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2. Target Validation

a) Validation table

Prolactin receptor (Prlr)	Hyperprolactinemia.	1	(1-5)
	Breast Cancer	2	(6-11)
	Prostate cancer	2	(12-17)
	Telogen effluvium and hirsutism	3	(18-21)

	Female fertility	4	(22, 23)
	Metabolism and growth	4	(10, 24-26)
	Osteoporosis	4	(27)
Male fertility	4	(17)	

(b) Implicated Diseases

The prolactin receptor (Prlr) has been implicated in a myriad of human diseases (28). The evidence for this involvement is variable in strength and so the approach taken here is to concentrate on those disorders for which strong evidence has been obtained using transgenic and knockout mouse models. A caveat here is that modulation of prolactin (Prl) or Prlr levels produces systemic endocrine disruption and so we have confined our detailed examination to those examples where systemic endocrine disruption has been demonstrated to play no part in the observed phenotype, indicated by a validation level of 2 and 3. Where endocrine disruption in transgenic and knockout animals has not been excluded as a contributing factor we have used a level of 4. In the case of fertility, metabolism and growth, endocrine regulation by Prl is central to its effects and so at least a portion of the consequent endocrine disruption in knockout models represents a valid and direct action of Prl. We have not detailed the many examples where non-genetic animal and cell-based

models reveal biology that may impinge on a specific disease process and the reader is referred to detailed reviews of these areas (28-30).

Hyperprolactinemia

Hyperprolactinemia is a common endocrine disease causing reproductive defects such as amenorrhea, infertility, or galactorrhea/gynecomastia and impotence. Osteoporosis is also observed (5). Hyperprolactinemia may result from the pharmacological use of dopamine antagonist neuroleptics, from pathologies that impinge on the dopamine control of Prl secretion, and most frequently from adenomas of the Prl-secreting pituitary lactotrophs. In the latter case large tumours may cause secondary effects such as headache and visual disturbance. Hyperprolactinemia is currently the only indication for which ablation of prolactin action is used in the clinic. Treatment by dopamine agonism using improved analogues of bromocriptine such as pergolide, cabergoline or quinagolide is usually successful in reducing Prl levels, shrinking and controlling tumour mass and restoring fertility. Between 10-20% of patients exhibit resistance to dopamine agonists (4). These patients are treated by trans-sphenoidal surgery or radiotherapy only when infertility is a problem or visual disturbance occurs. This group may benefit from the future development of drugs that modulate signalling by the Prlr, in an analogous way to blockade of the growth hormone receptor in acromegalia using Pegvisomant. In light of the growing body of evidence regarding the potential carcinogenic (see below) and osteoporotic effects of elevated Prl it may no longer be wise to allow hyperprolactinemia to go undetected and untreated. The actual rate of elevated and hyperprolactinemia in the population is unknown.

Breast Cancer

The very large and prospective Nurses Health Studies I and II (<u>http://www.channing.harvard.edu/nhs/</u>) demonstrated that top quartile serum Prl (PRL) conferred a higher relative risk (2.03 fold 95% CI 1.24-3.31 p=0.01) of developing breast cancer compared to women with bottom quartile serum Prl (8), with ER+ PR+ tumours having an increased relative risk of 1.78 (95% CI, 1.28-2.50; P-trend < 0.001) compared to ER-tumours (0.76 95% CI, 0.43-1.32; P-trend=0.28) (31). Clinical studies using bromocriptine in advanced breast cancer showed no effect (32,

33). These observations have led to two hypotheses regarding Prl action for which experimental support has been provided. Firstly advanced breast cancer may have lost sensitivity to Prl, suggesting that Prl may act during the early stages of carcinogenesis (10). Secondly the mammary epithelial cell produces Prl via a second promoter not sensitive to dopamine, allowing for autocrine or paracrine stimulation of cancer cell proliferation to continue in the presence of low serum Prl (34).

Prolactin is a complete carcinogen in rodent models of breast cancer (35). More recent transgenic strategies provide further proof and insight. Models that increase serum Prl levels result in mammary cancer. For example, transgenic mice that over-express human growth hormone (hGH), which binds both Prlr and growth hormone receptors, develop mammary carcinoma while mice that over-express the growth hormone receptor-restricted ligand bovine GH do not (36). Over-expression of rat Prl using the lipocalin promoter to drive expression predominantly in mammary epithelium produces oestrogen receptor positive tumours at a higher rate than other mouse mammary cancer models (11). Prl knockout mice expressing the polyoma middle-T antigen oncogene develop tumours significantly later (37). Using the SV40 T transgene a similar delay in tumour onset is seen in Prlr knockout animals (10) via increased cell proliferation in very early hyperplastic lesions, which was lost as they progressed to invasive carcinoma. Here the effect of Prl was demonstrated to occur via the mammary epithelial cell Prlr and not via systemic effects on other hormones like oestrogen. This result indicates that anti Prl or Prlr therapies may be most beneficial in preventing the progression of preneoplastic lesions to carcinoma. Advances in imaging technology have led to these pre-cancerous lesions being more frequently detected via micro calcification, usually at multifocal sites and so presenting a treatment dilemma; watchful waiting for evidence of progression to carcinoma, with attendant risk of metastasis, or prophylactic radical mastectomy.

