

IL-27 supports germinal center function by enhancing IL-21 production and the function of T follicular helper cells

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Maturation and selection of high-affinity B cell clones in the germinal center (GC) relies on support from T follicular helper (T_{FH}) cells. T_{FH} cells are characterized by their localization to the B cell follicle and their high expression of the costimulatory molecules ICOS and PD1 and the cytokine IL-21, which promotes immunoglobulin (Ig) class switching and production by B cells. We show that the heterodimeric cytokine IL-27 is critical for the function of T_{FH} cells and for normal and pathogenic GC responses. IL-27 signaling to T cells results in the production of IL-21, a known autocrine factor for the maintenance of T_{FH} cells, in a STAT3-dependent manner. IL-27 also enhances the survival of activated CD4⁺ T cells and the expression of T_{FH} cell phenotypic markers. In vivo, expression of the IL-27R α chain is required to support IL-21 production and T_{FH} cell survival in a T cell-intrinsic manner. The production of high-affinity antibodies is reduced, and pristane-elicited autoantibodies and glomerulonephritis are significantly diminished, in *Il27ra*^{-/-} mice. Together, our data show a nonredundant role for IL-27 in the development of T cell-dependent antibody responses.

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Abbreviations used: 7AAD, 7-aminoactinomycin D; AD-HIES, autosomal dominant hyper IgE syndrome; Ag, antigen; DLN, draining LN; GC, germinal center; PAS, periodic acid-Schiff; PNA, peanut agglutinin; SLE, systemic lupus erythematosus; T_{FH}, T follicular helper; Tg, transgenic.

IL-27 is a heterodimeric cytokine consisting of the protein subunits IL27p28 and Epstein Barr virus-induced protein 3 (Pflanz et al., 2002). It signals through a heterodimeric receptor consisting of the ligand-specific IL-27R α chain and gp130 (Pflanz et al., 2004), which is shared with several other cytokines, including the structurally related cytokine IL-6. Like IL-6, IL-27 signaling involves the activation of Jak1, STAT1, and STAT3 (Batten and Ghilardi, 2007). Despite shared use of the gp130 chain, the contribution of the IL-27R α subunit makes IL-27 functionally distinct from IL-6 in that it promotes early aspects of T_H1 differentiation, such as up-regulation of the transcription factor T-bet and the IL-12 receptor β 2 chain while suppressing IL-6-driven T cell proliferation and T_H17 differentiation (Batten and Ghilardi, 2007).

In vivo, IL-27 acts to constrain inflammation under most circumstances that have been studied to date (Batten and Ghilardi, 2007). Several possible mechanisms for this immunosuppressive activity have been identified; IL-27 is known to antagonize T_H17 development (Batten et al., 2006; Stumhofer et al., 2006), induce IL-10 production (Awasthi et al., 2007; Fitzgerald et al., 2007; Stumhofer et al., 2007; Batten et al., 2008), and suppress IL-6-induced T cell proliferation (Batten et al., 2006). Nevertheless, IL-27 apparently plays a proinflammatory role in some situations. For example,

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Il27ra^{-/-} mice are protected from proteoglycan-induced arthritis (Cao et al., 2008), which is dependent on both B and CD4⁺ T cell activity (Banerjee et al., 1992; Hamel et al., 2008). Furthermore, deletion of *Il27ra* in the MRL/lpr model of lupus results in lower T_H1 cytokine production, diminished anti-dsDNA antibodies, and enhanced survival (Shimizu et al., 2005). Together, these results suggest that IL-27 is required in proinflammatory situations that depend on generation of high-affinity antibodies in vivo.

Antibody affinity maturation involves the selection of antigen (Ag)-specific B cell clones that have undergone productive somatic hypermutation. This occurs in germinal centers (GCs) in secondary lymphoid organs and relies on a specialized subset of CD4⁺ T helper cells termed T follicular helper (T_{FH}) cells (Yu et al., 2009a). Without T_{FH} cells, GCs are short lived and ineffective in generating high-affinity antibodies and B cell memory, whereas aberrant T_{FH} activity can drive autoimmune disease (King, 2009; Yu et al., 2009a). T_{FH} cells express CXCR5 and are thereby attracted to the GC by the B cell chemoattractant CXCL13. Concurrently, they down-regulate CCR7 which would otherwise retain them in the T cell zones. Besides being CXCR5⁺CCR7^{lo}, T_{FH} cells also express high levels of PD1 and ICOS (Yu et al., 2009a) and low levels of CD127 (IL7R; Lim and Kim, 2007) and CD62L (Fazilleau et al., 2009). Bcl-6 has recently been shown to be a lineage-specific transcription factor, repressing alternative T helper cell differentiation pathways and promoting T_{FH} development (Johnston et al., 2009; Nurieva et al., 2009; Yu et al., 2009b). The requirements for T_{FH} cell differentiation and maintenance are not fully understood but include sustained interactions with B cells (Haynes et al., 2007) and ICOS-ICOSL signaling (Nurieva et al., 2008). Furthermore, T_{FH}-derived IL-21 is known to provide crucial stimulation to B cells (Linterman et al., 2010; Zotos et al., 2010) and can act as an autocrine growth factor for T_{FH} cells (Nurieva et al., 2008; Vogelzang et al., 2008; Eddahri et al., 2009; Linterman et al., 2010). In this paper, we identify IL-27 as an essential cytokine for IL-21 induction, the function of T_{FH} cells and GC responses, and show that the severity of antibody-mediated autoimmune disease is reduced in the absence of IL-27 signaling in a murine model of lupus.

RESULTS

IL-27 induces IL-21 expression in vitro and in vivo

We tested the effect of IL-27 on FACS-purified naive T cells during anti-CD3/anti-CD28 stimulation and found that the addition of IL-27 resulted in greatly elevated *Il21* mRNA expression (Fig. 1 A), peaking at ~24–48 h. This was a specific response to ligand–receptor interaction because no effect was observed on IL-27Rα-deficient (*Il27ra*^{-/-}) T cells (Fig. 1 A). IL-27 was unable to significantly enhance IL-21 mRNA expression in the presence of the translational repressor cycloheximide (Fig. S1 A), suggesting that translation of intermediate proteins may mediate IL-21 expression. Analysis of the culture supernatants confirmed that IL-27 induces IL-21 protein production by >10-fold compared with untreated cells by 48 h (Fig. 1 B).

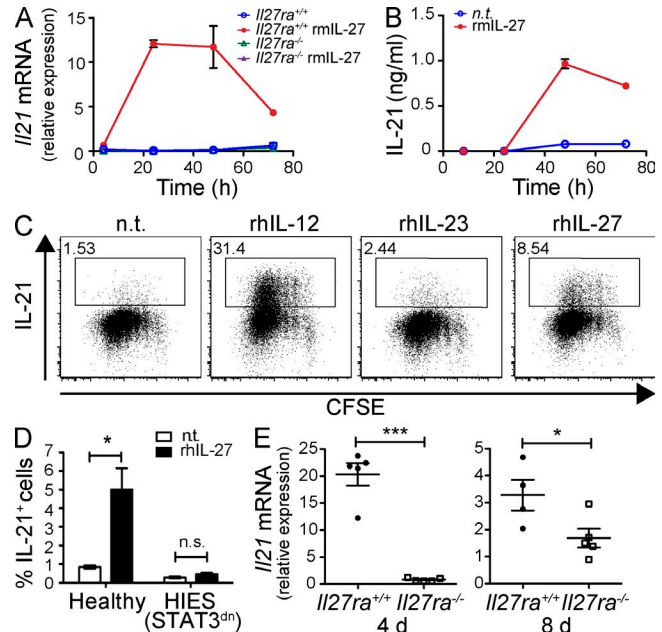


Figure 1. IL-27 induces IL-21 expression. (A) FACS-purified CD4⁺CD25⁻ T cells isolated from either *Il27ra*^{+/+} or *Il27ra*^{-/-} mice were stimulated with plate-bound anti-CD3 and soluble anti-CD28 under TH0 polarizing conditions and in the presence or absence of 20 ng/ml rmlIL-27 for the times indicated. *Il21* mRNA was determined by real-time RT-PCR and is presented relative to *rp19* mRNA. Data represent the mean of triplicate experimental samples and error bars indicate standard deviation. (B) CD4⁺ T cells isolated from C57BL/6 spleens were stimulated as in A (n.t., no treatment) and IL-21 protein was measured in the culture supernatant by ELISA. Error bars indicate standard deviation. (C) FACS-purified CD4⁺CD45RA⁺CXCR5⁻ cells from human tonsil were labeled with CFSE and stimulated with anti-CD3 + anti-CD28 + anti-CD2 beads (2:1 ratio of beads to cells) in the presence of no additional cytokine, 20 ng/ml rhIL-12 or IL-23, or 50 ng/ml rhIL-27 for 5 d. IL-21 expression was assessed by intracellular staining and flow cytometry and is plotted against CFSE division. The percentage of IL-21-expressing cells is indicated. Similar data were obtained from three individuals. (D) Naive CD4⁺ T cells were FACS purified from three healthy controls and three AD-HIES patients and stimulated as in C. The mean percentage of IL-21⁺ cells ± SEM is given. (E) Groups of *Il27ra*^{+/+} (filled circles) and *Il27ra*^{-/-} (open squares) mice were immunized with OVA (30 µg/mouse) in CFA. 4 and 8 d after immunization, CD4⁺ T cells were isolated from the spleens and IL-21 mRNA was determined by real time RT-PCR (relative to *rp19*). Data from individual animals are shown. Bars indicate the mean of five animals ± SEM. *, *P* < 0.05; ***, *P* < 0.001 (unpaired Student's *t* test). One of three (A and B) or two (E) independent experiments is shown. C shows a single donor representative of six. D shows data combined from three independent donors.

We also found that the addition of rhIL-27 during stimulation induced IL-21 expression by naive CD4⁺ T cells purified from human tonsil (Fig. 1 C) and peripheral blood (Fig. 1 D). These data suggest that IL-27 has a similar role in human and mouse IL-21 induction. In comparison to IL-27, IL-12 was a more potent IL-21 stimulant in cultures of human CD4⁺ T cells (Fig. 1 C). Because IL-27 stimulation up-regulated the expression of IL-12Rβ2 (Fig. S1 B), thereby sensitizing the cells to the effects of IL-12, these two cytokines may

work together to enhance IL-21 production. Indeed, IL-12 and IL-27 had an additive effect on IL-21 secretion by murine CD4⁺ T cells (Fig. S1 C).

A prominent feature of IL-27 signaling is the activation of STAT1; however, induction of IL-21 protein expression was not dependent on activation of this transcription factor because IL-27 was able to induce IL-21 expression by STAT1^{-/-} CD4⁺ T cells (Fig. S1 D). To investigate the importance of STAT3 activation for IL-27-induced IL-21 expression, we used naive CD4⁺ T cells from autosomal dominant hyper IgE syndrome (AD-HIES) as a result of dominant-negative mutations in *STAT3* (Minegishi et al., 2007). The addition of rhIL-27 during the stimulation of AD-HIES CD4⁺ T cells could no longer induce IL-21 expression (Fig. 1 D), indicating that the induction of IL-21 by IL-27 is STAT3 dependent.

