Q. W., Wu, J. Q., Rowan, M. J., & Anwyl, R. (2005). Beta-amyloid inhibition of long-term potentiation is mediated via tumor necrosis factor. *Eur J Neurosci* 22, 2827–2832.
- Wang, J. M., Liu, L., Irwin, R. W., Chen, S., & Brinton, R. D. (2008). Regenerative potential of allopregnanolone. *Brain Res Rev* 57, 398–409.
- Wang, G. Y., Chen, G. H., Li, H., Huang, Y., Wang, G. S., Jiang, N., et al. (2009). Rapamycin-treated mature dendritic cells have a unique cytokine secretion profile and impaired allostimulatory capacity. *Transpl Int* 22, 1005–1016.
- Wang, H., Christensen, D. J., Vitek, M. P., Sullivan, P. M., & Laskowitz, D. T. (2009). APOE genotype affects outcome in a murine model of sepsis: Implications for a new treatment strategy. *Anaesth Intensive Care* 37, 38–45.
- Watson, G. S., Cholerton, B. A., Reger, M. A., Baker, L. D., Plymate, S. R., Asthana, S., et al. (2005). Preserved cognition in patients with early Alzheimer disease and amnesic mild cognitive impairment during treatment with rosiglitazone: A preliminary study. *Am J Geriatr Psychiatry* 13, 950–958.
- Weberpals, M., Hermes, M., Hermann, S., Kummer, M. P., Terwel, D., Semmler, A., et al. (2009). NOS2 gene deficiency protects from sepsis-induced long-term cognitive deficits. *J Neurosci* 29, 14177–14184.
- Weir, M. R., Diekmann, F., Flechner, S. M., Lebranchu, Y., Mandelbrot, D. A., Oberbauer, R., et al. (2010). mTOR inhibition: the learning curve in kidney transplantation. *Transpl Int* 23, 447–460.
- Weksler, M. E., Gouras, G., Relkin, N. R., & Szabo, P. (2005). The immune system, amyloid-beta peptide, and Alzheimer's disease. *Immunol Rev* 205, 244–256.
- Weller, R. O., Preston, S. D., Subash, M., & Carare, R. O. (2009). Cerebral amyloid angiopathy in the aetiology and immunotherapy of Alzheimer disease. *Alzheimers Res Ther* 1, 6.
- Wen, T. S., Randall, D. C., & Zolman, J. F. (1994). Protein accumulation in cerebrospinal fluid during -90 degrees head-down tilt in rabbit. *J Appl Physiol* 77, 1081–1086.
- Wenisch, C., Varijanonta, S., Looareesuwan, S., Graninger, W., Pichler, R., & Wernsdorfer, W. (1994). Soluble intercellular adhesion molecule-1 (ICAM-1), endothelial leukocyte adhesion molecule-1 (ELAM-1), and tumor necrosis factor receptor (55 kDa TNF-R) in patients with acute *Plasmodium falciparum* malaria. *Clin Immunol Immunopathol* 71, 344–348.
- Wheeler, D., Knapp, E., Bandaru, V. V., Wang, Y., Knorr, D., Poirier, C., et al. (2009). Tumor necrosis factor- $\alpha$ -induced neutral sphingomyelinase-2 modulates synaptic plasticity by controlling the membrane insertion of NMDA receptors. *J Neurochem* 109, 1237–1249.
- White, N. J., Turner, G. D., Medana, I. M., Dondorp, A. M., & Day, N. P. (2010). The murine cerebral malaria phenomenon. *Trends Parasitol* 26, 11–15.
- Whiteman, M., Li, L., Rose, P., Tan, C. H., Parkinson, D., & Moore, P. (2010). The effect of hydrogen sulfide donors on lipopolysaccharide-induced formation of inflammatory mediators in macrophages. *Antioxid Redox Signal* 12, 1147–1154.
- Wu, J. P., Kuo, J. S., Liu, Y. L., & Tzeng, S. F. (2000). Tumor necrosis factor- $\alpha$  modulates the proliferation of neural progenitors in the subventricular/ventricular zone of adult rat brain. *Neurosci Lett* 292, 203–206.