Prostate Cancer

Prolactin is trophic for the normal prostate, where it operates with androgens. Greater castration-induced prostate regression occurs with additional hypophysectomy (38), androgen replacement after castration restores prostate weight less effectively with hypophysectomy (39, 40), and pituitary grafts reduce the rate and extent of prostate regression induced by castration (41) via androgen-independent mechanisms (42). Prl is capable of producing stromal hyperplasia and intraepithelial dysplastic features with long exposure (12, 13). Prl can also operate directly; Prl and the Prlr are expressed in human and rodent prostate (14) by the epithelial cells with a weak signal in the fibro-muscular stroma (43). *In vitro* Prl is mitogenic for cultured prostate epithelial cells (44) and in organ cultures normal morphology was best maintained by the addition of androgen and Prl (45). Expression in the prostate of both Prl and the Prlr is increased by androgen treatment *in vivo*, while Prlr level is also increased by Prl (15, 46). Prl has been viewed as an autocrine/paracrine growth factor (14, 15), or a survival factor (16) for prostate epithelial cells. Transgenic expression of Prl in the mouse prostate results in massive enlargement, stromal hyperplasia, ductal dilation and focal epithelial dysplasia (13). Knockout of the Prlr in animals expressing the SV40T oncogene in the prostate caused a reduction in intraepithelial neoplasia and prevented tumour formation (17).

Clinical investigation has shown that serum Prl levels in men with prostate disease are generally not different from age-matched controls when measured at presentation (47) or up to 13 years prior to diagnosis (48). During oestrogen, anti-oestrogen, anti-androgen or GnRH analogue therapy for prostate cancer, Prl levels can increase and are predictive of poor prognosis (49). This has lead to 11 reported small uncontrolled clinical trials of adjuvant bromocriptine during antiandrogen therapy; some of which report increased response rates when bromocriptine is used (50), despite the enrolment of late stage patients where tumour Prlr levels can be diminished or lost (51), and where the potential autocrine/paracrine effects of Prl produced by the prostate epithelium may operate. The use of a molecular mimic of phosphorylated Prl, S197D, successfully inhibited tumour initiation and the growth of human DU145-derived tumours in nude mice (52). Thus Prl may play a role in escape from androgen-dependence and anti Prl therapy may prove useful as an adjuvant to anti androgen therapy.

Telogen effluvium and hirsutism

The hair follicle cycle is comprised of a period of active hair shaft elongation followed by involution of the follicle and a quiescent state during which the hair shaft is retained by the follicle. Re entry of the follicle into the active state results in the production of a new hair, which in many species including humans ejects the old hair. This process is hormonally regulated, by systemic and local factors that form a zone of follicle reactivation that moves across the skin. The hair follicle expresses the Prlr in a cycle dependent manner and is only sensitive to Prl at specific cycle points (18-21). In Prlr knockout mice the timing of the first cycle of follicle reactivation- the first moult, occurs at a younger age, the resulting hair is longer and coarser, and the sexual dimorphism in these characteristics is lost (20). Importantly when Prlr knockout hair follicles and skin are transplanted to normal host animals the same effects occur, demonstrating direct Prl action on the hair follicle and discounting a systemic hormonal disruption as the cause (18). The human hair follicle is sensitive to Prl, opening the opportunity to pharmacologically regulate this pathway in cases of telogen effluvium and hirsutism.

Endocrine disruption

Modulation of Prl alters systemic hormone levels. In mice hyperprolactinemia causes increased androgen levels (12). Null mutation of Prl or Prlr in mice (23, 53) causes reduced levels of oestrogen, progesterone, and PTHRP (54). This central regulatory role of Prl produces a caveat for the remaining disease indications in Table 1; some of the effects may be directly attributable to Prlr and some may be due to the altered levels of other hormones. For this reason they are listed at level 4 and are not discussed further.