Having established that IL-27 is sufficient to induce IL-21 production in vitro, we next sought to determine whether IL-27 signaling is required for IL-21 induction in vivo. To this end, we immunized WT and *Il27ra*^{-/-} mice with OVA/CFA and measured *Il21* mRNA expression in splenic CD4⁺ T cells 4 and 8 d after immunization. CD4⁺ T cells from *Il27ra*^{-/-} mice contained significantly diminished levels of *Il21* mRNA, demonstrating that IL-27 signals are nonredundant for IL-21 expression in vivo (Fig. 1 E). High levels of IL-21 mRNA expression by WT cells at day 4 compared with day 8 may reflect IL-21 expression by recently activated CD4⁺ T cells in addition to the fully differentiated T_{FH} cells (King, 2009).

T_{FH} cells are reduced in the absence of *Il27ra*

Because *Il27ra*^{-/-} mice had reduced IL-21 expression, and because IL-21 is both a differentiation factor for and hallmark cytokine of T_{FH} cells (Nurieva et al., 2008; Vogelzang et al., 2008; Eddahri et al., 2009; Yu et al., 2009a; Linterman et al., 2010), we investigated whether these mice had a reduced T_{FH} cell population. To this end, we immunized mice twice with TNP-OVA in Freund's adjuvant and analyzed CD4⁺ T cell phenotypes in spleen and LNs 7 d after the second immunization. A statistically significant reduction in both the percentage and absolute number of PD1⁺CXCR5⁺CD4⁺ T cells was observed in the spleens and draining LN (DLN) of *Il27ra*^{-/-} mice (Fig. 2). To ensure proper discrimination between T_{FH} and activated T cells, we stained with additional markers and found that *Il27ra*^{-/-} mice have a reduction in CD4⁺ T cells with a CXCR5⁺, PD1⁺, ICOS⁺, CCR7^{lo}, CD62L^{lo}, and CD127^{lo} cell phenotype (Fig. S2 A), a population matching the published phenotype of T_{FH} cells. Interestingly, more ICOS⁺ cells were observed in the LN compared with spleen, both in the CXCR5-positive and -negative gates, suggesting that there may be differences in the phenotype or activation level of CD4⁺ cells in the DLN (Fig. S2 A). Although the percentage of T_{FH} cells was not significantly different between WT and *Il27ra*^{-/-} mice before immunization (Fig. 2 B), we used a double immunization strategy to ensure that the response to immunization overcame any small baseline differences in T_{FH} cell numbers and to provide sufficient opportunity for the development of high-affinity

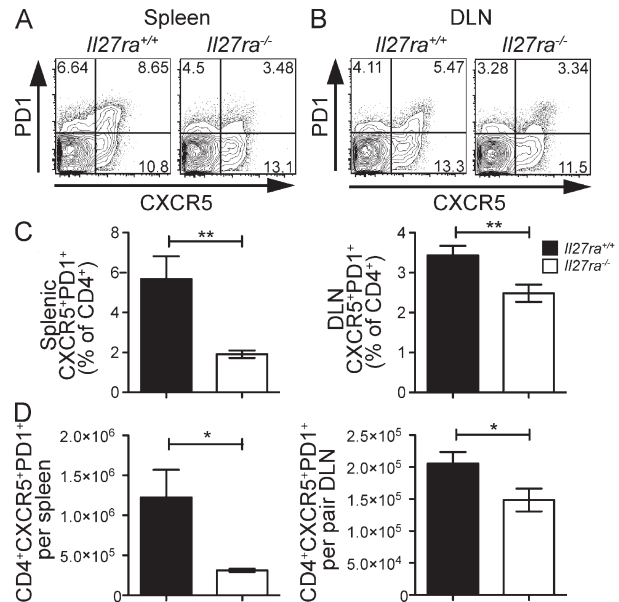


Figure 2. *Il27ra*^{-/-} mice contain fewer T_{FH} cells than *Il27ra*^{+/+} mice. Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized twice with TNP-OVA in adjuvant and, 7 d after the second immunization, tissue was collected for analysis. (A and B) Representative flow cytometric analysis of CXCR5 and PD1 expression in the CD4⁺B220⁻ gate in the spleen (A) and DLN (B). (C) The mean percentage ± SEM of CXCR5⁺PD1⁺ cells in each spleen or pair of DLN. (D) The numbers of CXCR5⁺PD1⁺ cells per organ were calculated for spleen and DLN by multiplying the percentage obtained by flow cytometry by the total cell count per organ. The mean of at least six animals per group is given and error bars indicate SEM. *, P < 0.05 (unpaired Student's *t* test). These data are representative of four individual experiments.

antibody. However, a similar requirement for *Il27ra* was observed 7 d after a single immunization (Fig. S2, C and D; and not depicted).

We reproducibly noted a significantly reduced population of T_{FH} cells in both the spleen and DLN of *Il27ra*^{-/-} mice; however, we consistently saw that the magnitude of the reduction was greatest in the spleen (Fig. 2 and Fig. S2). Inhibition of lymphocyte migration using FTY720 administration did not prevent the tissue-specific effect (Fig. S2 C), suggesting that IL-27-dependent lymphocyte migration is not responsible for these tissue-specific differences. Furthermore, although blockade of IL-21 using a soluble IL21R-Fc molecule diminished the percentage of CXCR5⁺PD1⁺ cells in WT mice, particularly in the LN, it was unable to further suppress T_{FH} cell numbers in *Il27ra*^{-/-} mice (Fig. S2 D), suggesting that these two cytokines work in the same pathway.

Reduced GC activity in the absence of *Il27ra*

Mouse strains that have a deficiency of IL-21 or T_{FH} cells also display abortive GC reactions (de Vinuesa et al., 2000; Nurieva et al., 2008; Vogelzang et al., 2008). Thus, to confirm that T_{FH} cell function was diminished, we examined the formation of GC in the spleens of immunized *Il27ra*^{-/-} mice. Flow cytometric analysis showed that significantly fewer Fas⁺GL7⁺ GC B cells were present in the absence of *Il27ra* signaling

(Fig. 3, A and B). Histological examination showed that although *Il27ra*^{-/-} mice did develop some peanut agglutinin (PNA)⁺ structures resembling GC, these were greatly reduced in size compared with WT controls (Fig. 3, C and D). The PNA-positive GC area per spleen was found to be significantly reduced in *Il27ra*^{-/-} mice (Fig. 3 D).

To examine whether this GC defect translated into lower serum levels of high-affinity antibodies, we used an established model (Roes and Rajewsky, 1993) whereby high-affinity antibodies to the immunizing hapten, TNP, can be measured in the serum by virtue of their ability to bind to sparsely haptenated BSA molecules (TNP₂-BSA). The level of total (high and low affinity) anti-TNP antibody, as detected using TNP₂₈-BSA, was similar in *Il27ra*^{-/-} and *Il27ra*^{+/+} sera (Fig. 3 E). However, the level of high-affinity anti-TNP antibodies (detected using TNP₂-BSA) was reduced in *Il27ra*^{-/-} mice compared with WT mice (Fig. 3 F), indicating that affinity maturation is compromised in the absence of IL-27 signaling. *Il27ra*^{-/-} mice had reduced levels of class-switched high-affinity antibodies, including IgG1, IgG2a, IgG2b, and IgG3 (Fig. 3 G), but not IgE (not depicted), which is expected because this isotype is inhibited by the GC transcription factor Bcl-6

(Harris et al., 1999). In general, antibody derived from GC-independent responses appears to be unaffected by the loss of IL-27 signaling. Previous studies showed that *Il27ra*^{-/-} mice displayed normal levels of total serum Ig, with the exception of IgG2a (Chen et al., 2000; Miyazaki et al., 2005). In line with these observations, we found that the early IgM response to the T cell-independent Ag, TNP-Ficoll, was similar in *Il27ra*^{+/+} and *Il27ra*^{-/-} mice (Fig. 3 H). Collectively, global absence of IL-27 receptor results in a reduction in T_{FH} cells, along with diminished GC formation and selective reduction of high-affinity antibodies, whereas T cell-independent and low-affinity antibodies remain largely unchanged.

Defects in T_{FH} and GC function in *Il27ra*^{-/-} mice are T cell intrinsic

The observation that IL-27 directly promotes IL-21 expression by CD4⁺ T cells suggested that the defect in T_{FH} cell number and GC function in *Il27ra*^{-/-} mice could result from a T cell-intrinsic defect. However, other cells, including B cells, express the IL-27 receptor, and thus it remained formally possible that the T_{FH} defect in *Il27ra*^{-/-} mice was indirect. To discriminate these two possibilities, we generated