- Xia, Z., DePierre, J. W., & Nassberger, L. (1996). Tricyclic antidepressants inhibit IL-6, IL-1 beta and TNF- $\alpha$  release in human blood monocytes and IL-2 and interferon- $\gamma$  in T cells. *Immunopharmacology* 34, 27–37.
- Xie, X. M., & Smart, T. G. (1991). A physiological role for endogenous zinc in rat hippocampal synaptic neurotransmission. *Nature* 349, 521–524.
- Xing, H. Q., Hayakawa, H., Izumo, K., Kubota, R., Gelpi, E., Budka, H., et al. (2009). In vivo expression of proinflammatory cytokines in HIV encephalitis: an analysis of 11 autopsy cases. *Neuropathology* 29, 433–442.
- Yamaguchi, Y., Fujio, K., Shoda, H., Okamoto, A., Tsuno, N. H., Takahashi, K., et al. (2007). IL-17B and IL-17C are associated with TNF- $\alpha$  production and contribute to the exacerbation of inflammatory arthritis. *J Immunol* 179, 7128–7136.
- Yamamoto, M., Kiyota, T., Horiba, M., Buescher, J. L., Walsh, S. M., Gendelman, H. E., et al. (2007). Interferon- $\gamma$  and tumor necrosis factor- $\alpha$  regulate amyloid-beta plaque deposition and beta-secretase expression in Swedish mutant APP transgenic mice. *Am J Pathol* 170, 680–692.
- Yamasu, K., Onoe, H., Soma, G., Oshima, H., & Mizuno, D. (1989). Secretion of tumor necrosis factor during fetal and neonatal development of the mouse: Ontogenic inflammation. *J Biol Response Mod* 8, 644–655.
- Yan, S. D., Chen, X., Fu, J., Chen, M., Zhu, H., Roher, A., et al. (1996). RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. *Nature* 382, 685–691.
- Yang, Y. L., Li, J. P., Li, K. Z., & Dou, K. F. (2004). Tumor necrosis factor alpha antibody prevents brain damage of rats with acute necrotizing pancreatitis. *World J Gastroenterol* 10, 2898–2900.
- Yeung, M. C., Pulliam, L., & Lau, A. S. (1995). The HIV envelope protein gp120 is toxic to human brain-cell cultures through the induction of interleukin-6 and tumor necrosis factor- $\alpha$ . *AIDS* 9, 137–143.
- Yeung, F., Hoberg, J. E., Ramsey, C. S., Keller, M. D., Jones, D. R., Frye, R. A., et al. (2004). Modulation of NF- $\kappa$ B-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 23, 2369–2380.
- Yokota, S. (2003). Influenza-associated encephalopathy—pathophysiology and disease mechanisms. *Nippon Rinsho* 61, 1953–1958.
- Zhang, Y. H., Lin, J. X., Yip, Y. K., & Vilcek, J. (1988). Enhancement of cAMP levels and of protein kinase activity by tumor necrosis factor and interleukin 1 in human fibroblasts: Role in the induction of interleukin 6. *Proc Natl Acad Sci USA* 85, 6802–6805.
- Zhang, Q., Raouf, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., et al. (2010). Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 464, 104–107.
- Zhang, Q., Itagaki, K., & Hauser, C. J. (2009). Mitochondrial DNA is released by shock and activates neutrophils via p38 MAP-kinase. *Shock* 34, 55–59.
- Zhao, C., Deng, W., & Gage, F. H. (2008). Mechanisms and functional implications of adult neurogenesis. *Cell* 132, 645–660.
- Zhu, J., Wu, X., Goel, S., Gowda, N. M., Kumar, S., Krishnegowda, G., et al. (2009). MAPK-activated protein kinase 2 differentially regulates *Plasmodium falciparum* glycosylphosphatidylinositol induced production of TNF- $\alpha$  and IL-12 in macrophages. *J Biol Chem* 284, 15750–15761.
- Zuckerbraun, B. S., Chin, B. Y., Bilban, M., d'Avila, J. D., Rao, J., Billiar, T. R., et al. (2007). Carbon monoxide signals via inhibition of cytochrome c oxidase and generation of mitochondrial reactive oxygen species. *FASEB J* 21, 1099–1106.