Target	Model	Disease	Refere nce	Source
Prlr	Prlr knockout mice	Hyperprolactinemia Breast Cancer Prostate Cancer Hair disorders	(23)	Jackson Laboratory 600 Main Street Bar Harbor, Maine 04609 USA Paul Kelly Centre de Recherche Croissance et Signalisation Inserm U845 Faculté de Médecine Paris Descartes - Site Necker 156 rue de

C. Disease Models.

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Prl	Prl knockout	Hyperprolactinemia Breast Cancer Prostate Cancer Hair disorders	(53)	Nelson Horseman. Department of Molecular and Cell Physiology University of Cincinnati Ohio USA
Prl	Prl transgenic Prostate specific	Prostate cancer	(12, 55)	Jan Tornell <u>Sektionen för fysiologi</u> , Medicinaregatan 11 Endokrin Box 434 405 30 GÖTEBORG SWEDEN
Prl	Prl transgenic Mammary specific	Mammary cancer	(11)	Linda Schuller Department: Comparative Biosciences, School of Veterinary Medicine 4354B Veterinary Medicine Bldg, 2015 Linden Dr., Madison, WI USA
Prl	Prl transgenic Whole body expression	Hyperprolactinemia	(12)	Jan Tornell <u>Sektionen för Fysiologi</u> , Medicinaregatan 11 Endokrin Box 434 405 30 GÖTEBORG SWEDEN
Prl	Pituitary transplant	Hyperprolactinemia		

Prlr S2	Transgenic	(56-58)	Paul Kelly
	expressing a		
	short form of		
	the Prlr		

The Prlr (23) and Prl (53) knockout models have defined the actions of Prl and are the major experimental models available. Both are constitutive in all tissues from birth and so suffer from the usual range of caveats that apply to this approach. There are no knockout models using a floxed approach to provide an inducible knockout.

The prostate (55) and mammary specific (11) transgenic models of forced Prl expression produce dysplasia and tumours respectively. The mammary model is the only mouse model that produces high frequency of oestrogen receptor positive cancer. The prostate model has not been combined with an oncogene to determine if the kinetics of carcinogenesis are altered- the most likely outcome would be decreased latency to tumourigenesis due to local prostate expression of Prl.

The Prl whole body transgenic (12) resulted from an attempt to create an inducible model of Prl expression using the metallothionine promoter. No inducible lines were found but a line exhibiting constitutive Prl over-expression was derived. This provides a model of hyperprolactinemia. Hyperprolactinemia can be easily produced experimentally by pituitary transplantation to the kidney capsule or mammary fat pad, which removes hypothalamic dopamine inhibition of pituitary Prl secretion in the transplant. Transgenic over expression of the Prlr short forms, both in wild type and a Prlr knockout background provides insight regarding the relative activity of these isoforms (56-58).

3. Drugs and Biotherapeutics.

a. Current Status

All current artificial modulators of the Prlr are based on amino acid substitutions using the natural ligands for the Prlr. The natural ligands of the Prlr in humans and other primates are GH, Prl, and during pregnancy placental lactogen. In non-Primates GH does not bind to the Prlr. A single ligand molecule dimerises two Prlr molecules, similar to other cytokine receptors, but the mechanism is not fully defined. Whether the receptor is pre-dimerised, or dimerised by ligand binding, and how this alters receptor and ligand conformation, is contentious (29).

The prototypic inhibitor of the Prlr was discovered and patented (US00418561 1989) by John Kopchick and colleagues, who replaced bovine growth hormone Gly120 with large residues such as arginine (G120R) or lysine (G120K) to produce an antagonist of the growth hormone receptor due to disruption of interaction between ligand Gly120 and receptor Trp104. Genentech followed with a similar approach using human growth hormone (US6800740 1995) that was patented (US6429186 1994) for use in breast cancer (59) and for antagonism of the Prlr by the co-administration of zinc (WO05058232). A further 8 mutations and pegylation produced the drug Pegvisomant (Somavert®), used for the treatment of human growth hormone excess (60). Pegvisomant, due to the extra 8 mutations, does not antagonise or agonise the Prlr and so there is currently no drug approved for human use that acts as a Prlr antagonist.

The homologous Prl mutant to hGH-G120R is hPrl-G129R and it is unpatented due to early poor results (61). The ability of these mutant ligands to antagonise the Prlr has been extensively characterised and mixed antagonism and agonism has been described. The mutation causes a 10-fold reduction in affinity, so a complicating feature is the large concentrations required to overcome endogenous Prl levels. The agonistic activity of hPrl-G129R can be removed by deletion of 9 N-terminal amino acids, a strategy developed by comparison to ovine placental lactogen and patented (WO03057729) by Vincent Goffin, Paul Kelly and colleagues. The problem of lower affinity remains but a more pure antagonism is produced (29, 62-64).

Brooks and colleauges have taken a different approach, patenting (US6995244 2003) Prl mutations that do not interfere with ligand-receptor binding, but instead interfere with the hypothesised conformational change in Prl that may be induced by initial then subsequent ligand binding to two receptor molecules (61).