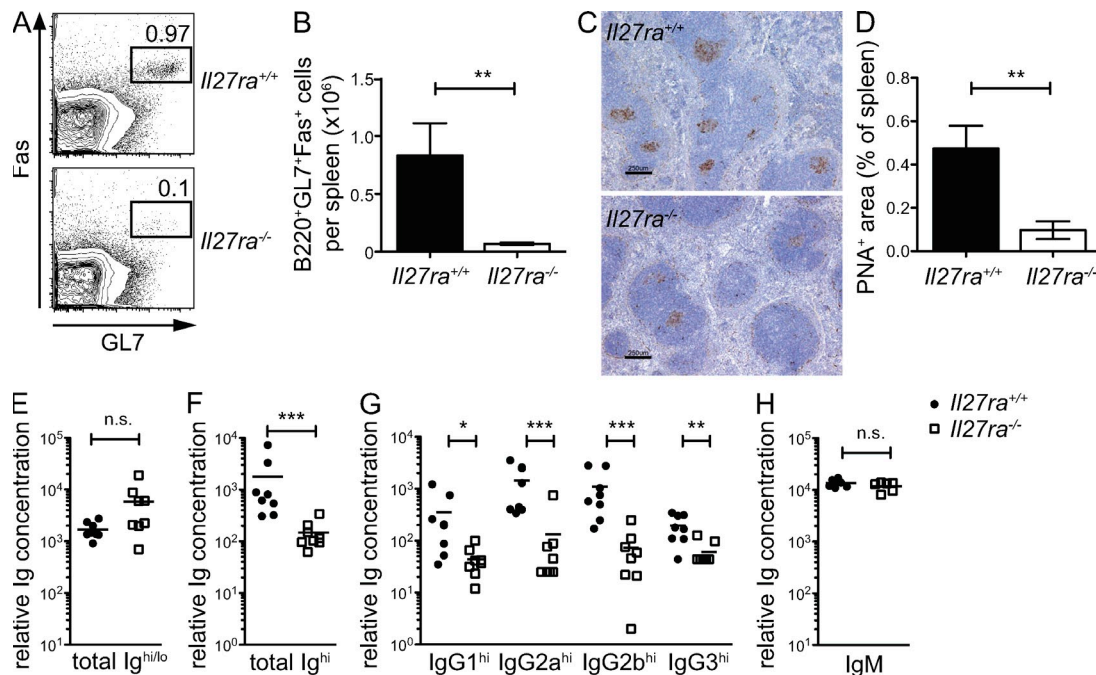


Figure 3. GC dysfunction in *Il27ra*^{-/-} mice. Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized twice with TNP-OVA in adjuvant and, 7 d after the second immunization, tissues and sera were collected for analysis. (A) Representative flow cytometric analysis for Fas and GL7 expression in the splenic B220⁺CD4⁻ cell gate. (B) The number of GL7⁺Fas⁺B220⁺CD4⁻ cells in the spleen of each mouse was calculated by multiplying the percentage obtained by flow cytometry by the total cell count per organ. The mean of at least six animals per group is given and error bars indicate SEM. (C) Representative spleen sections stained with PNA (brown) to detect GC. Bars, 250 μ m. (D) Slide scanning and image analysis software were used to quantify the PNA⁺ area in each of eight spleens per genotype. The mean percentage of PNA⁺ area is given and error bars indicate SEM. (E–H) ELISA using plates coated with 5 μ g/ml BSA-TNP₂₈ (E) or BSA-TNP₂ (F and G) for analysis of total anti-TNP and high-affinity anti-TNP antibodies, respectively, in the serum of mice immunized as in A. Anti-TNP antibodies were detected with either anti-mouse Ig (E and F) or antibodies against specific mouse Ig isotypes (G). (H) Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized with 100 μ g TNP-Ficoll i.p. and sera were collected 5 d later. Anti-TNP-IgM levels were assessed by ELISA as in E and detected using anti-mouse IgM antibodies. Relative anti-TNP antibody concentration is given for each mouse, and bars indicate the group mean where $n = 6$ –8. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$ (unpaired Student's t test). The data are indicative of three individual experiments.

mixed BM chimeras using *Il27ra*^{-/-} (CD45.2⁺ and Thy1.2⁺) BM mixed at a 1:1 ratio with congenic *Il27ra*^{+/+} (CD45.1⁺ and Thy1.2⁺) BM and transferred into lethally irradiated *Il27ra*^{+/+}CD45.1⁺Thy1.1⁺ hosts such that *Il27ra*^{-/-} donors, WT donors, and remnant WT host T cells could each be discriminated in the reconstituted chimeric mice. In such mice, WT and *Il27ra*^{-/-} T cells have the same exposure to the mixed population of WT and KO Ag-presenting cells and B cells. FACS analysis of the blood of reconstituted chimeric mice revealed a similar contribution of both genotypes to T and B cell compartments before immunization (Fig. S3, A and B), confirming that absence of the *Il27ra* does not impair repopulation. The chimeric mice were immunized twice, and 7 d after the second immunization the contributions of the WT and *Il27ra*^{-/-} cells to the total CD4⁺ and CD4⁺CXCR5⁺PD1⁺ T_{FH} cell populations were analyzed by flow cytometry. In the total CD4⁺ gate, WT cells contributed with somewhat increased frequency after immunization, with the WT/KO ratio being 1.644 ± 0.14 (mean \pm SEM) in the spleen and 1.366 ± 0.088 in the DLN (Fig. 4, A and B). However, WT T cells contributed highly disproportionately to the PD1⁺CXCR5⁺ T_{FH} gate, with a WT/KO ratio of 3.08 ± 0.3 in the spleen and 2.689 ± 0.26 in the DLN (Fig. 4, A and B). These data suggest that *Il27ra*^{-/-} T cells have an intrinsic defect in their follicular helper capacity, and this defect cannot be compensated for by the presence of WT APC and B cells. Because the T_{FH} cell compartment of the mixed chimeric mice was comprised mainly of WT cells, GC function was restored and the levels of high-affinity class-switched Ig were comparable with mice reconstituted with 100% WT cells (Fig. S3 C).

To conclusively rule out direct effects of IL-27 on B cells in this model, we constructed BM chimeras using B cell-deficient μ MT BM plus either WT or *Il27ra*^{-/-} BM (at a 80:20 ratio) such that all B cells in the resultant chimeras were derived from either the WT (control) or the *Il27ra*^{-/-} BM (a B cell-specific *Il27ra* deletion), whereas the majority of non-B cells were derived from WT BM. The absence of IL-27 signaling in the B cell compartment alone did not affect the development of T_{FH} cells in the spleen or DLN (Fig. 4, C and D). Although the μ MT chimeras generally produced lower amounts of high-affinity IgG compared with mice reconstituted with only WT BM, we noted no difference based on the *Il27ra* genotype of the B cells (Fig. 4 E). Thus, absence of IL-27 signaling in the B cell compartment alone affected neither the development of T_{FH} cells nor the production of high-affinity antibody.

An adoptive transfer system, in which OVA-specific *Il27ra*^{+/+} or *Il27ra*^{-/-} DO11.10 transgenic (Tg) CD4⁺ T cells were injected into either *Il27ra*^{+/+} or *Il27ra*^{-/-} hosts before immunization, further reinforced the T cell-intrinsic nature of the GC defect. Although complete removal of IL-27Ra from the system recapitulated the *Il27ra*^{-/-} phenotype, as expected, the percentage of T cells that expressed PD1 and CXCR5 was not significantly changed based on donor *Il27ra* genotype (Fig. S3 D). These data suggest that the differentiation of T_{FH} cells can occur in the absence of IL-27 signaling.

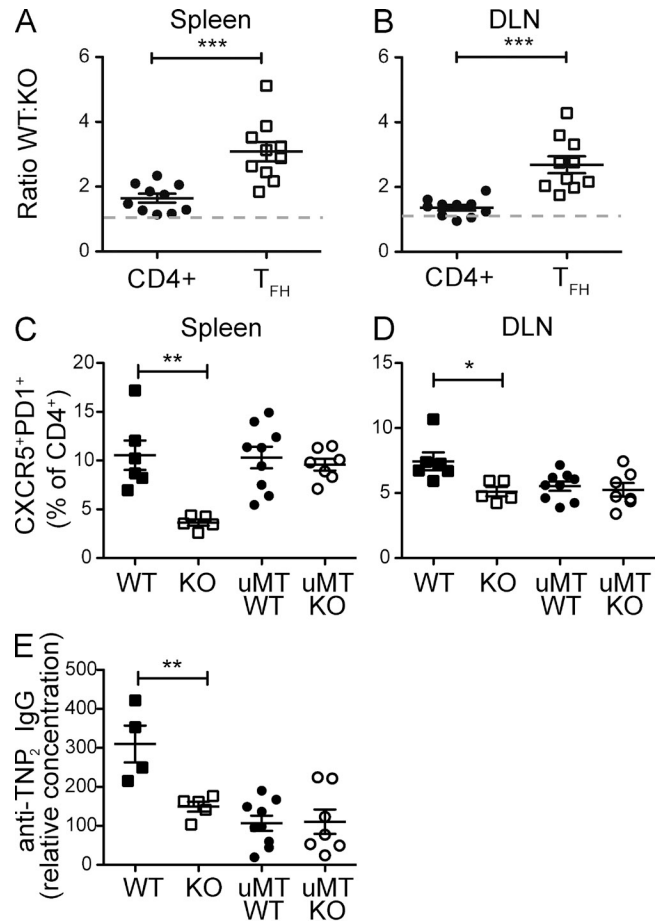
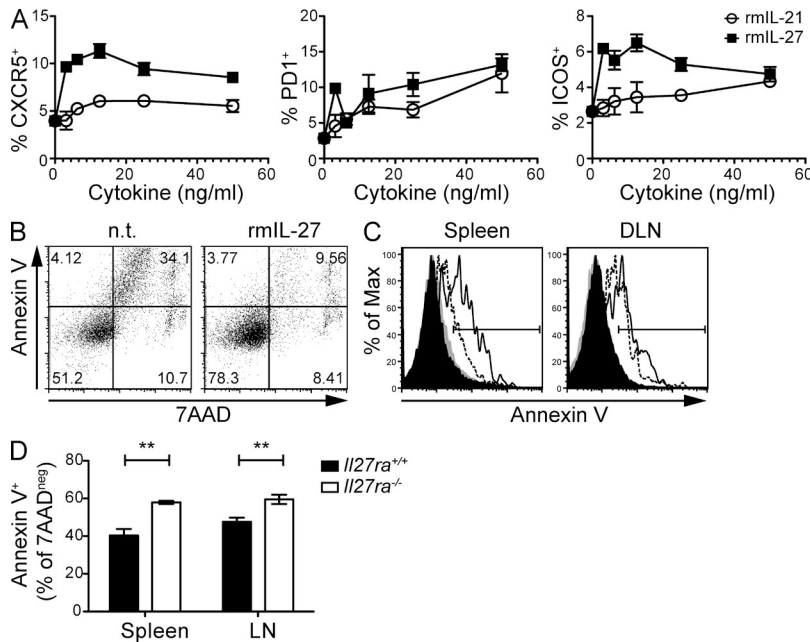


Figure 4. The GC defect in *Il27ra*^{-/-} mice is T cell intrinsic.

(A and B) WT (CD45.1⁺):*Il27ra*^{-/-} (CD45.2⁺) BM chimeric mice were immunized twice with TNP-OVA, as described, and tissue collected and assessed 7 d after the second injection. The ratio of CD45.1/CD45.2 cells is given for total CD4⁺ cells and CD4⁺CXCR5⁺PD1⁺ cells in the spleen (A) or DLN (B) for each of 10 chimeric animals. The gray dashed line indicates equivalency of WT and *Il27ra*^{-/-} cells (ratio of 1). (C and D) BM chimeric mice were reconstituted with BM from WT mice only, *Il27ra*^{-/-} mice only, WT and μ MT mice (1:4 ratio), or *Il27ra*^{-/-} and μ MT mice (1:4 ratio) and immunized twice as described. Tissue was collected 7 d after the second injection and the percentage of CXCR5⁺PD1⁺ cells in the CD4⁺B220⁻ gate in the spleen (C) and DLN (D) was determined by flow cytometry. (E) Serum high-affinity antibody was measured using ELISA plates coated with TNP₂-BSA and detected with anti-mouse IgG antibody. Data are given as a relative concentration using pooled serum from a control. Each set of data are indicative of two individual experiments. Bars indicate the mean \pm SEM. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$ (unpaired Student's *t* test).