Walker and colleagues have made molecular mimics of phosphorylated Prl, hPrl-S179D (US6890738 2003), based on observed antagonism of Prl action by phosphoprolactin (65). Controversy has raged between Goffin and colleagues, and Walker and colleagues regarding the mechanism of S179D action. The argument has turned on whether S179D is an agonist or antagonist of Prl action, complicated by assay details such as the ratio of background unphosphorylated Prl, tissue and species of origin, relative concentrations, hormone preparation etc. Generally S179D antagonises the biological effects of Prl in whole animal experiments, but this is not always reflected at the molecular level, where S179D exerts mixed agonism and antagonism in receptor interaction and activation assays. It now appears that S179D generates a qualitatively different signal from the Prlr compared to other ligands and that this response is context specific (65). Possible mechanisms include variable final receptor conformations causing differential activation of associated signalling pathways. Thus S179D, and indeed perhaps all mutants of Prl and GH, may be considered as context specific receptor activity modulators (SPRMs) analogous to the Specific Oestrogen Receptor Modulators (SERMs) such as Tamoxifen and Raloxifen (65). So the resolution of the SPRM controversy may also take a similar path to that which surrounded the SERMs.

Ligands and antibodies

Reagent	Patent	Group	Company	Reference
hGH-G120R	US6800740 1995	Fuh and Wells	Genentec	(66)
+Zn	US6429186 1994	Clark et al		(61)
bGH-120R	US00418561 1989	Kopchick et al.	Ohio University	(60)
hPrl-G129R	No	Goffin et al	INSERM U808	(64)

Pegvisomant	US00418561	http://www.somave	Pfizer	(60)
	1989 and	rt.com/		
Somavert®	others.			
D1-9hPrl-		Goffin et al	INSERM U808	(64)
G129R				
hPrl-S179D	US6890738	Walker et al	UC Riverside	(65)
	2003			
hPrl mutant	US6995244	Brooks et al		(64)
	2003			
Anti Prlr			AbCam	Catalogue
antibody				number
SPM213				Ab17831
Anti Prlr		Clevenger et al.	Zymed	Catalogue
antibody				number 35-
1A2B1				9200
Anti Prlr		Kelly et al.	AbCam	Catalogue
antibody-U5				number
				Ab2772
Prl ovine and			Sigma	
human				
Prl		http://www.humc.e	NIDDK	
		du/hormones/		

b. Next frontiers.

Currently available Prlr modulators produce either no detectable signal from the receptor (Δ 1-9-hPrl-G129R), or a modified signal (hPrl-S179D). Δ 1-9-hPrl-G129R and hPrl-G229R display a 10 fold loss of affinity and all molecules have a very short serum half life of 15-30 minutes. Use of pegylation to increase the half life of Pegvisomant to 70 hours caused reduced potency, and so it is likely that this strategy would further decrease the potency of any Prlr modulators based on mutation of the natural ligand. Thus although approaches to date have made great progress in identifying key ligand mutations, it is clear that they need be developed further to

become therapeutics. Especially pressing is the restoration of binding affinity, which ideally should be an order of magnitude higher than endogenous ligands. Low affinity modulators can overcome endogenous ligands by 10-100 fold concentration excess, and if the modulator elicits no pathogenic signal from the receptor, or no signal at all, this may represent a feasible approach. Alternative strategies include the generation of antibodies that block ligand activation of the Prlr, a strategy that has proved successful for a number of cell surface receptors. More distant is the development of small molecules capable of interfering with ligand binding, with receptor dimerisation, or with the associated signalling machinery. Defining and then targeting effectors down-stream of the Prlr provides a third approach that may offer the added advantage of selective ablation or stimulus of a specific Prl action, while maintaining others.

4. Function and localisation

a. In homeostasis.

There are a number of different protein isoforms of the Prlr that are produced via alternative splicing and post translational processing. These isoforms contain various deletions of the motifs responsible for ligand binding, membrane localisation and association with members of the signalling cascade such as Jak2 or MapK (Figure 1). Multiple receptor isoforms provide the potential for complex modulation of signalling via homodimer competition for ligand, and via heterodimerisation of the various isoforms and differentially associated signalling molecules. In Primates this may extend to the GHR via ligand-induced PRLR/GHR heterodimerisation (67). The major outputs include the Jak-Stat, MapK, Akt and Rac pathways to modulate cell proliferation, differentiation motility and survival (6, 28, 29).

b. In Disease

How the various signalling events that can be generated by the Prlr relate to disease remains to be defined.

5. Characteristic structural features.

Figure 1 shows the structural features and conserved sequence motifs of the Prlr relative to the mouse and human gene structure. The structural features are organised into discrete exons, reflecting the molecular evolution of the cytokine receptor family (68). Receptor diversity is generated via alternate exon splicing to produce receptors with different intracellular motifs, and thus interactions with the Jak/Stat pathway, the MapK pathway via Fak and the Akt pathway via Src (6, 29). The major signalling dichotomy exists between the short form that activates MapK alone and the long form that activates all three pathways. The Prlr has not been crystallised but the homologous GHR provides a model (60), and Prl has been the subject of intense structural study (30). The receptors may exist as pre-dimerised molecules or solo molecules that sequentially bind site 1 and then site 2 of the ligand, and so undergo a conformational change that results in activation via the close apposition of associated kinases (61).

Modulation of the activity of the Prlr has focussed primarily on the ligand binding site and in particular on mutation of site 2 of Prl, as binding here is thought to be the key step in receptor activation. Specific modulation of the signalling response of the receptor may be possible via this means, as indicated by the effects of S179D. A high affinity receptor-blocking pure-antagonist or a binding site blocking monoclonal antibody could be used to promote complete ablation of signalling. Other approaches to modulating the Prlr include generating small molecules molecules that can specifically interrupt the association of one or more signalling molecules with the receptor or that prevent the formation of receptor dimers, presumed to involve the membrane proximal regions. Elucidation and targeting of the down stream effectors of the transcriptional and other actions of the Prlr may also provide a future therapeutic strategy.

In summary the Prlr mediates the detrimental effects of hyperprolactinemia, which is treated in the clinic using dopamine agonists with high but not complete success. Raised prolactin levels play a role in the aetiology of breast and possibly prostate cancer. The first action of prolactin in these cancers may be to hasten progression of preneoplastic lesions to carcinoma, via effects on cell proliferation. Prototype Prlr modulators have been developed and so anti prolactin receptor therapy may find a place as a preventative measure, especially as imaging technology improves the

discovery of these early lesions. A subset of cancers may retain sensitivity to prolactin and so anti prolactin receptor therapy may also have a role as an adjuvant to other anti-hormonal agents.

6. References.

1. Colao A, Di Sarno A, Guerra E, De Leo M, Mentone A, Lombardi G. Drug insight: Cabergoline and bromocriptine in the treatment of hyperprolactinemia in men and women. Nature Clinical Practice Endocrinology & Metabolism 2006;2(4):200-10.

2. Colao A, Vitale G, Di Sarno A, *et al.* Prolactin and prostate hypertrophy: a pilot observational, prospective, case-control study in men with prolactinoma. Journal of Clinical Endocrinology & Metabolism 2004;89(6):2770-5.

3. Crosignani PG. Current treatment issues in female hyperprolactinaemia. European Journal of Obstetrics, Gynecology, & Reproductive Biology 2006;125(2):152-64.

4. Molitch ME. Dopamine resistance of prolactinomas. Pituitary 2003;6(1):19-27.

5. Molitch ME. Prolactin-secreting tumors: what's new? Expert Review of Anticancer Therapy 2006;6 Suppl 9:S29-35.

6. Clevenger CV, Furth PA, Hankinson SE, Schuler LA. The role of prolactin in mammary carcinoma. Endocrine Reviews 2003;24(1):1-27.

7. Hankinson SE, Willett WC, Manson JE, *et al.* Plasma sex steroid hormone levels and risk of breast cancer in postmenopausal women. Journal of the National Cancer Institute 1998;90(17):1292-9.

8. Hankinson SE, Willett WC, Michaud DS, *et al.* Plasma prolactin levels and subsequent risk of breast cancer in postmenopausal women. Journal of the National Cancer Institute 1999;91(7):629-34.

9. Nouhi Z, Chughtai N, Hartley S, Cocolakis E, Lebrun JJ, Ali S. Defining the role of prolactin as an invasion suppressor hormone in breast cancer cells. Cancer Research 2006;66(3):1824-32.

10. Oakes SR, Robertson FG, Kench JG, *et al.* Loss of mammary epithelial prolactin receptor delays tumor formation by reducing cell proliferation in low-grade preinvasive lesions. Oncogene 2007;26(4):543-53.

11. Rose-Hellekant TA, Arendt LM, Schroeder MD, Gilchrist K, Sandgren EP, Schuler LA. Prolactin induces ERalpha-positive and ERalpha-negative mammary cancer in transgenic mice. Oncogene 2003;22(30):4664-74.

12. Wennbo H, Kindblom J, Isaksson OG, Tornell J. Transgenic mice overexpressing the prolactin gene develop dramatic enlargement of the prostate gland. Endocrinology 1997;138(10):4410-5.