However, lack of *Il27ra* on the transferred CD4⁺ T cells significantly reduced their recovery rates (Fig. S3 E), with a corresponding reduction in Fas⁺GL7⁺ GC B cells (Fig. S3 F), suggesting that IL-27 supports the survival and/or proliferation of the DO11.10⁺ cells in a T cell-dependent manner. The effect was most striking when DO11.10⁺.*Il27ra*^{+/+} and DO11.10⁺.*Il27ra*^{-/-} cells were transferred into *Il27ra*^{-/-} recipients (Fig. S3 E), which is consistent with the notion that the transferred DO11.10⁺.*Il27ra*^{+/+} T cells have a greater



competitive advantage when the endogenous recipient T cells, which also participate in the response, are unable to respond to IL-27. Alternatively, the increased recovery of DO11.10⁺.*Il27ra*^{+/+} T cells from KO hosts might be the result of increased IL-27 levels in mice that lack expression of the receptor, as is observed in other receptor-deficient mice (Cho et al., 2007). Differences between WT and *Il27ra*^{-/-} cells were not evident in the LN in the adoptive transfer system (unpublished data). Together, the BM chimera and adoptive transfer data described strongly support a T cell-intrinsic defect in GC responses in *Il27ra*^{-/-} mice.

IL-27 enhances the expression of T_{FH} cell markers and supports CD4⁺ T cell survival after Ag activation

Our data suggest that IL-27 is important for the differentiation and/or maintenance of T_{FH} cells. To test whether IL-27 stimulation of CD4⁺ T cells directly induces differentiation of T_{FH} cells, we performed in vitro experiments using OT-II Tg T cells. The cells were activated by OVA peptide in the presence of irradiated splenocytes and various concentrations of rmIL-27 or rmIL-21 for 5 d and then restimulated with anti-CD3 for 4 h (RNA expression) or 24 h (protein expression), as described by Nurieva et al. (2008). Under these conditions, the addition of rmIL-27 resulted in a concentration-dependent enhancement of the expression of CXCR5, PD1 and ICOS mRNA, and proteins on the cell surface (Fig. 5 and Fig. S4 A). In our hands, this enhancement was superior to rmIL-21. However, when purified CD4⁺ T cells were stimulated in the absence of accessory cells, ICOS, but not PD1 or CXCR5, was induced (not depicted; Pot et al., 2009). Moreover, IL-27 had little effect on Bcl-6 mRNA expression (Fig. S4 A), either under the conditions described in this paragraph or after a short-term primary stimulation (4 or 24 h; not depicted), suggesting that IL-27 did not induce full T_{FH}

Figure 5. IL-27 promotes survival of T_{FH} cells.

(A) OTII Tg CD4⁺ T cells were cultured with irradiated splenic APC plus 0.3 μM OVA₃₂₃₋₃₃₉ peptide under TH0 conditions and in the presence of either no additional cytokine or various concentrations of rmIL-21 or rmIL-27 for 5 d. After 5 d, CD4⁺ T cells were restimulated with anti-CD3 for 24 h and CXCR5, PD1, and ICOS expression were assessed by flow cytometry. The positive gate was determined based on histogram plots for each antibody and the mean percentage positive ± SD for triplicate experimental samples is shown. (B) DO11.10 tg.*rag2*^{-/-} splenocytes were activated with 0.03 μM OVA₃₂₃₋₃₃₉ in the presence or absence (n.t.) of rmIL-27 for 72 h, and viability was assessed by flow cytometry using annexin V and 7AAD staining. The CD4⁺B220⁻ gate is shown. (C and D) Viability of cells in groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice was assessed by flow cytometry 4 d after immunization with TNP-OVA in CFA. (C) Representative plots for annexin V staining in the CD4⁺B220⁻7AAD⁻ gate (*Il27ra*^{+/+}, black fill; *Il27ra*^{-/-}, gray fill) or CXCR5⁺PD1⁺CD4⁺B220⁻7AAD⁻ cell gate (*Il27ra*^{+/+}, dotted line; *Il27ra*^{-/-}, solid line) in the spleen or DLN. (D) The mean percentage of annexin V⁺ cells in the CD4⁺CXCR5⁺PD1⁺7AAD⁻ gate. Error bars indicate SEM with *n* = 5 mice per group. Each set of data are indicative of two individual experiments. **, *P* < 0.01 (unpaired Student's *t* test).

differentiation. In vitro stimulation of OTII TCR Tg T cells in the presence of rmIL-21 resulted in enhanced development of GC in adoptive hosts (Nurieva et al., 2008); however, stimulation with rmIL-27 was not able to enhance B helper activity under these conditions (Fig. S4 B). Although IL-21 is highly up-regulated by IL-27 stimulation, maximal expression appears to reach around 1 ng/ml (Fig. 1 B), which is markedly less than the 50 ng/ml IL-21 used in those in vitro T_{FH} differentiation systems. Moreover, other consequences of IL-27 signaling that diverge from that of IL-21 may also contribute to the differences observed. These data suggest that although IL-27 enhances the expression of T_{FH} effector molecules and cytokines, it is not able to do so in the absence of additional signals and is not sufficient to produce fully differentiated T_{FH} cells.

In adoptive transfer experiments, IL-27 signaling influenced the amount of Ag-specific CD4⁺ T cells that were present after immunization (Fig. S3, D–F). In in vitro culture systems, we noted that the addition of rmIL-27 to Ag-stimulated CD4⁺ T cells enhanced the survival of these strongly stimulated T cells (Fig. 5 B and Fig. S4 C). The survival advantage conferred to cells by the addition of rmIL-27 was not dependent on IL-21R signaling (Fig. S4 D). To investigate whether the altered cell survival observed in vitro was reflected by changes in T_{FH} cell survival in vivo, we examined the percentage of apoptotic (annexin V⁺) total CD4⁺ and T_{FH} cells in immunized *Il27ra*^{+/+} and *Il27ra*^{-/-} mice. We chose a time point of 4 d after immunization to examine early changes in viability and examined the percentage of annexin V (apoptotic) cells in the live (7-aminoactinomycin D [7AAD]⁻) gate. Although we found that the percentage of annexin V⁺ cell in

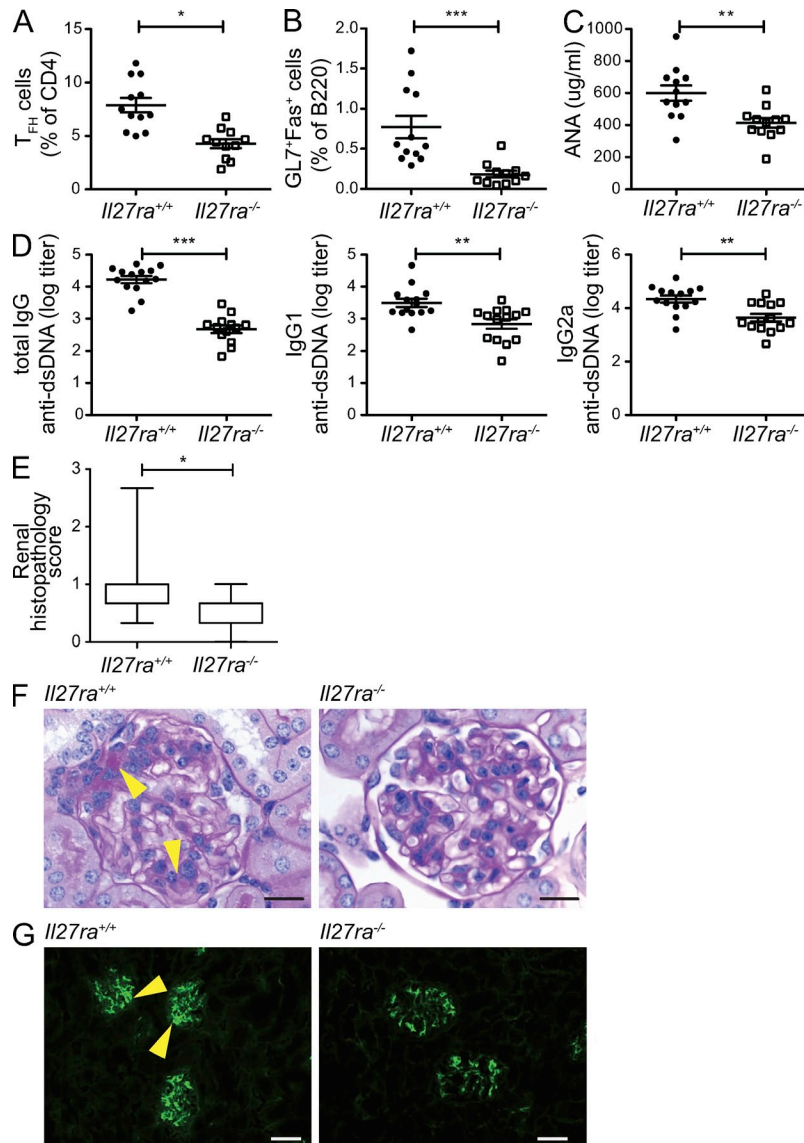


Figure 6. The deletion of *Il27ra* ameliorates pathology in the pristane-induced lupus model. A single 0.5-ml i.p. injection of pristane was administered to groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice. 42 wk after induction, mice were euthanized and examined for immune activation parameters and lesions consistent with SLE. (A and B) Flow cytometric analysis of splenocytes. The percentages of CXCR5⁺PD1⁺ (T_{FH}-like) cells in the CD4⁺B220⁻ gate (A) and the percentages of GL7⁺Fas⁺ (GC B cells) in the B220⁺CD4⁻ gate (B) were analyzed. (C) Serum anti-nuclear antibody (ANA). (D) Serum total IgG (left), IgG1 (middle), and IgG2a (right) anti-dsDNA antibody titers. In all graphs bars indicate mean \pm SEM. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$ (unpaired Student's *t* test). (E and F) Kidney sections were stained with PAS and the glomerulopathy scored by blinded observers. (E) Mean renal lesion scores are depicted as a box plot with max to min whiskers. *, $P < 0.05$ (unpaired Student's *t* test). (F) Representative sections. Arrowheads, lumpy accumulations of PAS-positive material. Bars, 20 μ m. (G) Immunofluorescence for IgG immune complex deposition in the kidney. Lumpy IgG deposits (arrowheads) correspond to PAS positive mesangial deposits. Bars, 50 μ m. Data represent one experiment with 16 animals per group.

the CD4⁺ T cell pool was similar between WT and *Il27ra*^{-/-} mice (Fig. 5 C), the percentage of annexin V⁺ cells in the PD1⁺CXCR5⁺ T_{FH} cell gate was significantly higher in *Il27ra*^{-/-} spleen and LN compared with WT (Fig. 5, C and D). Together, these data suggest that IL-27 is important for the survival of T_{FH} cells as well as enhancing the expression of T_{FH} cell effector proteins.