13. Kindblom J, Dillner K, Robertson F, Ormandy C, Tornell J, Wennbo H. Prostate hyperplasia in a transgenic mouse with prostate-sprecific expression of prolactin. Endocrinology 2003;144:2269-78.

14. Nevalainen MT, Valve EM, Ingleton PM, Nurmi M, Martikainen PM, Harkonen PL. Prolactin and prolactin receptors are expressed and functioning in human prostate. J Clin Invest 1997;99(4):618-27.

15. Nevalainen MT, Valve EM, Ahonen T, Yagi A, Paranko J, Harkonen PL. Androgen-dependent expression of prolactin in rat prostate epithelium in vivo and in organ culture. FASEB J 1997;11(14):1297-307.

16. Ahonen TJ, Harkonen PL, Laine J, Rui H, Martikainen PM, Nevalainen MT. Prolactin is a survival factor for androgen-deprived rat dorsal and lateral prostate epithelium in organ culture. Endocrinology 1999;140(11):5412-21.

17. Robertson FG, Harris J, Naylor MJ, *et al.* Prostate development and carcinogenesis in prolactin receptor knockout mice. Endocrinology 2003;144(7):3196-205.

18. Craven AJ, Nixon AJ, Ashby MG, *et al.* Prolactin delays hair regrowth in mice. Journal of Endocrinology 2006;191(2):415-25.

19. Nixon AJ, Ford CA, Wildermoth JE, Craven AJ, Ashby MG, Pearson AJ. Regulation of prolactin receptor expression in ovine skin in relation to circulating prolactin and wool follicle growth status. Journal of Endocrinology 2002;172(3):605-14.

20. Craven AJ, Ormandy CJ, Robertson FG, *et al.* Prolactin signaling influences the timing mechanism of the hair follicle: analysis of hair growth cycles in prolactin receptor knockout mice. Endocrinology 2001;142(6):2533-9.

21. Pearson AJ, Ashby MG, Wildermoth JE, Craven AJ, Nixon AJ. Effect of exogenous prolactin on the hair growth cycle. Experimental Dermatology 1999;8(4):358-60.

22. Binart N, Helloco C, Ormandy CJ, *et al.* Rescue of preimplantatory egg development and embryo implantation in prolactin receptor-deficient mice after progesterone administration. Endocrinology 2000;141(7):2691-7.

23. Ormandy CJ, Camus A, Barra J, *et al.* Null mutation of the prolactin receptor gene produces multiple reproductive defects in the mouse. Genes Dev 1997;11:167-78.

24. Freemark M, Fleenor D, Driscoll P, Binart N, Kelly P. Body weight and fat deposition in prolactin receptor-deficient mice. Endocrinology 2001;142(2):532-7.

25. Nilsson L, Binart N, Bohlooly YM, *et al.* Prolactin and growth hormone regulate adiponectin secretion and receptor expression in adipose tissue. Biochemical & Biophysical Research Communications 2005;331(4):1120-6.

26. Fleenor D, Oden J, Kelly PA, *et al.* Roles of the lactogens and somatogens in perinatal and postnatal metabolism and growth: studies of a novel mouse model combining lactogen resistance and growth hormone deficiency. Endocrinology 2005;146(1):103-12.

27. Clement-Lacroix P, Ormandy CJ, Lepescheux L, *et al.* Osteoblasts are a new target for prolactin: analysis of bone formation in prolactin receptor knockout mice. Endocrinology 1999;140:96-105.

28. Bole-Feysot C, Goffin V, Edery M, Binart N, Kelly PA. Prolactin (PRL) and its receptor: actions, signal transduction pathways and phenotypes observed in PRL receptor knockout mice. Endocrine Reviews 1998;19(3):225-68.

29. Goffin V, Bernichtein S, Touraine P, Kelly PA. Development and potential clinical uses of human prolactin receptor antagonists. Endocrine Reviews 2005;26(3):400-22.

30. Goffin V, Shiverick KT, Kelly PA, Martial JA. Sequence-function relationships within the expanding family of prolactin, growth hormone, placental lactogen, and related proteins in mammals. Endocrine Reviews 1996;17(4):385-410.

31. Tworoger SS, Eliassen AH, Rosner B, Sluss P, Hankinson SE. Plasma prolactin concentrations and risk of postmenopausal breast cancer. Cancer Research 2004;64(18):6814-9.

32. Anonymous. Clinical trial of 2-Br- -ergocryptine (CB154) in advanced breast cancer. European Journal of Cancer (Oxford) 1972;8(2):155-6.

33. Engelsman E, Heuson JC, Blonk Van Der Wijst J, *et al.* Controlled clinical trial of L-dopa and nafoxidine in advanced breast cancer: an E.O.R.T.C. study. British Medical Journal 1975;2(5973):714-5.

34. Manhes C, Goffin V, Kelly PA, Touraine P. Autocrine prolactin as a promotor of mammary tumour growth. Journal of Dairy Research 2005;72 Spec No:58-65.

35. Welsch CW, Nagasawa H. Prolactin and murine mammary tumorigenesis: a review. Cancer Research 1977;37(4):951-63.

36. Wennbo H, Gebre-Medhin M, Gritli-Linde A, Ohlsson C, Isaksson OG, Tornell J. Activation of the prolactin receptor but not the growth hormone receptor is important for induction of mammary tumors in transgenic mice. Journal of Clinical Investigation 1997;100(11):2744-51.

37. Vomachka AJ, Pratt SL, Lockefeer JA, Horseman ND. Prolactin genedisruption arrests mammary gland development and retards T-antigen-induced tumor growth. Oncogene 2000;19(8):1077-84.

38. Huggins C, Russell P. Endocrinology 1946;39:1-7.

39. Vanderlaan W. Observations on the hormonal control of the prostate. Lab Invest 1960;9:185-90.

40. Grayhack J. Pituitary factors influencing the growth of the prostate. Cancer Inst Monographs 1963;12:198-9.

41. Kolbusz WE, Lee C, Grayhack JT. Delay of castration induced regression in rat prostate by pituitary homografts. J Urol 1982;127(3):581-4.

42. Smith C, Assimos D, Lee C, Grayhack JT. Metabolic action of prolactin in regressing prostate: independent of androgen action. Prostate 1985;6(1):49-59.

43. Ouhtit A, Morel G, Kelly PA. Visualization of gene expression of short and long forms of prolactin receptor in rat reproductive tissues. Biol Reprod 1993;49(3):528-36.

44. McKeehan WL, Adams PS, Rosser MP. Direct mitogenic effects of insulin, epidermal growth factor, glucocorticoid, cholera toxin, unknown pituitary factors and possibly prolactin, but not androgen, on normal rat prostate epithelial cells in serum-free, primary cell culture. Cancer Res 1984;44(5):1998-2010.

45. Nevalainen MT, Valve EM, Makela SI, Blauer M, Tuohimaa PJ, Harkonen PL. Estrogen and prolactin regulation of rat dorsal and lateral prostate in organ culture. Endocrinology 1991;129(2):612-22.

46. Nevalainen MT, Valve EM, Ingleton PM, Harkonen PL. Expression and hormone regulation of prolactin receptors in rat dorsal and lateral prostate. Endocrinology 1996;137(7):3078-88.

47. Harper ME, Wilson DW, Jensen HM, Pierrepoint CG, Griffiths K. Steroid hormone concentrations in relation to patient prognosis and prostate tumour grade. J Steroid Biochem 1987;27(1-3):521-4.

48. Eaton NE, Reeves GK, Appleby PN, Key TJ. Endogenous sex hormones and prostate cancer: a quantitative review of prospective studies. Br J Cancer 1999;80(7):930-4.

49. Matzkin H, Kaver I, Lewyshon O, Aylon D, Braf Z. The role of prolactin levels under GnRH analogue treatment in advanced prostatic carcinoma. Cancer 1988;61:2187-91.

50. Horti J, Figg WD, Weinberger B, Kohler D, Sartor O. A phase II study of bromocriptine in patients with androgen-independent prostate cancer. Oncol Rep 1998;5(4):893-6.

51. Leav I, Merk FB, Lee KF, *et al.* Prolactin receptor expression in the developing human prostate and in hyperplastic, dysplastic, and neoplastic lesions. Am J Pathol 1999;154(3):863-70.

52. Xu X, Kreye E, Kuo CB, Walker AM. A molecular mimic of phosphorylated prolactin markedly reduced tumor incidence and size when DU145 human prostate cancer cells were grown in nude mice. Cancer Research 2001;61(16):6098-104.

53. Horseman ND, Zhao W, Montecino-Rodriguez E, *et al.* Defective mammopoiesis, but normal hematopoiesis, in mice with a targeted disruption of the prolactin gene. EMBO Journal 1997;16(23):6926-35.

54. Clement-Lacroix P, Ormandy C, Lepescheux L, *et al.* Osteoblasts are a new target for prolactin: analysis of bone formation in prolactin receptor knockout mice. Endocrinology 1999;140(1):96-105.

55. Kindblom J, Dillner K, Ling C, Tornell J, Wennbo H. Progressive prostate hyperplasia in adult prolactin transgenic mice is not dependent on elevated serum androgen levels. Prostate 2002;53(1):24-33.

56. Manhes C, Kayser C, Bertheau P, *et al.* Local over-expression of prolactin in differentiating mouse mammary gland induces functional defects and benign lesions, but no carcinoma. Journal of Endocrinology 2006;190(2):271-85.