Loss of IL-27 signaling ameliorates disease in a murine lupus model

Our data suggest that IL-27 might play a pathogenic role in diseases that are characterized by affinity-matured autoantibody production such as systemic lupus erythematosus (SLE; Wellmann et al., 2005; Mietzner et al., 2008). To test this hypothesis, we used a model in which a single i.p. injection of tetramethylpentadecane (TMPD; commonly known as pristane) results in an SLE-like syndrome characterized by the development of a variety of autoantibodies,

polyclonal hypergammaglobulinemia, and glomerulonephritis (Satoh and Reeves, 1994; Reeves et al., 2009). The IgG hypergammaglobulinemia and the development of IgG1 and IgG2a anti-nuclear antibodies appear to be dependent on T cell help (Nacionales et al., 2009). Because genetic factors influence susceptibility to TMPD-induced lupus, we used *Il27ra*^{-/-} mice on the BALB/c background to study the importance of IL-27 in the development of disease in this model. 42 wk after TMPD injection, the *Il27ra*^{-/-} displayed significantly reduced populations of T_{FH} and GC B cells (Fig. 6, A and B), along

with significantly lower levels of serum anti-nuclear (Fig. 6 C) and anti-dsDNA IgG (Fig. 6 D) antibodies compared with WT controls. Although a global reduction in IgG2a production in *Il27ra*^{-/-} mice has been noted previously (Chen et al., 2000; Miyazaki et al., 2005), the reduction in anti-dsDNA antibodies was not confined to this isotype because IgG1 anti-dsDNA antibody was also significantly lower in *Il27ra*^{-/-} serum (Fig. 6 D). Consistent with the reduction in autoantibody production, histological analysis revealed a lower mean renal histopathology score in the *Il27ra*^{-/-} mice (Fig. 6, E and F) and a reduction in histological evidence of IgG immune complex deposition in the kidney (Fig. 6 G). In both groups, renal lesions consistent with SLE are mild, although there was greater individual variability with occurrence of more severe lesions in the WT group. These data demonstrate that the defects observed in GC function in the absence of IL-27 signaling are relevant in the disease setting.

DISCUSSION

The development of T_{FH} cells is not fully understood and represents a topic of intense current interest. Our data position IL-27 as a new factor that is critical for T_{FH} cell function. Stimulation of T cells with IL-27 promoted IL-21 production and expression of the T_{FH} hallmark proteins PD-1, CXCR5, and ICOS. In *Il27ra*^{-/-} mice, we observed a reduction in IL-21 production, fewer T_{FH} cells, reduced GC formation, and a reduction in high-affinity immunoglobulin production, indicating that IL-27 signaling is required for efficient T cell-dependent humoral immune responses. A functional consequence of IL-27R α ablation was partial protection in the pristane-induced lupus model, which is characterized by active GC development, production of multiple autoantibodies, and renal pathology. Collectively, these data strongly suggest that IL-27 is required for normal and pathogenic GC responses in vivo.

IL-27 also induced IL-21 expression by human CD4⁺ T cells, suggesting that the IL-27–IL-21 axis is conserved across species. However, our results differ from recent observations by Schmitt et al. (2009), who observed that rhIL-12, but not rhIL-27, induced IL-21 expression by human PBMC. The reason for this discrepancy remains unresolved at this time but may result from differences in the experimental setup and the strength of TCR stimulation, which we observed to be proportional to IL-21 production (unpublished data). Our data also suggest that although IL-12 and IL-27 can independently induce IL-21 (Fig. 1 C), they can also act in concert to induce maximal production of this cytokine (Fig. S1, B and C). Mechanistically, IL-21 induction was independent of STAT1 in murine T cells (Fig. S1 D) but failed to occur in STAT3-deficient T cells from human AD-HIES patients (Fig. 1 D).

IL-27 was unable to induce the functional T_{FH} phenotype in an in vitro system. This was surprising because IL-27 induced IL-21 production, which by itself is sufficient to induce functional T_{FH} cells (Nurieva et al., 2008). However, IL-21 levels peak only at 48 h and ~ 1 ng/ml when induced by IL-27, whereas direct stimulation with recombinant IL-21 was conducted at 50 ng/ml and affected cells from the outset (Nurieva et al., 2008). Consequently, these two in vitro stimulation experiments cannot be directly compared. Moreover, isolated stimulation with a single recombinant cytokine in vitro falls short of mimicking the much more complex stimulation environment in vivo, where IL-27 is clearly required for a functional T_{FH} response.

In addition to inducing hallmark T_{FH} genes, our data also suggest that IL-27 promotes the survival of T_{FH} cells. T_{FH} cells represent a highly activated population, which express high levels of Fas and low levels of IL-7R (Ma et al., 2009) and are exquisitely sensitive to activation-induced cell death (Marinova et al., 2006). In T cell transfer experiments, DO11.10⁺rag2^{-/-} T cells assumed the T_{FH} phenotype readily and independently of IL-27 signaling (Fig. S3 D). This is perhaps not surprising because the DO11.10⁺ cells uniformly express a high-affinity Ag-specific TCR and because Ag

affinity controls T_{FH} differentiation (Fazilleau et al., 2009). However, DO11.10⁺rag2^{-/-} T cells that lacked *Il27ra* were recovered at diminished numbers (Fig. S3 E), suggesting that IL-27 is important for the maintenance of T_{FH} cells. This hypothesis is supported by our observations that recombinant IL-27 enhances the survival of newly activated CD4⁺ T cells in vitro and that T_{FH} cells undergo increased apoptosis in vivo when *Il27ra* is genetically ablated. Although the cells studied in vitro were not fully differentiated T_{FH} cells, the death of recently activated CD4⁺ T cells would likely influence the number of cells that subsequently become T_{FH} cells. IL-27 also induced ICOS, and ICOS signals have been shown to contribute to the survival and expansion of Ag-specific T cells in an immunization system similar to the one used in this paper (Burmeister et al., 2008). Thus, although the relative contributions remain to be determined, ICOS induction, IL-21 production (Nurieva et al., 2008), and other IL-27-elicited effects may all contribute to the enhanced survival of T_{FH} cells.

Somewhat surprisingly, the reduction in T_{FH} cells was more pronounced in spleen compared with the DLN (Fig. 2 and Fig. S2). The transfer of TCR Tg cells confirmed that Ag-specific responses occur in the spleen after immunization in the flank (Fig. S4, C and D). Because inhibition of lymphocyte migration using FTY720 administration did not prevent the tissue-specific effect (Fig. S2 C), we favor a hypothesis in which the proximity of the LN to the site of injection results in increased local exposure to Ag and inflammatory cytokines (Fu et al., 2006). Strong Ag signals promote T_{FH} differentiation (Vogelzang et al., 2008; Fazilleau et al., 2009; Deenick et al., 2010), thus, the dependence on IL-27 signals may be reduced. Despite these possible compensatory effects close to the site of immunization, it is clear that IL-27 signaling is not redundant because significant reductions in T_{FH} cells in the DLN and reduced serum high-affinity antibodies remain.

Our data suggest that the T_{FH} -promoting effect of IL-27 is mediated through direct action on T cells. First, ICOS and IL-21 production were induced in T cell-only systems in vitro (Fig. 1; Pot et al., 2009). Furthermore, in chimeric animals that harbor both WT and *Il27ra*^{-/-} T cells, the *Il27ra*^{-/-} T cells are selectively underrepresented in the T_{FH} pool (Fig. 4, A and B). In contrast, no differences in the T_{FH} response or high-affinity Ig levels were noted when *Il27ra* was selectively ablated on B cells (Fig. 4, C and D), which are known to be functionally responsive to IL-27 in vitro (Yoshimoto et al., 2004; Larousserie et al., 2006). These data do not preclude the possibility that IL-27 influences certain minor aspects of the T cell-dependent humoral response, such as IgG2a class switching (Yoshimoto et al., 2004), through direct activities on B cells. Although the IL-27R is also expressed on myeloid cells, transfer of *Il27ra*^{+/+} DO11.10⁺rag2^{-/-} T cells into *Il27ra*^{-/-} recipients did not result in a significant decrease of T_{FH} differentiation (Fig. S3, D–F), suggesting that IL-27 signaling into myeloid cells plays no major role in this context.

IL-27 is induced by type I IFN (Pirhonen et al., 2007; Remoli et al., 2007), and *Ifnar1*^{-/-} mice display a similar phenotype to *Il27ra*^{-/-} mice with respect to both loss of T_{HH} function (Cucak et al., 2009) and protection from pristane-induced lupus (Nacionales et al., 2007), suggesting that IL-27 is an important downstream effector of type 1 IFN. In human SLE patients, a type 1 IFN signature is readily detectable, and antagonists are currently in clinical trials for the treatment of this disease (Yao et al., 2009; Niewold et al., 2010). Our data suggests that IL-27 might mediate the pathogenic effect of type I IFN and that therapeutic targeting of IL-27 may also be efficacious in SLE and other disorders characterized by excessive GC formation and high-affinity autoantibody production.

MATERIALS AND METHODS

Mice, cells, and reagents. *Il27ra*^{+/+} and *Il27ra*^{-/-} (Chen et al., 2000) mice (C57BL/6 background, *n* > 33 unless otherwise stated or, where indicated, BALB/c background, *n* > 11), OT-II TCR Tg (C57BL/6), and *Il27ra*. DO11.10 TCR Tg/rag2-deficient mice (*Il27ra*^{-/-}.DO11.10tg.rag2^{-/-} or *Il27ra*^{+/+}.DO11.10tg.rag2^{-/-} on the BALB/c background) were bred in a specific pathogen-free facility at The Garvan Institute or Genentech. *Stat1*^{-/-} mice (129Sv/Ev background) and 129Sv/Ev control mice were purchased from Taconic. The TCM (triple congenic) mice, Igh^aB6-CD45.1-Cross-B6.SJL, which express congenic markers at the CD45.1, Igh^a and Thy1.1 loci on the C57BL/6 background, and μMT mice (*Igh-6^{tm1Cgn}* targeted mutation; C57BL/6 background) were bred in a specific pathogen-free facility at Genentech. The rag2^{-/-} animals were purchased from The Jackson Laboratory. All live animal experiments were approved by the Genentech Institutional Animal Care and Use Committee or The Garvan Institute/St. Vincent's Animal Experimentation Ethics Committee. Human PBMC buffy coats were obtained from the Australian Red Cross Blood Service, and patients with the clinical diagnosis of AD-HIES as a result of mutations in *STAT3* were recruited from Immunology Clinics in Canberra and Sydney, Australia. All human experiments were approved by jurisdictional ethics committees in Canberra and Sydney. Unless otherwise indicated, all cytokines were purchased from R&D Systems and all antibodies were from BD. Cycloheximide was purchased from Sigma-Aldrich. FTY720 (Cayman Chemical) was reconstituted in DMSO and injected i.p. in H₂O. IL21R-Fc was produced at the Garvan Institute as previously described (Herber et al., 2007).

Isolation of lymphocyte subsets. Unless otherwise indicated, primary mouse CD4⁺ T cells were isolated by magnetic depletion using MACS kits (Miltenyi Biotec) according to the manufacturer's instructions. The purity ranged from 90 to 95%. Where indicated, FACS sorting was performed using a FACSaria instrument (BD) to obtain specific cell populations of >99% purity. After immunization, splenic leukocyte populations were isolated as follows: CD4⁺ (CD4⁺B220⁻), T_{HH} (CD4⁺CXCR5⁺PD1⁺), non-GC B (B220⁺CD38⁺), GC B (B220⁺CD38^{lo}), FDC (B220⁺CD35^{hi}CD32⁺), CD11b⁺ (CD11b⁺CD11c⁻B220⁻), and CD11c⁺ (CD11c⁺CD11b⁻B220⁻). For cell culture systems that required Ag-presenting cells, splenocyte samples were magnetically depleted of T cells with anti-CD90 (Thy-1.2) MACS MicroBeads (Miltenyi Biotec) and irradiated with a dose of 2,600 rads.

In vitro T cell stimulation. Primary mouse cells were cultivated in Iscove's modified Dulbecco's medium (Invitrogen) supplemented with 10% (volume/volume) heat-inactivated FBS (HyClone PerBio), 1 mM L-glutamine, 1% (volume/volume) penicillin and streptomycin (Invitrogen), and 55 μM 2-mercaptoethanol (MP Biomedicals). Splenic CD4⁺ T cells from C57BL/6, *Il27ra*^{-/-}, 129, or *Stat1*^{-/-} mice were activated for the indicated times in plates coated with 5 μg/ml of anti-CD3 and in the presence of 1 μg/ml anti-CD28. Unfractionated DO11.10tg.rag2^{-/-} or OTIItg splenocytes

were stimulated with OVA₃₂₃₋₃₃₉ peptide at the indicated concentrations. TH0 conditions consist of 5 μg/ml each of hamster anti-mouse IFN-γ (H22), rat anti-mouse IL-4 (BVD4-1D11), and rat anti-mouse IL-12 (C15.6) and 100 ng/ml TGFβRII-Fc (R&D Systems). Where indicated, rmIL-27 was added at a concentration of 20 ng/ml.

Human naive CD4⁺ T cells were FACS purified from tonsil cell or PBMC preparations based on the CD4⁺CD45RA⁺CXCR5⁻ phenotype. Cells were labeled with CFSE and stimulated with T cell activation and expansion beads (Miltenyi Biotec) at a bead/cell ratio of 2:1 in the presence of no additional cytokine, 20 ng/ml rhIL-12, 20 ng/ml rhIL-23, or 50 ng/ml rhIL-27 for 5 d. Where irradiated splenocytes were used as APC, total splenocytes were irradiated with 2,000 rads x-ray irradiation.

Immunization. For T cell-dependent immunization, groups of age- and sex-matched mice were immunized with TNP₁₄-OVA (30 μg/mouse) or OVA (100 μg/mouse), as indicated, and emulsified in 100 μl CFA (Sigma-Aldrich) by subcutaneous injection into the flank. Where a second immunization was required, this was performed 21 d after the initial injection using the same dose of Ag emulsified in IFA (Sigma-Aldrich) to a volume of 100 μl/mouse and injected subcutaneously into the alternate flank.

For T cell-independent immunization, groups of six mice per genotype were immunized i.p. with 100 μg TNP-aminoethylcarboxymethyl-Ficoll in PBS. Serum was harvested 5 d later.

Flow cytometric analysis. Cells were treated with Fc blocking Abs (anti-CD16/32 2.4G2) and then surface stained with the specified antibodies. To measure cell viability, cells were stained with FITC-conjugated annexin V and 7-AAD according to the manufacturer's instructions (BD). Viable cells exclude both stains. IL-21 expression in human T cells was assessed by intracellular staining using Alexa Fluor 647-labeled anti-human IL-21 (eBioscience; clone 3A3-N2). Samples were analyzed using a FACSCanto II or LSR II (BD) and data analyzed using Flow Jo software (Tree Star, Inc.). Contour profiles are presented as 5% probability contours with outliers.

ELISA. IL-21 was detected in culture supernatants using the mouse IL-21 DuoSet ELISA (R&D Systems) as per the manufacturer's instructions. To measure the relative amounts of TNP-specific antibodies in mouse serum, plates were coated with 5 μg/ml TNP₂-BSA or TNP₂₈-BSA (Biosearch Technology) overnight at 4°C. TNP-specific ELISA was otherwise performed as previously described (Roes and Rajewsky, 1993). To standardize and quantify relative amounts of TNP-specific Ig responses, all experimental samples were compared with a standardized dilution of pooled serum obtained from *Il27ra*^{-/-} mice immunized with TNP-OVA. This standard was given the arbitrary concentration of 100. In the case of IgM, an anti-TNP monoclonal (BD; clone G155-228) was used as a standard control.

For serum autoantibody measurement, mouse ANA, anti-Sm, and anti-nRNP antibodies were measured using ELISA kits from Alpha Diagnostic International, as per the manufacturer's instructions. Anti-dsDNA antibodies were measured by coating ELISA plates with 0.01% p-L-Lysine (Sigma-Aldrich) for 1 h at room temperature and then 2.5 μg/ml of calf thymus DNA (Sigma-Aldrich) in PBS at 4°C overnight. After blocking with PBS, 0.5% BSA + 10 ppm Procline, pH 7.4, for 1 h at room temperature, serially diluted mouse serum was added and incubated at room temperature for 2 h. Antibody was detected using 500 ng/ml biotinylated rat anti-mouse IgG, IgG1, or IgG2a (BD) for 1 h at room temperature, followed by HRP-conjugated streptavidin and visualized using TMB substrate. NZBF1 serum was used as a positive control.

BM chimeras. For mixed BM chimeras, triple congenic mice (CD45.1, Thy1.1, and Igh^a) were lethally irradiated (¹³⁷Cs; gamma source, 1,150 rad) and reconstituted with equal numbers of BM cells from *Il27ra*^{+/+} (CD45.1 and Thy1.2) and *Il27ra*^{-/-} (CD45.2 and Thy1.2) mice by i.v. injection. After 8 wk of reconstitution, the mice were bled to assess reconstitution by flow cytometry and immunized as described in Immunization.

For μ MT chimeras, Rag2^{-/-} mice were lethally irradiated (¹³⁷Cs; gamma source, 1100 rad) and reconstituted with BM from B cell-deficient μ MT mice plus either WT or IL-27ra^{-/-} BM (at a 80:20 ratio), such that all B cells in the resultant chimeras were derived from either the WT (control) or the IL27ra^{-/-} BM (a B cell-specific IL27ra deletion), with the majority of non-B cells derived from WT BM. After 8 wk of reconstitution, the mice were bled to assess reconstitution by flow cytometry and immunized as described in Immunization.

Adoptive transfer. CD4⁺ T cells from either IL27ra^{+/+} or ^{-/-} DO11.10tg. rag2^{-/-} were isolated by magnetic sorting and 10⁶ CD4⁺ cells were transferred to either IL27ra^{+/+} or IL27ra^{-/-} hosts (on a BALB/c genetic background) via tail vein injection. The next day, the mice were immunized with OVA in CFA/IFA as described in Immunization and tissue harvested 7 d after the second immunization. DO11.10 TCR Tg cells were detected using directly fluorochrome-labeled clonotypic KJ1-26 antibody (BD).

Statistical analysis. Data were analyzed with Prism software (GraphPad Software, Inc.) to calculate an unpaired two-way Student's *t* test.

RT-PCR. Total RNA from FACS-sorted or cultured cells was isolated with the RNeasy kit using on-column DNase I digestion (QIAGEN). Taqman quantitative RT-PCR was performed according to the manufacturer's instructions (Applied Biosystems). A Lightcycler480 (Roche) instrument was used in the case of human samples. For each sample, triplicate test reactions and a control reaction lacking reverse transcription were analyzed for expression of the gene of interest and results were normalized to those of the housekeeping ribosomal protein L19 mRNA or hGAPDH. Arbitrary units given are the fold change relative to RPL19 (mouse) or GAPDH (human) and multiplied by 1,000. Primer sequences for each target are provided in Table S1.

Histochemical analysis. For GC quantification, spleens were fixed in 10% neutral buffered formalin and subsequently embedded in paraffin. To detect GC, 5- μ m sections were stained with biotin-conjugated PNA followed by visualization with HRP-linked streptavidin and diaminobenzidine. The sections were lightly counterstained with hematoxylin. Slides were scanned on a NanoZoomer digital pathology system (Olympus), and Matlab image analysis software (The MathWorks) was used to quantitate the percentage of each spleen that was positive for PNA staining.

For kidney glomerulopathy, tissue collected at necropsy was fixed in formalin, paraffin embedded, sectioned at <4 μ m and subjected to Periodic acid-Schiff stain (PAS). Tissue was evaluated by blinded observers using arbitrary severity scores on scales of 0–3 for interstitial nephritis, arteritis/periarthritis and glomerulopathy. Groupwise comparison of mean lesion severity scores was analyzed by ANOVA. For kidney and immune complex deposition, tissue collected at necropsy was snap frozen in OCT, sectioned, and stained by direct immunofluorescence using 10 μ g/ml of goat anti-mouse IgG H⁺L FITC (Jackson ImmunoResearch Laboratories) for 60 min at room temperature.

Pristane-induced lupus. SLE was induced by a single i.p. injection of 0.5 ml pristane (2,6,10,14-tetramethylpentadecane) to groups of IL27ra^{+/+} and IL27ra^{-/-} mice on the BALB/c background. Mice were sacrificed 42 wk after induction and examined for lesions consistent with SLE by histology, the presence of autoantibodies in the serum by ELISA, and the presence of T_{HH} and GCB cells in the spleen by flow cytometry.

Online supplemental material. Fig. S1 shows a requirement for protein translation, but not STAT1 signaling, for IL-21 induction by IL-27, that IL-27 induces IL12RB2 mRNA expression by human T cells, and that IL-12 and IL-27 can additively promote IL-21 production. Fig. S2 further defines the surface marker expression of T_{HH} cells, which are reduced in IL27ra^{-/-} mice, and shows that lack of T_{HH} cells is still evident after inhibition of lymphocyte migration and that blockade of IL-21 signaling does not further

suppress T_{HH} generation in IL27ra^{-/-} mice. Fig. S3 shows reconstitution of lymphocyte populations and serum levels of high-affinity antibody in BM chimeric mice as described in Fig. 4, and it shows T_{HH} and GC B cell generation when WT or IL27ra^{-/-} TCR Tg T cells are transferred to WT or IL27ra^{-/-} hosts. Fig. S4 details the effect of IL-27 on T_{HH} differentiation and T cell survival in vitro. Table S1 is a list of primers and probes used for RT-PCR. Online supplemental material is available at <http://www.jem.org/cgi/content/full/jem.20100064/DC1>.