57. Binart N, Imbert-Bollore P, Baran N, Viglietta C, Kelly PA. A short form of the prolactin (PRL) receptor is able to rescue mammopoiesis in heterozygous PRL receptor mice. Molecular Endocrinology 2003;17(6):1066-74.

58. Saunier E, Dif F, Kelly PA, Edery M. Targeted expression of the dominantnegative prolactin receptor in the mammary gland of transgenic mice results in impaired lactation. Endocrinology 2003;144(6):2669-75.

59. Fuh G, Wells JA. Prolactin receptor antagonists that inhibit the growth of breast cancer cell lines. Journal of Biological Chemistry 1995;270(22):13133-7.

60. Kopchick JJ, Parkinson C, Stevens EC, Trainer PJ. Growth hormone receptor antagonists: discovery, development, and use in patients with acromegaly. Endocrine Reviews 2002;23(5):623-46.

61. Goffin V, Tallet E, Jomain J, Kelly PA. Development of prolactin receptor antagonists: same goal, different ways. Recent Patents on Endocrine, Metabolic & Immune Drug Discovery 2007;1:41-52.

62. Goffin V, Bernichtein S, Kayser C, Kelly PA. Development of new prolactin analogs acting as pure prolactin receptor antagonists. Pituitary 2003;6(2):89-95.

63. Goffin V, Kinet S, Ferrag F, Binart N, Martial JA, Kelly PA. Antagonistic properties of human prolactin analogs that show paradoxical agonistic activity in the Nb2 bioassay. Journal of Biological Chemistry 1996;271(28):16573-9.

64. Goffin V, Touraine P, Culler MD, Kelly PA. Drug Insight: prolactin-receptor antagonists, a novel approach to treatment of unresolved systemic and local hyperprolactinemia? Nature Clinical Practice Endocrinology & Metabolism 2006;2(10):571-81.

65. Walker AM. Therapeutic potential of S179D prolactin--from prostate cancer to angioproliferative disorders: the first selective prolactin receptor modulator. Expert Opinion on Investigational Drugs 2006;15(10):1257-67.

66. Fuh G, Cunningham BC, Fukunaga R, Nagata S, Goeddel DV, Wells JA. Rational design of potent antagonists to the human growth hormone receptor. Science 1992;256(5064):1677-80. 67. Biener E, Martin C, Daniel N, *et al.* Ovine placental lactogen-induced heterodimerization of ovine growth hormone and prolactin receptors in living cells is demonstrated by fluorescence resonance energy transfer microscopy and leads to prolonged phosphorylation of signal transducer and activator of transcription (STAT)1 and STAT3. Endocrinology 2003;144(8):3532-40.

68. Ormandy CJ, Binart N, Helloco C, Kelly PA. Mouse prolactin receptor gene: genomic organization reveals alternative promoter usage and generation of isoforms via alternative 3'-exon splicing. DNA & Cell Biology 1998;17(9):761-70.

7. Figures

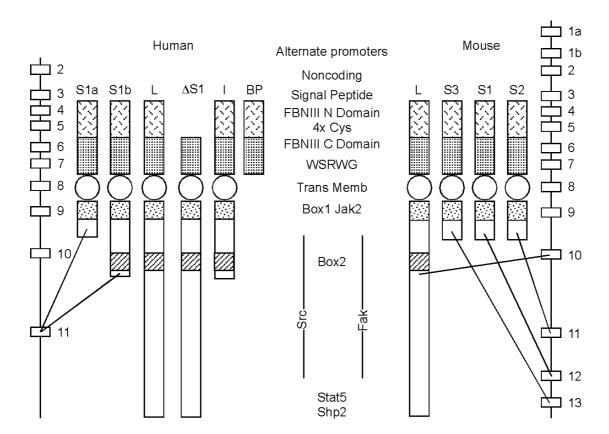


Figure 1. Prlr structural motifs and genomic organization.

Exons of the prolactin receptor gene (numbered boxes) of the human (left hand side) and mouse (right hand side) are shown beside their protein product so that the indicated structural features (variously shaded shapes and middle text) correspond horizontally with the exon(s) that encode them. Alternative splicing of exons 10-13 produces different proteins, indicated by lines, and produces multiple protein products designated as short (S) long (L), intermediate (I) or binding protein (BP). Human S1b is produced by variable splicing of exon 11 with prematurely terminated exon 10. Human long and delta S1 use exon 10 while human intermediate shows a frame shift transcription of exon 10. Human BP is produced by protease activity. Alternative splicing of mouse exons 10-13 produces the four Prlr isoforms shown. Tissue specific alternate promoter usage is common.