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SUPPLEMENTAL MATERIAL

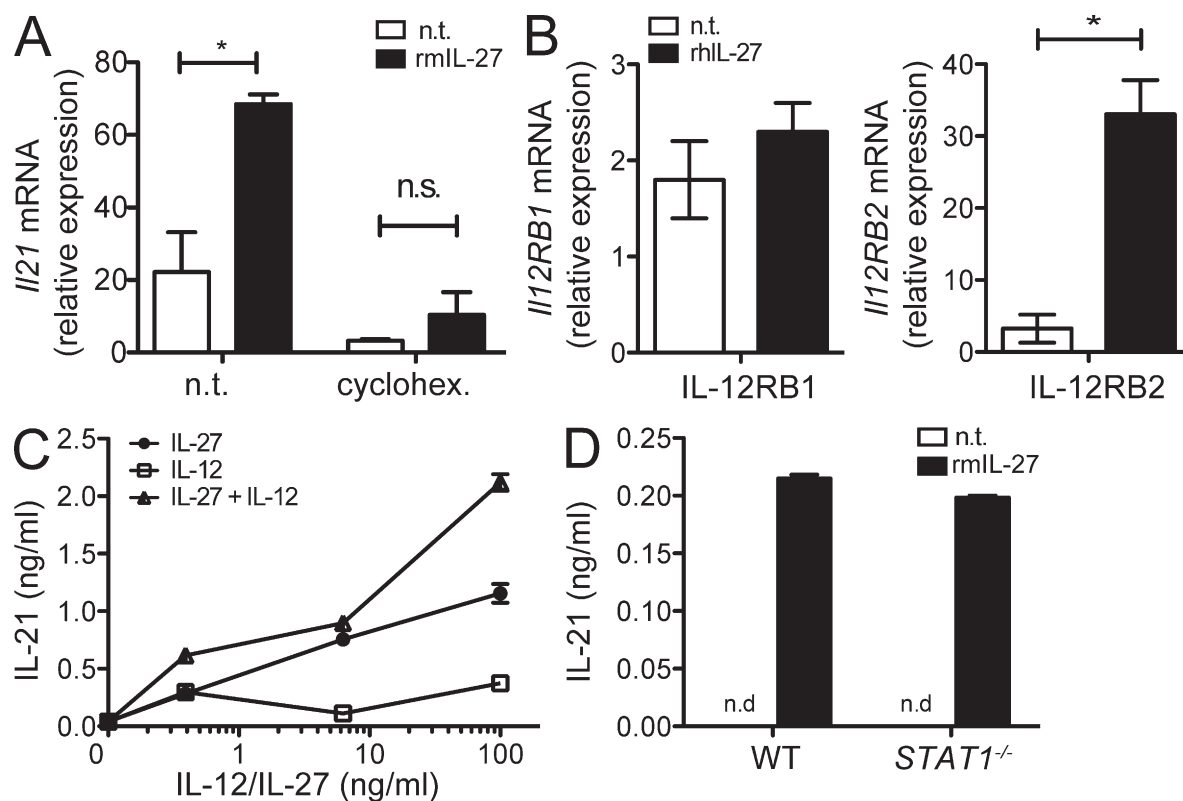
Batten et al., <http://www.jem.org/cgi/content/full/jem.20100064/DC1>

Figure S1. Regulation of IL-21 by IL-27. CD4⁺ T cells from C57BL/6 mice were stimulated with plate-bound anti-CD3 and soluble anti-CD28 under TH0 polarizing conditions in the presence (filled symbols) or absence (open symbols) of rmIL-27 and in the presence or absence of cycloheximide for 5 h. The mean of triplicate experimental samples is given \pm SD. Data are representative of three independent experiments. (B) Naive CD4⁺ T cells isolated from human PBMC were stimulated with T cell activation beads in the presence (filled bars) or absence (open bars) of rhIL-27 for 3 d. The expression of *IL12RB1* (left) and *IL-12RB2* (right) mRNA expression was measured by real-time RT-PCR and is given relative to human *gapdh*. Combined data from three independent donors and experiments is shown \pm SD. (C) CD4⁺ T cells enriched from C57BL/6 splenocytes were stimulated with plate-bound anti-CD3 and soluble anti-CD28 in the presence of varying concentrations of rmIL-27, rmIL-12, or both rmIL-27 and rmIL-12 for 72 h. IL-21 in the culture supernatant was measured by ELISA and the mean \pm SD of triplicate experimental samples is shown. (D) CD4⁺ T cells from STAT1^{+/+} (SvEv) or STAT1^{-/-} mice were stimulated as in A for 72 h. IL-21 in the culture supernatant was measured by ELISA. Data in D are representative of two experiments. n.t., no treatment; n.d., not detectable. *, $P < 0.05$ (unpaired Student's *t* test). Bars indicate mean \pm SEM.

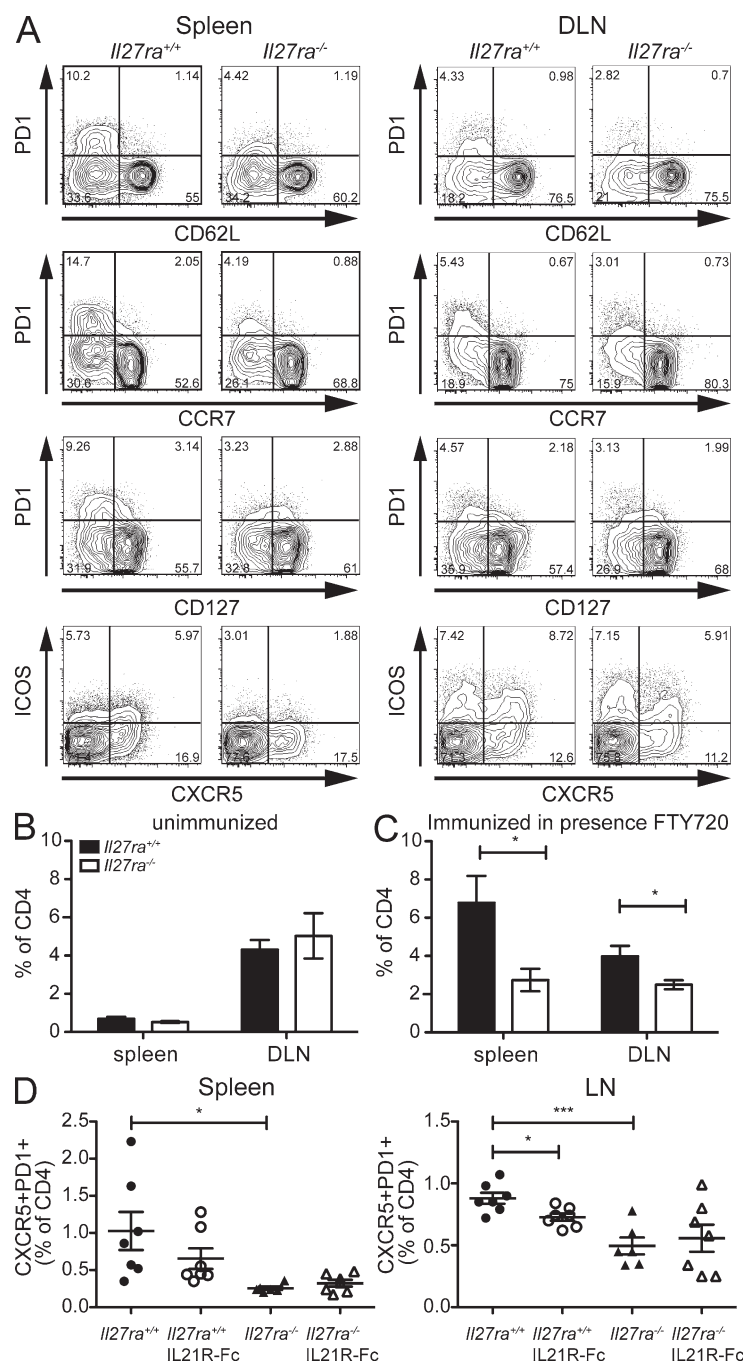


Figure S2. Detailed analysis of T_H cell reduction in *Il27ra*^{-/-} mice. (A) Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized twice with TNP-OVA in adjuvant and, 7 d after the second immunization, tissue was collected for analysis. Representative flow cytometric analysis for T_H marker expression in the spleen (left) and DLN (right). For all plots the CD4⁺B220⁻ gate is shown. (B) The percentage of CXCR5⁺PD1⁺ cells in the CD4⁺B220⁻ gate in the spleen and DLN of unimmunized age-matched *Il27ra*^{+/+} and *Il27ra*^{-/-} mice. Data are representative of three independent experiments. (C) Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized once with TNP-OVA in CFA and FTY720 was administered i.p. every second day beginning on the day of immunization. Efficacy of the FTY720 treatment was confirmed by a >90% reduction in normal T cell number in the blood. 7 d after the immunization, tissue was collected for analysis. The percentage of CXCR5⁺PD1⁺ cells in the CD4⁺B220⁻ gate in the spleen and DLN is given. (D) Groups of *Il27ra*^{+/+} and *Il27ra*^{-/-} mice were immunized once with TNP-OVA in CFA. Where indicated, 20 μ g/mouse of IL-21R-Fc was administered i.v. every second day beginning 1 d before immunization. 7 d after immunization, tissue was collected for analysis. The percentage of CXCR5⁺PD1⁺ cells in the CD4⁺B220⁻ gate in the spleen and DLN is given. For C and D, a single experiment with seven animals per group is represented and error bars indicate SEM. *, $P < 0.05$; ***, $P < 0.001$ (unpaired Student's *t* test). Bars indicate mean \pm SEM.

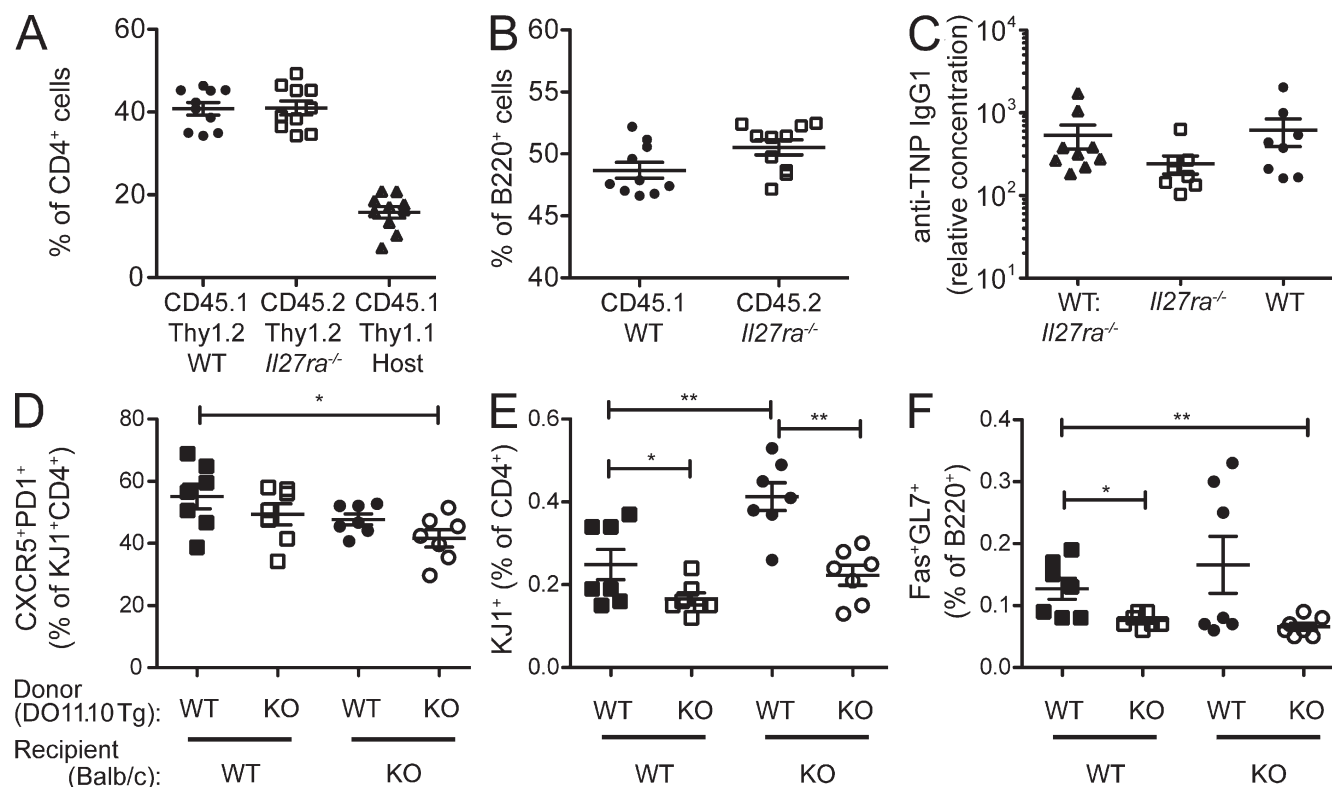


Figure S3. T cell-intrinsic defect in GC activity in *Il27ra*^{-/-} mice (A–C) TCM mice (CD45.1 and Thy1.1) were lethally irradiated and reconstituted with a 50:50% mix of BM from WT (CD45.1) and *Il27ra*^{-/-} (CD45.2) mice (A and B). 6 wk after BM transfer, the mice were bled to assess reconstitution by flow cytometry. (A) The percentage of WT (filled circles), *Il27ra*^{-/-} (open squares), and host (filled triangles) CD4⁺ T cells in the mixed chimeras. (B) The percentage of WT (CD45.1⁺ host plus donor; filled circles) and *Il27ra*^{-/-} (open squares) in the B220⁺ B cell gate in the mixed chimeras. (C) Serum IgG1 levels as determined by ELISA using TNP₂-BSA coating in the mixed chimera (triangles) compared with chimeric mice reconstituted with only *Il27ra*^{-/-} (squares) or *Il27ra*^{+/+} (WT; circles) BM. (D–F) DO11.10Tg.*Il27ra*^{+/+} (WT) or DO11.10Tg.*Il27ra*^{-/-} (KO) CD4⁺ T cells were adoptively transferred into either *Il27ra*^{+/+} (WT) or *Il27ra*^{-/-} (KO) hosts (as indicated) on the BALB/c background. The mice were immunized with OVA in CFA and the response analyzed 7 d later. Flow cytometric analysis was used to determine the percentage of CD4⁺KJ1⁺ donor cells that became CXCR5⁺PD1⁺ (D), KJ1⁺ donor cells as a percentage of total CD4⁺ cells (E), and the percentage of Fas⁺GL7⁺ GC B cells in the spleen (F). Bars indicate the mean ± SEM. Each dataset is representative of two independent experiments. *, P < 0.05; **, P < 0.01 (unpaired Student's *t* test).

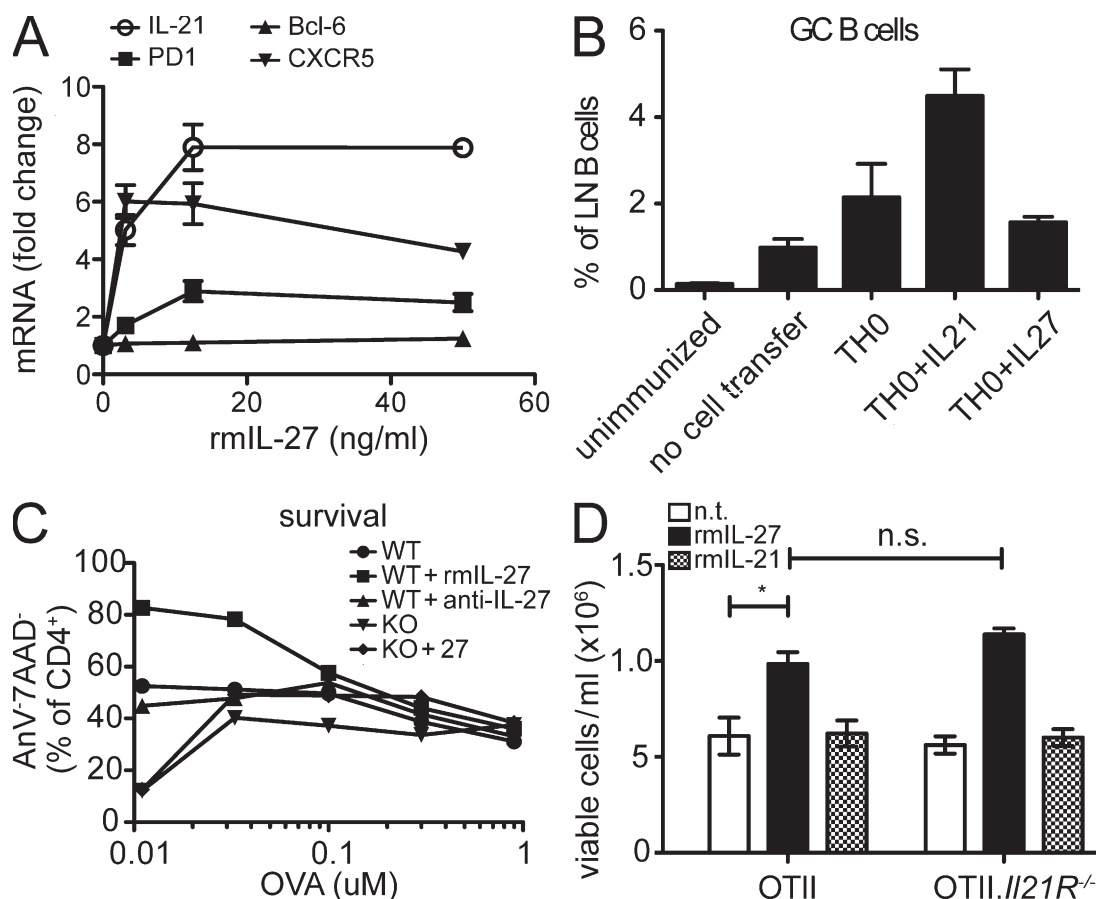


Figure S4. IL-27 does not promote T_{FH} differentiation. (A) OTII Tg $CD4^+$ T cells were cultured with irradiated splenic APC plus $0.3 \mu M$ OVA₃₂₃₋₃₃₉ peptide under TH0 conditions and in the presence of either no additional cytokine or various concentrations of rmlL-27 for 5 d. After 5 d, $CD4^+$ T cells were restimulated with anti-CD3 for 4 h and IL-21, PD1, Bcl-6, and CXCR5 mRNA levels were assessed by real-time RT-PCR. mRNA levels for each probe set were normalized to the housekeeping gene Rpl19 and the fold increase relative to samples stimulated in the absence of rmlL-27 are given. The mean \pm SEM of triplicate experimental samples is shown and data are representative of two experiments. (B) Thy1.1⁺ OTII TCR Tg $CD4^+$ T cells were isolated by magnetic purification and cultured with irradiated splenic APC plus OVA₃₂₃₋₃₃₉ peptide under TH0 conditions alone (blocking antibodies against IFN- γ and IL-4 and TGFBR11-Fc) or with the addition of 50 ng/ml rmlL-21 or 50 ng/ml rmlL-27 for 5 d. Cells were then adoptively transferred to naive Thy1.2 congenic hosts ($n = 4-8$ per group) before recipient mice were subcutaneously immunized with 100 μg OVA in IFA. Two additional control groups were included that did not receive cell transfers; one group was immunized as described, whereas the other group remained unimmunized. 7 d after immunization, differentiation of GC B cells in the LN were assessed by flow cytometry. The graph shows the mean percentage of GL7⁺Fas⁺B220⁺ cells in the DLN. Error bars indicate SEM. (C) DO11.10tg.rag2^{-/-} or DO11.10tg.rag2^{-/-}.II27ra^{-/-} splenocytes were activated with various concentrations of OVA₃₂₃₋₃₃₉ in the presence or absence of 20 ng/ml rmlL-27 or 10 μg /ml of anti-IL-27 for 72 h and the percentage of AnV-neg and 7AAD-neg (viable) cells in the $CD4^+$ gate was determined by flow cytometry. Data are representative of three independent experiments. (D) OTII.II21R^{+/+} and OTII.II21R^{-/-} splenocytes were stimulated with OVA₃₂₃₋₃₃₉ peptide for 72 h in the absence of additional cytokine (open bars) or in the presence of rmlL-27 (black bars) or rmlL-21 (gray bars). 7AAD^{neg} $CD4^+$ cells were assessed by flow cytometry and multiplied by total cells counts to give viable $CD4^+$ cells per ml of culture medium. The mean of three replicate samples are shown, and these data are representative of two independent experiments. Error bars indicate SEM. *, $P < 0.05$, (unpaired Student's t test).

Table S1. Primers and probes for real-time RT-PCR

Gene	Primers	Probe
<i>mIi21</i>	5'-CTCCCGTGTGAGGGATT-3'; 5'-TCACAGTTGGGCAATAAGATG-3'	5'-AGCCACAGCTTGAGAAGCACCAGA-3'
<i>mIi27p28</i>	5'-TCAGGTGTCATCCAAGTGT-3'; 5'-GACAAGCTCCAGGGAGTGA-3'	5'-GGTAGGTATAGAGCAGCTGGGGCCAG-3'
<i>mEbi3</i>	5'-GGCCTGCTGAGTCTGAATA-3'; 5'-AGTCAAGTGAATTATCCAGTGCTT-3'	5'-CTTCCATGTACTGGGCTGCTCCG-3'
<i>mRpl19</i>	5'-ATCCGCAAGCCTGTGACTGT-3'; 5'-TCGGGCCAGGGTGTGTTT-3'	5'-TTCCCGGGCTCGTTGCCG-3'
<i>mBcl6</i>	Inventoried Taqman assay Mm00477633_m1	
<i>hGAPDH</i>	5'-CTCTGCTCCTCTGTTTCGAC-3'; 5'-ACGACCAAATCCGTTGACTC-3'	Roche UPL probe #60
<i>hIL12RB1</i>	5'-CGGCTGACCCTGAAAGAG-3'; 5'-CAGCCCTTGACAGCCTTC-3'	Roche UPL probe #78
<i>hIL12RB2</i>	5'-TCCAGATCCAGCAAATAGCA-3'; 5'-GTCCAAGGGCAGCTGTGT-3'	Roche UPL probe #82